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September 8, 2000

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UTILITY PATENT APPLICATION TRANSMITTAL

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Title: Amino Polyol Amine Oxidase Polynucleotides and Related Polypeptides and
Methods of Use

APPLICATION ELEMENTS

1. ☐ Fee Transmittal Form (Submit an original, and a duplicate for fee processing)
2. ☒ Specification [Total Pages 83]
(Preferred arrangement set forth below)
 - Descriptive title of the Invention
 - Cross Reference to Related Applications
 - Background of the Invention
 - Brief Summary of the Invention
 - Brief Description of the Drawings (if filed)
 - Detailed Description
 - Claim(s)
 - Abstract of the Disclosure
3. ☒ Drawing(s) (35 USC 113) [Total Sheets 2]
 - a. ☐ Formal
 - b. ☒ Informal

09/08/00



4. ☐ Oath or Declaration [Total Pages ____]
- a. ☐ Newly executed (original or copy)
- b. ☐ Copy from a prior application (37 CFR 1.63(d))
(for continuation/divisional with Box 16 completed)
- i. ☐ DELETION OF INVENTOR(S)
Signed statement attached deleting inventor(s) named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b).
5. ☐ Microfiche Computer Program (Appendix)
6. ☒ Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)
- a. ☒ Computer Readable Copy
- b. ☒ Paper Copy (identical to computer copy) [Total Pages 87]
- c. ☒ Statement verifying identity to above copies

ACCOMPANYING APPLICATION PARTS

7. ☐ Assignment Papers (cover sheet & document(s))
8. ☐ 37 CFR 3.73(b) Statement ☐ Power of Attorney
(where there is an assignee)
9. ☐ English Translation Document (if applicable)
10. ☒ Information Disclosure Statement (IDS/PTO-1449) ☐ Copies of IDS Citations
11. ☐ Preliminary Amendment
12. ☒ Return Receipt Postcard (MPEP 503) (Should be specifically itemized)
13. ☐ Small Entity Statement(s) ☐ Statement filed in prior application
Status still proper and desired
14. ☐ Certified Copy of Priority document(s)
15. ☐ Other:

16. If a **CONTINUING APPLICATION**, check the appropriate box, and supply the requisite information below and in a preliminary amendment:

☐ Continuation ☐ Divisional ☒ Continuation-in-part (CIP) of prior application
No. 09/352,159 and 09/352,168.

Prior application information: Examiner _____ Group/Art Unit:

For CONTINUATION or DIVISIONAL APPS only: The entire disclosure of the prior application, from which an oath or declaration is supplied under Box 4b, is considered a part of the disclosure of the accompanying continuation or divisional application and is hereby incorporated by reference. The incorporation can only be relied upon when a portion has been inadvertently omitted from the submitted application parts.

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Respectfully submitted,



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AMINO POLYOL AMINE OXIDASE POLYNUCLEOTIDES AND RELATED
POLYPEPTIDES AND METHODS OF USE

5

Technical Field

The present invention relates generally to the detection and isolation of fumonisin and AP1 degrading enzymes and to compositions and methods for degradation of fumonisin, a structurally related mycotoxin, or its hydrolysis product AP1. This method has broad application in agricultural biotechnology and crop agriculture and in the improvement of food grain quality.

Cross Reference To Related Application

This application is a continuation-in-part of U.S. Application No. 09/352,159, which claims the benefit of U.S. Provisional Application No. 60/135,391, filed May 21, 1999 and U.S. Provisional Application No. 60/092,936, filed July, 15 1998 all of which are hereby incorporated by reference. This application also claims the benefit of U.S. Application No. 09/352,168, which is hereby incorporated by reference.

Background of the Invention

Fungal diseases are common problems in crop agriculture. Many strides have been made against plant diseases as exemplified by the use of hybrid plants, pesticides and improved agricultural practices. However, as any grower or home gardener can attest, the problems of fungal plant disease continue to cause difficulties in plant cultivation. Thus, there is a continuing need for new methods and materials for solving the problems caused by fungal diseases of plants.

These problems can be met through a variety of approaches. For example, the infectious organisms can be controlled through the use of agents that are selectively biocidal for the pathogens. Another method is interference with the mechanism by which the pathogen invades the host crop plant. Yet another method, in the case of pathogens that cause crop losses, is interference with the mechanism by which the pathogen causes

injury to the host crop plant. Still another method, in the case of pathogens that produce toxins that are undesirable to mammals or other animals that feed on the crop plants, is interference with toxin production, storage, or activity. This invention falls into the latter two categories.

5 Since their discovery and structural elucidation in 1988 (Bezuidenhout *et al.*,
Journal Chem Soc, Chem Commun 1988: 743-745 (1988)), fumonisins have been
recognized as a potentially serious problem in maize-fed livestock. They are linked to
several animal toxicoses including leukoencephalomalacia (Marasas, *et al.*, Onderstepoort
10 Journal of Veterinary Research 55: 197-204 (1988); Wilson, *et al.*, American Association
of Veterinary Laboratory Diagnosticians: Abstracts 33rd Annual Meeting, Denver,
Colorado, October 7-9, 1990, Madison, Wisconsin, USA) and porcine pulmonary edema
(Colvin, *et al.*, Mycopathologia 117: 79-82 (1992)). Fumonisins are also suspected
carcinogens (Geary W (1971) Coord Chem Rev 7: 81; Gelderblom, *et al.*, Carcinogenesis
12: 1247-1251 (1991); Gelderblom, *et al.*, Carcinogenesis 13: 433-437 (1992)). *Fusarium*
15 isolates in section *Liseola* produce fumonisins in culture at levels from 2 to >4000 ppm
(Leslie, *et al.*, Phytopathology 82: 341-345 (1992)). Isolates from maize (predominantly
mating population A) are among the highest producers of fumonisin. (Leslie *et al.*, *supra*).
Fumoinisin levels detected in field-grown maize have fluctuated widely depending on
location and growing season, but both preharvest and postharvest surveys of field maize
20 have indicated that the potential for high levels of fumonisins exists (Murphy, *et al.*, J Agr
Food Chem 41: 263-266 (1993)). Surveys of food and feed products have also detected
fumoinisin (Holcomb, *et al.*, J Agr Food Chem 41: 764-767 (1993); Hopmans, *et al.*, J Agr
Food Chem 41: 1655-1658 (1993); Sydenham, *et al.*, J Agr Food Chem 39: 2014-2018
(1991)). The etiology of *Fusarium* ear mold is poorly understood, although physical
25 damage to the ear and certain environmental conditions can contribute to its occurrence
(Nelson, Mycopathologia 117: 29-36 (1992)). *Fusarium* can be isolated from most field
grown maize, even when no visible mold is present. The relationship between seedling
infection and stalk and ear diseases caused by *Fusarium* is not clear. Genetic resistance to
visible kernel mold has been identified (Gendloff, *et al.*, Phytopathology 76: 684-688
30 (1986); Holley, *et al.*, Plant Dis 73: 578-580 (1989)), but the relationship to visible mold
to fumoinisin production has yet to be elucidated.

Fumonisin have been shown in *in vitro* mammalian cell studies to inhibit sphingolipid biosynthesis through inhibition of the enzyme sphingosine N-acetyl transferase, resulting in the accumulation of the precursor sphinganine (Norred, *et al.*, *Mycopathologia* 117: 73-78 (1992); Wang, *et al.*, *Biol Chem* 266: 14486 (1991); Yoo, *et al.*, *Toxicol Appl Pharmacol* 114: 9-15 (1992); Nelson, *et al.*, *Annu Rev Phytopathol* 31:233-252 (1993)). It is likely that inhibition of this pathway accounts for at least some of fumonisin's toxicity, and support for this comes from measures of sphinganine: sphingosine ratios in animals fed purified fumonisin (Wang, *et al.*, *J Nutr* 122: 1706-1716 (1992)). Fumonisin also affect plant cell growth (Abbas, *et al.*, *Weed Technol* 6: 548-552 (1992); Vanasch, *et al.*, *Phytopathology* 82: 1330-1332 (1992); Vesonder, *et al.*, *Arch Environ Contam Toxicol* 23: 464-467 (1992)). Kuti *et al.*, (Abstract, Annual Meeting American Phytopathological Society, Memphis, TN: APS Press 1993) reported on the ability of exogenously added fumonisins to accelerate disease development and increase sporulation of *Fusarium moniliforme* and *Fusarium oxysporum* on tomato.

Enzymes that degrade the fungal toxin fumonisin to its de-esterified form (e.g. AP1 from FB1) have been identified in US patent no. 5,716,820, issued February 10, 1998, US patent no. 5,792,931, issued August 11, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997, and all hereby incorporated by reference. It is understood that AP1 as used here designates the hydrolyzed form of any fumonisin, FB1, FB2, FB3, FB4, or any other AP1-like compounds, including synthetically produced AP1 like compounds, that contain a C-2 or C-1 amine group and one or more adjacent hydroxyl groups. Plants expressing a fumonisin esterase enzyme, infected by fumonisin producing fungus, and tested for fumonisin and AP1 were found to have low levels of fumonisin but high levels of AP1. AP1 is less toxic than fumonisin to plants and probably also to animals but contamination with AP1 is still a concern (Lamprecht, *et al.*, *Phytopathology*, 84:383-391 (1991)). The preferred result would be complete detoxification of fumonisin to a non-toxic form. Therefore enzymes capable of degrading AP1 are necessary for the further detoxification of fumonisin.

The present invention provides newly discovered polynucleotides and related polypeptides of amino polyol amine oxidase (abbreviated APAO, formerly known as AP1 catabolase, US patent no. 5,716,820, *supra*, US patent no. 5,792,931, *supra*; US patent no.

6,025,188, *supra*, pending US application no. 08/888,950, *supra*; trAPAO is the abbreviation for a truncated, but still functional APAO), capable of oxidatively deaminating the AP1 to a compound identified as the 2-oxo derivative of AP1 or its cyclic ketal form (abbreviated as 2-OP, formerly called AP1-N1, US patent no. 5,716,820, US patent no. 5,792,931, US patent no. 6,025,188, *supra*; pending US application no. 08/888,950, *supra*), isolated from *Exophiala spinifera*, ATCC 74269. The partially purified APAO enzyme from *Exophiala spinifera* has little or no activity on intact FB1, a form of fumonisin. However, recombinant APAO enzyme from *Exophiala spinifera*, expressed in *E. coli*, has significant but reduced activity on intact FB1 and other B-series fumonisins. APAO or trAPAO thus could potentially be used without fumonisin esterase since the amine group is the major target for detoxification. Alternatively, fumonisin esterase and APAO (or trAPAO) can be used together for degrading toxins.

APAO is a type of flavin amine oxidase (EC 1.4.3.4, enzyme class nomenclature, see *Enzyme Nomenclature 1992*, Recommendations of the Nomenclature Committee of the IUBMB on the Nomenclature and Classification of Enzymes, Academic Press, Inc. (1992)). One class of flavin amine oxidases in mammals is known as monoamine oxidases, where they participate in the conversion of amines involved in neuronal function. A prokaryotic flavin amine oxidase that deaminates putrescine has been described (Ishizuka *et al.*, *J. Gen Microbiol.* 139:425-432 (1993)). A single fungal gene, from *Aspergillus niger* has been cloned (Schilling *et al.*, *Mol Gen Genet.* 247:430-438 (1995)). It deaminates a variety of alkyl and aryl amines, but when tested for its ability to oxidize AP1, was found to not contain AP1 oxidizing activity.

The toxicity of fumonisins and their potential widespread occurrence in food and feed makes it imperative to find detoxification or elimination strategies to remove the compound from the food chain.

Summary of the Invention

The present invention provides polynucleotides and related polypeptides of newly discovered APAOs. SEQ ID NO: 5 contains the nucleotide sequence of an active, truncated APAO (trAPAO), SEQ ID NO: 10 contains the nucleotide sequence of trAPAO with an additional lysine and SEQ ID NO: 22, 35, 37, 39, 41, 43, and 45 comprise full length nucleotide sequences of APAOs isolated from different organisms. In addition,

APAO can be modified to eliminate glycosylation sites and/or cysteine residues, for example, see SEQ ID NOS: 32, 48, 50, and 52. Another aspect of the present invention is the method of predicting possible mutagenesis sites on APAO by developing a 3-dimensional model of APAO and then identifying the possible sites that may contribute to misfolding of the protein. The present invention also includes the 3-dimensional model of APAO generated by a computer modeling program, preferably the *Modeler* program. For expression in a plant, the polynucleotide of the present invention can be operably linked to a targeting sequence. It is an object of the present invention to provide transgenic plants comprising the nucleic acids of the present invention.

Therefore, in one aspect, the present invention relates to an isolated APAO encoding polynucleotide ligated to a fumonisin esterase encoding polynucleotide wherein the APAO encoding polynucleotide comprises a member selected from (a) a polynucleotide encoding a polypeptide of the present invention; (b) a polynucleotide having at least 70% sequence identity to the polynucleotides of the present invention; and (c) a polynucleotide of the present invention. The isolated nucleic acid can be DNA. The isolated nucleic acid can also be RNA. Examples of fumonisin esterase genes include, but are not limited to ESP1 and BEST1.

In another aspect, the present invention relates to vectors comprising the polynucleotides of the present invention, including ligated and non-ligated polynucleotides. Also the present invention relates to recombinant expression cassettes, comprising a polynucleotide of the present invention operably linked to a promoter.

In another aspect, the present invention is directed to a host cell into which has been introduced the polynucleotides of the present invention, including a host cell comprising a fumonisin esterase ligated to an APAO or a fumonisin esterase not ligated to an APAO.

In yet another aspect, the present invention relates to a transgenic plant or plant cell comprising a recombinant expression cassette with a promoter operably linked to any of the isolated polynucleotides of the present invention. Preferred plants containing the recombinant expression cassette of the present invention include but are not limited to maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley, tomato, and millet. The present invention also provides transgenic seed from the transgenic plant.

In another aspect, the present invention relates to an isolated protein selected from the group consisting of (a) a polypeptide comprising at least 70% sequence identity to a polypeptide of the present invention; (b) a polypeptide encoded by a nucleic acid of the present invention; and (c) a polypeptide characterized by a polypeptide of the present invention.

This invention further provides methods of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a structurally related mycotoxin breakdown product, by applying APAO as a spray or wash. Additionally, fumonisins and related mycotoxins can be degraded by the application of both fumonisin esterase enzymes and APAO enzymes. Mycotoxins can be degraded in harvested grain, during the processing of harvested grain, in animal feed, or in plant tissue as, for example, during the use of the plant for silage or as a spray on grain, fruit or vegetables. Further, this invention provides methods of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a structurally related mycotoxin breakdown product, by transforming the APAO polynucleotide, alone or in combination with polynucleotides encoding a fumonisin esterase, into plant cells.

The polynucleotides of the present invention can also be used as a selectable marker for plant transformation. By transforming plant cells with an expression cassette comprising a polynucleotide of the present invention and then placing the plant cells on media containing FB1, AP1 or a phytotoxic analog, only the plant cells expressing the polynucleotide of the present invention would survive.

Another embodiment of the present invention is the use of the enzyme fumonisin esterase and APAO by themselves or in combination as reagents for detecting fumonisin and structurally related toxins.

Brief Description of the Drawings

Figure 1 shows a 3-dimensional model of APAO (1B) based on the crystal structure of a related amine oxidase from maize, maize polyamine oxidase (MPAO) (1A). The sites for possible mutation of APAO to alter glycosylation sites or cysteine residues are shown.

Figure 2 shows a 3-dimensional model of APAO (2B) based on the crystal structure of a related amine oxidase from maize MPAO (2B). The substrate binding holes are shown as circles.

Detailed Description of the Invention

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Unless mentioned otherwise, the techniques employed or contemplated herein are standard methodologies well known to one of ordinary skill in the art. The materials, methods and examples are illustrative only and not limiting. The following is presented by way of illustration and is not intended to limit the scope of the invention.

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of botany, microbiology, tissue culture, molecular biology, chemistry, biochemistry and recombinant DNA technology, which are within the skill of the art. Such techniques are explained fully in the literature. See, e.g., J. H. Langenheim and K. V. Thimann, *Botany: Plant Biology and Its Relation to Human Affairs* (1982) John Wiley; *Cell Culture and Somatic Cell Genetics of Plants*, Vol. 1 (I. K. Vasil, ed. 1984); R. V. Stanier, J. L. Ingraham, M. L. Wheelis, and P. R. Painter, *The Microbial World*, (1986) 5th Ed., Prentice-Hall; O. D. Dhringra and J. B. Sinclair, *Basic Plant Pathology Methods*, (1985) CRC Press; Maniatis, Fritsch & Sambrook, *Molecular Cloning: A Laboratory Manual* (1982); *DNA Cloning*, Vols. I and II (D. N. Glover ed. 1985); *Oligonucleotide Synthesis* (M. J. Gait ed. 1984); *Nucleic Acid Hybridization* (B. D. Hames & S. J. Higgins eds. 1984); and the series *Methods in Enzymology* (S. Colowick and N. Kaplan, eds., Academic Press, Inc.).

Units, prefixes, and symbols may be denoted in their SI accepted form. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. Numeric ranges are inclusive of the numbers defining the range. Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes. The terms defined below are more fully defined by reference to the specification as a whole.

In describing the present invention, the following terms will be employed, and are intended to be defined as indicated below.

By "microbe" is meant any microorganism (including both eukaryotic and prokaryotic microorganisms), such as fungi, yeast, bacteria, actinomycetes, algae and protozoa, as well as other unicellular structures.

A "fumonisin-producing microbe" is any microbe capable of producing the mycotoxin fumonisin or analogs thereof. Such microbes are generally members of the fungal genus *Fusarium*, as well as recombinantly derived organisms, which have been genetically altered to enable them to produce fumonisin or analogs thereof.

By "degrading fumonisin" is meant any modification to fumonisin, AP1, or any derivative of fumonisin or AP1 which causes a decrease or loss in its toxic activity, such as degradation to less than 1%, 5%, 10%, or 50% of original toxicity, with less than 10% being preferred. Such a change can comprise cleavage of any of the various bonds, oxidation, reduction, the addition or deletion of a chemical moiety, or any other change that affects the activity of the molecule. In a preferred embodiment, the modification includes hydrolysis of the ester linkage in the molecule as a first step and then oxidative deamination. Furthermore, chemically altered fumonisin can be isolated from cultures of microbes that produce an enzyme of this invention, such as growing the organisms on media containing radioactively-labeled fumonisin, tracing the label, and isolating the degraded toxin for further study. The degraded fumonisin can be compared to the active compound for its phytotoxicity or mammalian toxicity in known sensitive species, such as porcines, rabbits, and equines or in cell or tissue culture assays. Such toxicity assays are known in the art. For example, in plants a whole leaf bioassay can be used in which solutions of the active and inactive compound are applied to the leaves of sensitive plants. The leaves may be treated *in situ* or, alternatively, excised leaves may be used. The relative toxicity of the compounds can be estimated by grading the ensuing damage to the plant tissues and by measuring the size of lesions formed within a given time period. Other known assays can be performed at the cellular level, employing standard tissue culture methodologies e.g., using cell suspension cultures.

By "fumonisin esterase" is meant any enzyme capable of hydrolysis of the ester linkage in fumonisin or a structurally similar molecule such as AAL toxin. Two examples of such enzymes are ESP1 and BEST1 found in US patent no. 5,716,820, issued February 10, 1998; US patent no. 5,792,931, issued August 11, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997.

By "structurally related mycotoxin" is meant any mycotoxin having a chemical structure related to a fumonisin or AP1 such as AAL toxin, fumonisin B1, fumonisin B2, fumonisin B3, fumonisin B4, fumonisin C1, fumonisin A1 and A2, and their analogs or hydrolyzed forms, as well as other mycotoxins having similar chemical structures, including synthetically made analogs that contain a C-2 or C-1 amine group and one or more adjacent hydroxyl groups, that would be expected to be degraded by the activity of an enzyme of the present invention. The present invention is the first flavin amine oxidase known to attack a primary amine not located at C-1 (i.e. C-2 of AP1) and resulting in a keto rather than an aldehydic product.

It is understood that "AP1" or "amino polyol" as used here is to designate the hydrolyzed form of any fumonisin, FB1, FB2, FB3, FB4, AAL, or any other AP1-like compound, including a compound made synthetically, that contains a C-2 or C-1 amine group and one or more adjacent hydroxyl groups.

By "amplified" is meant the construction of multiple copies of a nucleic acid sequence or multiple copies complementary to the nucleic acid sequence using at least one of the nucleic acid sequences as a template. Amplification systems include the polymerase chain reaction (PCR) system, ligase chain reaction (LCR) system, nucleic acid sequence based amplification (NASBA, Cangene, Mississauga, Ontario), Q-Beta Replicase systems, transcription-based amplification system (TAS), and strand displacement amplification (SDA). See, e.g., *Diagnostic Molecular Microbiology: Principles and Applications*, D. H. Persing *et al.*, Ed., American Society for Microbiology, Washington, DC (1993). The product of amplification is termed an amplicon.

The term "conservatively modified variants" applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refer to those nucleic acids that encode identical or conservatively modified variants of the amino acid sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein. For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are "silent variations" and represent one species of conservatively modified variation. Every nucleic acid sequence herein that encodes a polypeptide also

describes every possible silent variation of the nucleic acid. One of ordinary skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine, one exception is *Micrococcus rubens*, for which GTG is the methionine codon (Ishizuka, *et al.*, *J. Gen'l Microbiol*, 139:425-432 (1993)) can be modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid, which encodes a polypeptide of the present invention, is implicit in each described polypeptide sequence and incorporated herein by reference.

As to amino acid sequences, one of skill will recognize that individual substitutions, deletions or additions to a nucleic acid, peptide, polypeptide, or protein sequence which alters, adds or deletes a single amino acid or a small percentage of amino acids in the encoded sequence is a "conservatively modified variant" when the alteration results in the substitution of an amino acid with a chemically similar amino acid. Thus, any number of amino acid residues selected from the group of integers consisting of from 1 to 15 can be so altered. Thus, for example, 1, 2, 3, 4, 5, 7, or 10 alterations can be made. Conservatively modified variants typically provide similar biological activity as the unmodified polypeptide sequence from which they are derived. For example, substrate specificity, enzyme activity, or ligand/receptor binding is generally at least 30%, 40%, 50%, 60%, 70%, 80%, or 90%, preferably 60-90% of the native protein for its native substrate. Conservative substitution tables providing functionally similar amino acids are well known in the art.

The following six groups each contain amino acids that are conservative substitutions for one another:

- 1) Alanine (A), Serine (S), Threonine (T);
- 2) Aspartic acid (D), Glutamic acid (E);
- 3) Asparagine (N), Glutamine (Q);
- 4) Arginine (R), Lysine (K);
- 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and
- 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

See also, Creighton (1984) *Proteins* W.H. Freeman and Company.

As used herein, "consisting essentially of" means the inclusion of additional sequences to an object polynucleotide where the additional sequences do not selectively hybridize, under stringent hybridization conditions, to the same cDNA as the

polynucleotide and where the hybridization conditions include a wash step in 0.1X SSC and 0.1% sodium dodecyl sulfate at 65°C.

By "encoding" or "encoded", with respect to a specified nucleic acid, is meant comprising the information for translation into the specified protein. A nucleic acid encoding a protein may comprise non-translated sequences (e.g., introns) within translated regions of the nucleic acid, or may lack such intervening non-translated sequences (e.g., as in cDNA). The information by which a protein is encoded is specified by the use of codons. Typically, the amino acid sequence is encoded by the nucleic acid using the "universal" genetic code. However, variants of the universal code, such as is present in some plant, animal, and fungal mitochondria, the bacterium *Mycoplasma capricolum* (*Proc. Natl. Acad. Sci. (USA)*, 82: 2306-2309 (1985)), or the ciliate *Macronucleus*, may be used when the nucleic acid is expressed using these organisms.

When the nucleic acid is prepared or altered synthetically, advantage can be taken of known codon preferences of the intended host where the nucleic acid is to be expressed. For example, although nucleic acid sequences of the present invention may be expressed in both monocotyledonous and dicotyledonous plant species, sequences can be modified to account for the specific codon preferences and GC content preferences of monocotyledonous plants or dicotyledonous plants as these preferences have been shown to differ (Murray *et al.* Nucl. Acids Res. 17: 477-498 (1989) and herein incorporated by reference). Thus, the maize preferred codon for a particular amino acid might be derived from known gene sequences from maize. Maize codon usage for 28 genes from maize plants is listed in Table 4 of Murray *et al.*, *supra*.

As used herein, "heterologous" in reference to a nucleic acid is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by deliberate human intervention. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form. A heterologous protein may originate from a foreign species or, if from the same species, is substantially modified from its original form by deliberate human intervention.

By "host cell" or "recombinantly engineered cell" is meant a cell, which contains a vector and supports the replication and/or expression of the expression vector. Host cells

may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, *Pichia*, insect, plant, amphibian, or mammalian cells. Preferably, host cells are monocotyledonous or dicotyledonous plant cells, including but not limited to maize, sorghum, sunflower, soybean, wheat, alfalfa, rice, cotton, canola, barley, millet, and tomato. A particularly preferred monocotyledonous host cell is a maize host cell.

The term "hybridization complex" includes reference to a duplex nucleic acid structure formed by two single-stranded nucleic acid sequences selectively hybridized with each other.

The term "introduced" in the context of inserting a nucleic acid into a cell, means "transfection" or "transformation" or "transduction" and includes reference to the incorporation of a nucleic acid into a eukaryotic or prokaryotic cell where the nucleic acid may be incorporated into the genome of the cell (e.g., chromosome, plasmid, plastid or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (e.g., transfected mRNA).

The term "isolated" refers to material, such as a nucleic acid or a protein, which is substantially or essentially free from components which normally accompany or interact with it as found in its naturally occurring environment. The isolated material optionally comprises material not found with the material in its natural environment. Nucleic acids, which are "isolated", as defined herein, are also referred to as "heterologous" nucleic acids.

Unless otherwise stated, the term "APAO nucleic acid" means a nucleic acid comprising a polynucleotide ("APAO polynucleotide") encoding an APAO polypeptide. The term APAO, unless otherwise stated can encompass both APAO and the functional, truncated version of APAO designated trAPAO.

As used herein, "nucleic acid" includes reference to a deoxyribonucleotide or ribonucleotide polymer in either single- or double-stranded form, and unless otherwise limited, encompasses known analogues having the essential nature of natural nucleotides in that they hybridize to single-stranded nucleic acids in a manner similar to naturally occurring nucleotides (e.g., peptide nucleic acids).

By "nucleic acid library" is meant a collection of isolated DNA or RNA molecules, which comprise and substantially represent the entire transcribed fraction of a genome of a specified organism. Construction of exemplary nucleic acid libraries, such as genomic and cDNA libraries, is taught in standard molecular biology references such as Berger and

Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology*, Vol. 152, Academic Press, Inc., San Diego, CA (Berger); Sambrook *et al.*, *Molecular Cloning - A Laboratory Manual*, 2nd ed., Vol. 1-3 (1989); and *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (1994 Supplement).

As used herein "operably linked" includes reference to a functional linkage between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. Generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in the same reading frame.

The term "ligated" or "ligation" refers to the process of forming phosphodiester bonds between two or more polynucleotides, which most often are double stranded DNAs. Techniques for ligation are well known in the art and protocols are described in standard laboratory manuals and references, such as, Sambrook, *et al. Molecular Cloning: A Laboratory Manual*, 2ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1989). The two polynucleotides can include, but are not limited to, a polynucleotide, which can function as a promoter, ligated to a polynucleotide capable of encoding a polypeptide or linking two polynucleotides each capable of encoding a polypeptide. In the case of joining two polynucleotides that each encode a polypeptide, a polynucleotide spacer region between the two polynucleotides may or may not be present. The spacer region may encode a polypeptide containing a protease cleavage site. Optionally, the spacer region may contain a polynucleotide cleavage site such as but not limited to a site for RNase cleavage or a self-cleaving ribozyme (*See, e.g., Tanner, FEMS Microbiol Rev*, 23(3):257-75 (1999)). Alternatively, the transcription of the two or more ligated polynucleotides may result in a polycistronic message. An example of a spacer sequence that would direct translation of downstream coding sequences is an intervening ribosomal entry site (IRES) (*See, e.g., Liu, et al., Anal Biochem*, 280(1):20-28 (2000)). The length of the spacer region may be of any length that results in a functional polypeptide or polypeptides. For example, the spacer region may be from 1 nucleotide to 1000 nucleotides, preferably 24 nucleotides in length.

As used herein, the term "plant" includes reference to whole plants, plant organs (e.g., leaves, stems, roots, etc.), seeds and plant cells and progeny of same. Plant cell, as used herein includes, without limitation, seeds suspension cultures, embryos, meristematic regions, callus tissue, leaves, roots, shoots, gametophytes, sporophytes, pollen, and microspores. The class of plants, which can be used in the methods of the invention, is generally as broad as the class of higher plants amenable to transformation techniques, including both monocotyledonous and dicotyledonous plants including species from the genera: *Cucurbita*, *Rosa*, *Vitis*, *Juglans*, *Fragaria*, *Lotus*, *Medicago*, *Onobrychis*, *Trifolium*, *Trigonella*, *Vigna*, *Citrus*, *Linum*, *Geranium*, *Manihot*, *Daucus*, *Arabidopsis*, *Brassica*, *Raphanus*, *Sinapis*, *Atropa*, *Capsicum*, *Datura*, *Hyoscyamus*, *Lycopersicon*, *Nicotiana*, *Solanum*, *Petunia*, *Digitalis*, *Majorana*, *Ciahorium*, *Helianthus*, *Lactuca*, *Bromus*, *Asparagus*, *Antirrhinum*, *Heterocallis*, *Nemesis*, *Pelargonium*, *Panieum*, *Pennisetum*, *Ranunculus*, *Senecio*, *Salpiglossis*, *Cucumis*, *Browaalia*, *Glycine*, *Pisum*, *Phaseolus*, *Lolium*, *Oryza*, *Avena*, *Hordeum*, *Secale*, *Allium*, and *Triticum*. A particularly preferred plant is *Zea mays*.

As used herein, "polynucleotide" includes reference to a deoxyribopolynucleotide, ribopolynucleotide, or analogs thereof that have the essential nature of a natural ribonucleotide in that they hybridize, under stringent hybridization conditions, to substantially the same nucleotide sequence as naturally occurring nucleotides and/or allow translation into the same amino acid(s) as the naturally occurring nucleotide(s). A polynucleotide can be full-length or a subsequence of a native or heterologous structural or regulatory gene. Unless otherwise indicated, the term includes reference to the specified sequence as well as the complementary sequence thereof. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides" as that term is intended herein. Moreover, DNAs or RNAs comprising unusual bases, such as inosine, or modified bases, such as tritylated bases, to name just two examples, are polynucleotides as the term is used herein. It will be appreciated that a great variety of modifications have been made to DNA and RNA that serve many useful purposes known to those of skill in the art. The term polynucleotide as it is employed herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including *inter alia*, simple and complex cells.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers.

As used herein "promoter" includes reference to a region of DNA upstream from the start of transcription and involved in recognition and binding of RNA polymerase and other proteins to initiate transcription. A "plant promoter" is a promoter capable of initiating transcription in plant cells. Exemplary plant promoters include, but are not limited to, those that are obtained from plants, plant viruses, and bacteria which comprise genes expressed in plant cells such *Agrobacterium* or *Rhizobium*. Examples are promoters that preferentially initiate transcription in certain tissues, such as leaves, roots, seeds, fibres, xylem vessels, tracheids, or sclerenchyma. Such promoters are referred to as "tissue preferred". A "cell type" specific promoter primarily drives expression in certain cell types in one or more organs, for example, vascular cells in roots or leaves. An "inducible" or "regulatable" promoter is a promoter, which is under environmental control. Examples of environmental conditions that may effect transcription by inducible promoters include anaerobic conditions or the presence of light. Another type of promoter is a developmentally regulated promoter, for example, a promoter that drives expression during pollen development. Tissue preferred, cell type specific, developmentally regulated, and inducible promoters constitute the class of "non-constitutive" promoters. A "constitutive" promoter is a promoter, which is active under most environmental conditions.

The term "APAO polypeptide or trAPAO polypeptide" refers to one or more amino acid sequences. The term is also inclusive of fragments, variants, homologs, alleles or precursors (e.g., preproteins or proproteins) thereof. An "APAO or trAPAO protein" comprises an APAO or trAPAO polypeptide.

As used herein "recombinant" includes reference to a cell or vector, that has been modified by the introduction of a heterologous nucleic acid or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found in identical form within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under expressed or not expressed at all as a result of deliberate human intervention. The term "recombinant" as used herein does not encompass the alteration of the cell or vector by naturally occurring events (e.g.,

spontaneous mutation, natural transformation/transduction/transposition) such as those occurring without deliberate human intervention.

As used herein, a "recombinant expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements, which permit transcription of a particular nucleic acid in a target cell. The recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid to be transcribed, and a promoter.

The term "residue" or "amino acid residue" or "amino acid" are used interchangeably herein to refer to an amino acid that is incorporated into a protein, polypeptide, or peptide (collectively "protein"). The amino acid may be a naturally occurring amino acid and, unless otherwise limited, may encompass known analogs of natural amino acids that can function in a similar manner as naturally occurring amino acids.

The term "selectively hybridizes" includes reference to hybridization, under stringent hybridization conditions, of a nucleic acid sequence to a specified nucleic acid target sequence to a detectably greater degree (e.g., at least 2-fold over background) than its hybridization to non-target nucleic acid sequences and to the substantial exclusion of non-target nucleic acids. Selectively hybridizing sequences typically have about at least 40% sequence identity, preferably 60-90% sequence identity, and most preferably 100% sequence identity (i.e., complementary) with each other.

The terms "stringent conditions" or "stringent hybridization conditions" include reference to conditions under which a probe will hybridize to its target sequence, to a detectably greater degree than other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences can be identified which can be up to 100% complementary to the probe (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Optimally, the probe is approximately 500 nucleotides in length,

but can vary greatly in length from less than 500 nucleotides to equal to the entire length of the target sequence.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (*e.g.*, 10 to 50 nucleotides) and at least about 60°C for long probes (*e.g.*, greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide or Denhardt's. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C. Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl, *Anal. Biochem.*, 138:267-284 (1984): $T_m = 81.5\text{ }^{\circ}\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61 (\% \text{ form}) - 500/L$; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe. T_m is reduced by about 1 °C for each 1% of mismatching; thus, T_m , hybridization and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with $\geq 90\%$ identity are sought, the T_m can be decreased 10 °C. Generally, stringent conditions are selected to be about 5 °C lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4 °C lower than the thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10 °C lower than the thermal melting point (T_m); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20 °C

lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T_m , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T_m of less than 45 °C (aqueous solution) or 32 °C (formamide solution) it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen, *Laboratory Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 "Overview of principles of hybridization and the strategy of nucleic acid probe assays", Elsevier, New York (1993); and *Current Protocols in Molecular Biology*, Chapter 2, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995). Unless otherwise stated, in the present application high stringency is defined as hybridization in 4X SSC, 5X Denhardt's (5g Ficoll, 5g polyvinylpyrrolidone, 5 g bovine serum albumin in 500ml of water), 0.1 mg/ml boiled salmon sperm DNA, and 25 mM Na phosphate at 65°C, and a wash in 0.1X SSC, 0.1% SDS at 65°C.

As used herein, "transgenic plant" includes reference to a plant, which comprises within its genome a heterologous polynucleotide. Generally, the heterologous polynucleotide is stably integrated within the genome such that the polynucleotide is passed on to successive generations. The heterologous polynucleotide may be integrated into the genome alone or as part of a recombinant expression cassette. "Transgenic" is used herein to include any cell, cell line, callus, tissue, plant part or plant, the genotype of which has been altered by the presence of heterologous nucleic acid including those transgenics initially so altered as well as those created by sexual crosses or asexual propagation from the initial transgenic. The term "transgenic" as used herein does not encompass the alteration of the genome (chromosomal or extra-chromosomal) by conventional plant breeding methods or by naturally occurring events such as random cross-fertilization, non-recombinant viral infection, non-recombinant bacterial transformation, non-recombinant transposition, or spontaneous mutation.

As used herein, "vector" includes reference to a nucleic acid used in transfection of a host cell and into which can be inserted a polynucleotide. Vectors are often replicons. Expression vectors permit transcription of a nucleic acid inserted therein.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides or polypeptides: (a) “reference sequence”, (b) “comparison window”, (c) “sequence identity”, (d) “percentage of sequence identity”, and (e) “substantial identity”.

5 (a) As used herein, “reference sequence” is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

10 (b) As used herein, “comparison window” means includes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence may be compared to a reference sequence and wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20
15 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

20 Methods of alignment of nucleotide and amino acid sequences for comparison are well known in the art. The local homology algorithm (Best Fit) of Smith and Waterman, *Adv. Appl. Math* may conduct optimal alignment of sequences for comparison. 2: 482 (1981); by the homology alignment algorithm (GAP) of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970); by the search for similarity method (Tfasta and Fasta) of Pearson and Lipman, *Proc. Natl. Acad. Sci.* 85: 2444 (1988); by computerized implementations of these
25 algorithms, including, but not limited to: CLUSTAL in the PC/Gene program by Intelligenetics, Mountain View, California, GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, Wisconsin, USA; the CLUSTAL program is well described by Higgins and Sharp, *Gene* 73: 237-244 (1988); Higgins and Sharp, *CABIOS* 5: 151-153
30 (1989); Corpet, *et al.*, *Nucleic Acids Research* 16: 10881-90 (1988); Huang, *et al.*, *Computer Applications in the Biosciences* 8: 155-65 (1992), and Pearson, *et al.*, *Methods in Molecular Biology* 24: 307-331 (1994). The preferred program to use for optimal

global alignment of multiple sequences is PileUp (Feng and Doolittle, *Journal of Molecular Evolution*, 25:351-360 (1987) which is similar to the method described by Higgins and Sharp, *CABIOS*, 5:151-153 (1989) and hereby incorporated by reference). The BLAST family of programs which can be used for database similarity searches includes:

5 BLASTN for nucleotide query sequences against nucleotide database sequences; BLASTX for nucleotide query sequences against protein database sequences; BLASTP for protein query sequences against protein database sequences; TBLASTN for protein query sequences against nucleotide database sequences; and TBLASTX for nucleotide query sequences against nucleotide database sequences. See, *Current Protocols in Molecular*

10 *Biology*, Chapter 19, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995).

GAP uses the algorithm of Needleman and Wunsch (J. Mol. Biol. 48: 443-453, 1970) to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps. GAP considers all possible alignments and

15 gap positions and creates the alignment with the largest number of matched bases and the fewest gaps. It allows for the provision of a gap creation penalty and a gap extension penalty in units of matched bases. GAP must make a profit of gap creation penalty number of matches for each gap it inserts. If a gap extension penalty greater than zero is chosen, GAP must, in addition, make a profit for each gap inserted of the length of the gap times

20 the gap extension penalty. Default gap creation penalty values and gap extension penalty values in Version 10 of the Wisconsin Genetics Software Package are 8 and 2, respectively. The gap creation and gap extension penalties can be expressed as an integer selected from the group of integers consisting of from 0 to 100. Thus, for example, the gap creation and gap extension penalties can be 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 30, 40, 50,

25 or greater.

GAP presents one member of the family of best alignments. There may be many members of this family, but no other member has a better quality. GAP displays four figures of merit for alignments: Quality, Ratio, Identity, and Similarity. The Quality is the metric maximized in order to align the sequences. Ratio is the quality divided by the

30 number of bases in the shorter segment. Percent Identity is the percent of the symbols that actually match. Percent Similarity is the percent of the symbols that are similar. Symbols that are across from gaps are ignored. A similarity is scored when the scoring matrix value

for a pair of symbols is greater than or equal to 0.50, the similarity threshold. The scoring matrix used in Version 10 of the Wisconsin Genetics Software Package is BLOSUM62 (see Henikoff & Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using the BLAST 2.0 suite of programs using default parameters. Altschul *et al.*, *Nucleic Acids Res.* 25:3389-3402 (1997).

As those of ordinary skill in the art will understand, BLAST searches assume that proteins can be modeled as random sequences. However, many real proteins comprise regions of nonrandom sequences, which may be homopolymeric tracts, short-period repeats, or regions enriched in one or more amino acids. Such low-complexity regions may be aligned between unrelated proteins even though other regions of the protein are entirely dissimilar. A number of low-complexity filter programs can be employed to reduce such low-complexity alignments. For example, the SEG (Wooten and Federhen, *Comput. Chem.*, 17:149-163 (1993)) and XNU (Claverie and States, *Comput. Chem.*, 17:191-201 (1993)) low-complexity filters can be employed alone or in combination.

(c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences includes reference to the residues in the two sequences, which are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g. charge or hydrophobicity) and therefore do not change the functional properties of the molecule. Where sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences, which differ by such conservative substitutions, are said to have "sequence similarity" or "similarity". Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, *e.g.*, according to the algorithm of Meyers and

Miller, *Computer Applic. Biol. Sci.*, 4: 11-17 (1988) *e.g.*, as implemented in the program PC/GENE (Intelligenetics, Mountain View, California, USA).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

(e) (i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has between 50-100% sequence identity, preferably at least 50% sequence identity, preferably at least 60% sequence identity, preferably at least 70%, more preferably at least 80%, more preferably at least 90% and most preferably at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of between 40-100%, preferably at least 55%, preferably at least 60%, more preferably at least 70%, 80%, 90%, and most preferably at least 95%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. The degeneracy of the genetic code allows for many amino acids substitutions that lead to variety in the nucleotide sequence that code for the same amino acid, hence it is possible that the DNA sequence could code for the same polypeptide but not hybridize to each other under stringent conditions. This may occur, *e.g.*, when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is that the polypeptide, which the first

nucleic acid encodes, is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e) (ii) The terms "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with between 55-100% sequence identity to a reference sequence preferably at least 55% sequence identity, preferably 60% preferably 70%, more preferably 80%, most preferably at least 90% or 95% sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970). An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution. In addition, a peptide can be substantially identical to a second peptide when they differ by a non-conservative change if the epitope that the antibody recognizes is substantially identical. Peptides, which are "substantially similar" share sequences as, noted above except that residue positions, which are not identical, may differ by conservative amino acid changes.

Fumonisin Degrading Organisms

The present invention is based on the discovery of organisms with the ability to degrade the mycotoxin fumonisin. In a search for a biological means of detoxifying fumonisins, several dematiaceous hyphomycetes were isolated from field-grown maize kernels. The fungi were found to be capable of growing on fumonisin B1 or B2 (FB1 or FB2) as a sole carbon source, degrading it partially or completely in the process. One species, identified as *Exophiala spinifera*, a "black yeast", was recovered from maize seed from diverse locations in the southeastern and south central US. The enzyme-active strain of *Exophiala spinifera* (ATCC 74269) was deposited (see US patent no. 5,716,820, issued February 10, 1998, US patent no. 5,792,931, issued August 11, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997). Other enzyme-active strains of *Exophiala spinifera* were used to isolate APAO polynucleotides. Isolate ESP002 was isolated from palm trees (ATCC 26089) and isolate ESP003 was isolated from maize seed. Another fungus from which APAO polynucleotides were isolated was *Rhinoctadiella atrovirens* (RAT 011).

Nucleic Acids

The present invention provides, *inter alia*, isolated nucleic acids of RNA, DNA, and analogs and/or chimeras thereof, comprising an APAO or trAPAO polynucleotide.

5 The present invention also includes polynucleotides optimized for expression in different organisms. For example, for expression of the polynucleotide in a maize plant, the sequence can be altered to account for specific codon preferences and to alter GC content as according to Murray *et al*, *supra*. Maize codon usage for 28 genes from maize plants is listed in Table 4 of Murray, *et al.*, *supra*.

10 The APAO or trAPAO nucleic acids of the present invention comprise isolated APAO or trAPAO polynucleotides which, are inclusive of:

(a) a polynucleotide encoding an APAO or trAPAO polypeptide of the sequences shown in SEQ ID NOS: 36, 38, 40, 42, 44, and 46, and conservatively modified and polymorphic variants thereof;

15 (b) a polynucleotide which selectively hybridizes to a polynucleotide of (a) or (b);

(c) a polynucleotide having at least 50% sequence identity with polynucleotides of (a) or (b);

(d) complementary sequences of polynucleotides of (a), (b), or (c); and

20 (e) a polynucleotide comprising at least 25 contiguous nucleotides from a polynucleotide of (a), (b), (c), or (d).

In addition, polynucleotides are presented that are a fusion of an APAO or trAPAO polynucleotide and the polynucleotide of a fumonisin esterase. The invention encompasses the sequences from *Exophiala* or *Rhinoctadiella* as well as sequences having sequence similarity with such sequences. It is recognized that the sequences of the invention can be used to isolate corresponding sequences in other organisms. Methods such as PCR, hybridization, and the like can be used to identify sequences having substantial sequence similarity to the sequences of the invention. See, for example, Sambrook, *et al.*, (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York) and Innis *et al.*, (1990) *PCR Protocols: Guide to Methods and Applications* (Academic Press, New York). Coding sequences isolated based on their sequence identity to the entire fumonisin degrading coding

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sequences set forth herein or to fragments thereof are encompassed by the present invention.

It is recognized that the sequences of the invention can be used to isolate similar sequences from other fumonisin degrading organisms. Likewise sequences from other fumonisin degrading organisms may be used in combination with the sequences of the present invention. See, for example, copending application entitled "Compositions and Methods for Fumonisin Detoxification", U.S. application serial number 60/092,953, filed concurrently herewith and herein incorporated by reference.

Plasmids containing the polynucleotide sequences of the invention were deposited with American Type Culture Collection (ATCC), Manassas, Virginia, and assigned Accession Nos. 98812, 98813, 98814, 98815, 98816, and PTA-32. These deposits will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. These deposits were made merely as a convenience for those of skill in the art and are not an admission that a deposit is required under 35 U.S.C. § 112.

Construction of Nucleic Acids

The isolated nucleic acids of the present invention can be made using (a) standard recombinant methods, (b) synthetic techniques, or combinations thereof. In some embodiments, the polynucleotides of the present invention will be cloned, amplified, or otherwise constructed from a fungus or bacteria.

The nucleic acids may conveniently comprise sequences in addition to a polynucleotide of the present invention. For example, a multi-cloning site comprising one or more endonuclease restriction sites may be inserted into the nucleic acid to aid in isolation of the polynucleotide. Also, translatable sequences may be inserted to aid in the isolation of the translated polynucleotide of the present invention. For example, a hexahistidine marker sequence provides a convenient means to purify the proteins of the present invention. The nucleic acid of the present invention - excluding the polynucleotide sequence - is optionally a vector, adapter, or linker for cloning and/or expression of a polynucleotide of the present invention. Additional sequences may be added to such cloning and/or expression sequences to optimize their function in cloning and/or expression, to aid in isolation of the polynucleotide, or to improve the introduction of the

polynucleotide into a cell. Typically, the length of a nucleic acid of the present invention less the length of its polynucleotide of the present invention is less than 20 kilobase pairs, often less than 15 kb, and frequently less than 10 kb. Use of cloning vectors, expression vectors, adapters, and linkers is well known in the art. Exemplary nucleic acids include such vectors as: M13, lambda ZAP Express, lambda ZAP II, lambda gt10, lambda gt11, pBK-CMV, pBK-RSV, pBluescript II, lambda DASH II, lambda EMBL 3, lambda EMBL 4, pWE15, SuperCos 1, SurfZap, Uni-ZAP, pBC, pBS+/-, pSG5, pBK, pCR-Script, pET, pSPUTK, p3'SS, pGEM, pSK+/-, pGEX, pSPORTI and II, pOPRSVI CAT, pOPI3 CAT, pXT1, pSG5, pPbac, pMbac, pMC1neo, pOG44, pOG45, pFRT β GAL, pNEO β GAL, pRS403, pRS404, pRS405, pRS406, pRS413, pRS414, pRS415, pRS416, lambda MOSSlox, and lambda MOSElox. Optional vectors for the present invention, include but are not limited to, lambda ZAP II, and pGEX. For a description of various nucleic acids see, for example, Stratagene Cloning Systems, Catalogs 1995, 1996, 1997 (La Jolla, CA); and, Amersham Life Sciences, Inc, Catalog '97 (Arlington Heights, IL).

Synthetic Methods for Constructing Nucleic Acids

The isolated nucleic acids of the present invention can also be prepared by direct chemical synthesis by methods such as the phosphotriester method of Narang *et al.*, *Meth. Enzymol.* 68: 90-99 (1979); the phosphodiester method of Brown *et al.*, *Meth. Enzymol.* 68: 109-151 (1979); the diethylphosphoramidite method of Beaucage *et al.*, *Tetra. Lett.* 22: 1859-1862 (1981); the solid phase phosphoramidite triester method described by Beaucage and Caruthers, *Tetra. Letts.* 22(20): 1859-1862 (1981), *e.g.*, using an automated synthesizer, *e.g.*, as described in Needham-VanDevanter *et al.*, *Nucleic Acids Res.*, 12: 6159-6168 (1984); and, the solid support method of US Patent No. 4,458,066. Chemical synthesis generally produces a single stranded oligonucleotide. This may be converted into double stranded DNA by hybridization with a complementary sequence, or by polymerization with a DNA polymerase using the single strand as a template. One of skill will recognize that while chemical synthesis of DNA is limited to sequences of about 100 bases, longer sequences may be obtained by the ligation of shorter sequences.

UTRs and Codon Preference

In general, translational efficiency has been found to be regulated by specific sequence elements in the 5' non-coding or untranslated region (5' UTR) of the RNA. Positive sequence motifs include translational initiation consensus sequences (Kozak, *Nucleic Acids Res.* 15:8125 (1987)) and the 5' cap structure (Drummond *et al.*, *Nucleic Acids Res.* 13:7375 (1985)). Negative elements include stable intramolecular 5' UTR stem-loop structures (Muesing *et al.*, *Cell* 48:691 (1987)) and AUG sequences or short open reading frames preceded by an appropriate AUG in the 5' UTR (Kozak, *supra*, Rao *et al.*, *Mol. and Cell. Biol.* 8:284 (1988)). Accordingly, the present invention provides 5' and/or 3' UTR regions for modulation of translation of heterologous coding sequences.

Further, the polypeptide-encoding segments of the polynucleotides of the present invention can be modified to alter codon usage. Altered codon usage can be employed to alter translational efficiency and/or to optimize the coding sequence for expression in a desired host or to optimize the codon usage in a heterologous sequence for expression in maize. Codon usage in the coding regions of the polynucleotides of the present invention can be analyzed statistically using commercially available software packages such as "Codon Preference" available from the University of Wisconsin Genetics Computer Group (see Devereaux *et al.*, *Nucleic Acids Res.* 12: 387-395 (1984)) or MacVector 4.1 (Eastman Kodak Co., New Haven, Conn.). Thus, the present invention provides a codon usage frequency characteristic of the coding region of at least one of the polynucleotides of the present invention. The number of polynucleotides (3 nucleotides per amino acid) that can be used to determine a codon usage frequency can be any integer from 3 to the number of polynucleotides of the present invention as provided herein. Optionally, the polynucleotides will be full-length sequences. An exemplary number of sequences for statistical analysis can be at least 1, 5, 10, 20, 50, or 100.

Sequence Shuffling

The present invention provides methods for sequence shuffling using polynucleotides of the present invention, and compositions resulting therefrom. Sequence shuffling is described in PCT publication No. 96/19256. See also, Zhang, J.- H., *et al. Proc. Natl. Acad. Sci. USA* 94:4504-4509 (1997) and Zhao, *et al.*, *Nature Biotech* 16:258-

261 (1998). Generally, sequence shuffling provides a means for generating libraries of polynucleotides having a desired characteristic, which can be selected or screened for. Libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides, which comprise sequence regions, which have substantial sequence identity and can be homologously recombined in vitro or in vivo. The population of sequence-recombined polynucleotides comprises a subpopulation of polynucleotides which possess desired or advantageous characteristics and which can be selected by a suitable selection or screening method. The characteristics can be any property or attribute capable of being selected for or detected in a screening system, and may include properties of: an encoded protein, a transcriptional element, a sequence controlling transcription, RNA processing, RNA stability, chromatin conformation, translation, or other expression property of a gene or transgene, a replicative element, a protein-binding element, or the like, such as any feature which confers a selectable or detectable property. In some embodiments, the selected characteristic will be an altered K_m and/or K_{cat} over the wild-type protein as provided herein. In other embodiments, a protein or polynucleotide generated from sequence shuffling will have a substrate binding affinity greater than the non-shuffled wild-type polynucleotide. In yet other embodiments, a protein or polynucleotide generated from sequence shuffling will have an altered pH optimum as compared to the non-shuffled wild-type polynucleotide. The increase in such properties can be at least 110%, 120%, 130%, 140% or greater than 150% of the wild-type value.

Recombinant Expression Cassettes

The present invention further provides recombinant expression cassettes comprising a nucleic acid of the present invention. A nucleic acid sequence coding for the desired polynucleotide of the present invention, for example a cDNA or a genomic sequence encoding a polypeptide long enough to code for an active protein of the present invention, can be used to construct a recombinant expression cassette which can be introduced into the desired host cell. A recombinant expression cassette will typically comprise a polynucleotide of the present invention operably linked to transcriptional initiation regulatory sequences which will direct the transcription of the polynucleotide in the intended host cell, such as tissues of a transformed plant.

For example, plant expression vectors may include (1) a cloned plant gene under the transcriptional control of 5' and 3' regulatory sequences and (2) a dominant selectable marker. Such plant expression vectors may also contain, if desired, a promoter regulatory region (e.g., one conferring inducible or constitutive, environmentally- or developmentally-regulated, or cell- or tissue-specific/selective expression), a transcription initiation start site, a ribosome binding site, an RNA processing signal, a transcription termination site, and/or a polyadenylation signal.

A plant promoter fragment can be employed which will direct expression of a polynucleotide of the present invention in all tissues of a regenerated plant. Such promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the 1'- or 2'- promoter derived from T-DNA of *Agrobacterium tumefaciens*, the Smas promoter, the cinnamyl alcohol dehydrogenase promoter (US Patent No. 5,683,439), the *Nos* promoter, the rubisco promoter, the GRP1-8 promoter, the 35S promoter from cauliflower mosaic virus (CaMV), as described in Odell et al., (1985), *Nature*, 313:810-812, rice actin (McElroy et al., (1990), *Plant Cell*, 163-171); ubiquitin (Christensen et al., (1992), *Plant Mol. Biol.* 12:619-632; and Christensen, et al., (1992), *Plant Mol. Biol.* 18:675-689); pEMU (Last, et al., (1991), *Theor. Appl. Genet.* 81:581-588); MAS (Velten et al., (1984), *EMBO J.* 3:2723-2730); and maize H3 histone (Lepetit et al., (1992), *Mol. Gen. Genet.* 231:276-285; and Atanassova et al., (1992), *Plant Journal* 2(3):291-300), the Rsyn7 as described in published PCT Application WO 97/44756, ALS promoter, as described in published PCT Application WO 96/30530, and other transcription initiation regions from various plant genes known to those of skill. For the present invention ubiquitin is the preferred promoter for expression in monocot plants.

Alternatively, the plant promoter can direct expression of a polynucleotide of the present invention in a specific tissue or may be otherwise under more precise environmental or developmental control. Such promoters are referred to here as "inducible" promoters. Environmental conditions that may effect transcription by inducible promoters include pathogen attack, anaerobic conditions, or the presence of light. Examples of inducible promoters are the *Adh1* promoter, which is inducible by hypoxia or

cold stress, the Hsp70 promoter, which is inducible by heat stress, and the PPDK promoter, which is inducible by light.

Examples of promoters under developmental control include promoters that initiate transcription only, or preferentially, in certain tissues, such as leaves, roots, fruit, seeds, or flowers. The operation of a promoter may also vary depending on its location in the genome. Thus, an inducible promoter may become fully or partially constitutive in certain locations.

If polypeptide expression is desired, it is generally desirable to include a polyadenylation region at the 3'-end of a polynucleotide coding region. The polyadenylation region can be derived from a variety of plant genes, or from T-DNA. The 3' end sequence to be added can be derived from, for example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene. Examples of such regulatory elements include, but are not limited to, 3' termination and/or polyadenylation regions such as those of the *Agrobacterium tumefaciens* nopaline synthase (nos) gene (Bevan et al., (1983), *Nucl. Acids Res.* 12:369-385); the potato proteinase inhibitor II (PINII) gene (Keil, et al., (1986), *Nucl. Acids Res.* 14:5641-5650; and An et al., (1989), *Plant Cell* 1:115-122); and the CaMV 19S gene (Mogen et al., (1990), *Plant Cell* 2:1261-1272).

An intron sequence can be added to the 5' untranslated region or the coding sequence of the partial coding sequence to increase the amount of the mature message that accumulates in the cytosol. Inclusion of a spliceable intron in the transcription unit in both plant and animal expression constructs has been shown to increase gene expression at both the mRNA and protein levels up to 1000-fold. Buchman and Berg, *Mol. Cell Biol.* 8: 4395-4405 (1988); Callis et al., *Genes Dev.* 1: 1183-1200 (1987). Such intron enhancement of gene expression is typically greatest when placed near the 5' end of the transcription unit. Use of maize introns Adh1-S intron 1, 2, and 6, the Bronze-1 intron are known in the art. See generally, *The Maize Handbook*, Chapter 116, Freeling and Walbot, Eds., Springer, New York (1994).

Plant signal sequences, including, but not limited to, signal-peptide encoding DNA/RNA sequences which target proteins to the extracellular matrix of the plant cell (Dratewka-Kos, et al., (1989), *J. Biol. Chem.* 264:4896-4900), the *Nicotiana plumbaginifolia* extension gene (DeLoose, et al., (1991), *Gene* 99:95-100), signal peptides

which target proteins to the vacuole like the sweet potato sporamin gene (Matsuka, et al., (1991), *PNAS* 88:834) and the barley lectin gene (Wilkins, et al., (1990), *Plant Cell*, 2:301-313), signal peptides which cause proteins to be secreted such as that of PR1b (Lind, et al., (1992), *Plant Mol. Biol.* 18:47-53), or the barley alpha amylase (BAA) (Rahmatullah, et al., *Plant Mol. Biol.* 12:119 (1989)) and hereby incorporated by reference), or from the present invention the signal peptide from the ESP1 or BEST1 gene, or signal peptides which target proteins to the plastids such as that of rapeseed enoyl-Acp reductase (Verwaert, et al., (1994), *Plant Mol. Biol.* 26:189-202) are useful in the invention. The barley alpha amylase signal sequence operably linked to the trAPAO or APAO polynucleotide is the preferred construct for expression in maize for the present invention.

The vector comprising the sequences from a polynucleotide of the present invention will typically comprise a marker gene, which confers a selectable phenotype on plant cells. Usually, the selectable marker gene will encode antibiotic resistance, with suitable genes including genes coding for resistance to the antibiotic spectinomycin (e.g., the aada gene), the streptomycin phosphotransferase (SPT) gene coding for streptomycin resistance, the neomycin phosphotransferase (NPTII) gene encoding kanamycin or geneticin resistance, the hygromycin phosphotransferase (HPT) gene coding for hygromycin resistance, genes coding for resistance to herbicides which act to inhibit the action of acetolactate synthase (ALS), in particular the sulfonylurea-type herbicides (e.g., the acetolactate synthase (ALS) gene containing mutations leading to such resistance in particular the S4 and/or Hra mutations), genes coding for resistance to herbicides which act to inhibit action of glutamine synthase, such as phosphinothricin or basta (e.g., the *bar* gene), or other such genes known in the art. The *bar* gene encodes resistance to the herbicide basta, and the ALS gene encodes resistance to the herbicide chlorsulfuron.

Alternatively, the invention, itself, could be used as a method for selection of transformants, in other words as a selectable marker. An APAO or trAPAO polynucleotide operably linked to a promoter and then transformed into a plant cell by any of the methods described in the present application would express the degradative enzyme. When the plant cells are placed in the presence of fumonisin, AP1, or a phytotoxic analog in culture only the transformed cells would be able to grow. In another embodiment, the plant cell could be transformed with both a polynucleotide for APAO and a polynucleotide for fumonisin esterase. The selective agent in this case could be either AP1 or fumonisin

or any structural analog. Thus, growth of plant cells in the presence of a mycotoxin favors the survival of plant cells that have been transformed to express the coding sequence that codes for one of the enzymes of this invention and degrades the toxin. When the APAO or trAPAO cassette with or without the fumonisin esterase polynucleotide, is co-transformed with another gene of interest and then placed in the presence of fumonisin, AP1 or a phytotoxic analog, this invention would allow for selection of only those plant cells that contain the gene of interest. In the past antibiotic resistance genes have been used as selectable markers. Given the current concerns by consumers and environmentalist over use of antibiotic genes and the possibility of resistant microorganisms arising due to this use, a non-antibiotic resistant selectable marker system such as the present invention, fulfills this very important need.

Typical vectors useful for expression of genes in higher plants are well known in the art and include vectors derived from the tumor-inducing (Ti) plasmid of *Agrobacterium tumefaciens* described by Rogers *et al.*, Meth. In Enzymol., 153:253-277 (1987). These vectors are plant integrating vectors in that on transformation, the vectors integrate a portion of vector DNA into the genome of the host plant. Exemplary *A. tumefaciens* vectors useful herein are plasmids pKYLX6 and pKYLX7 of Schardl *et al.*, Gene, 61:1-11 (1987) and Berger *et al.*, Proc. Natl. Acad. Sci. U.S.A., 86:8402-8406 (1989). Another useful vector herein is plasmid pBI101.2 that is available from CLONTECH Laboratories, Inc. (Palo Alto, CA).

Expression of Proteins in Host Cells

Using the nucleic acids of the present invention, one may express a protein of the present invention in a recombinantly engineered cell such as bacteria, yeast, insect, mammalian, or preferably plant cells. The cells produce the protein in a non-natural condition (e.g., in quantity, composition, location, and/or time), because they have been genetically altered through human intervention to do so.

It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of a nucleic acid encoding a protein of the present invention. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of isolated nucleic acids encoding a protein of the present invention will typically be achieved by operably linking, for example, the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding a protein of the present invention. To obtain high level expression of a cloned gene, it is desirable to construct expression vectors which contain, at the minimum, a strong promoter, such as ubiquitin, to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator. Constitutive promoters are classified as providing for a range of constitutive expression. Thus, some are weak constitutive promoters, and others are strong constitutive promoters. Generally, by "weak promoter" is intended a promoter that drives expression of a coding sequence at a low level. By "low level" is intended at levels of about 1/10,000 transcripts to about 1/100,000 transcripts to about 1/500,000 transcripts. Conversely, a "strong promoter" drives expression of a coding sequence at a "high level", or about 1/10 transcripts to about 1/100 transcripts to about 1/1,000 transcripts.

One of skill would recognize that modifications could be made to a protein of the present invention without diminishing its biological activity. Some modifications may be made to facilitate the cloning, expression, or incorporation of the targeting molecule into a fusion protein. Such modifications are well known to those of skill in the art and include, for example, a methionine added at the amino terminus to provide an initiation site, or additional amino acids (*e.g.*, poly His) placed on either terminus to create conveniently located restriction sites or termination codons or purification sequences.

A. Expression in Prokaryotes

Prokaryotic cells may be used as hosts for expression. Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Commonly used prokaryotic control sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang et al., Nature 198:1056 (1977)),

the tryptophan (trp) promoter system (Goeddel et al., Nucleic Acids Res. 8:4057 (1980)) and the lambda derived P L promoter and N-gene ribosome binding site (Shimatake *et al.*, Nature 292:128 (1981)). The inclusion of selection markers in DNA vectors transfected in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol.

The vector is selected to allow introduction of the gene of interest into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA. Expression systems for expressing a protein of the present invention are available using *Bacillus sp.* and *Salmonella* (Palva, *et al.*, *Gene* 22: 229-235 (1983); Mosbach, *et al.*, *Nature* 302: 543-545 (1983)). The pGEX-4T-1 plasmid vector from Pharmacia is the preferred *E. coli* expression vector for the present invention.

B. Expression in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines, plant and mammalian cells, are known to those of skill in the art. As explained briefly below, the present invention can be expressed in these eukaryotic systems. In some embodiments, transformed/transfected plant cells, as discussed *infra*, are employed as expression systems for production of the proteins of the instant invention.

Synthesis of heterologous proteins in yeast is well known. Sherman, F., *et al.*, *Methods in Yeast Genetics*, Cold Spring Harbor Laboratory (1982) is a well recognized work describing the various methods available to produce the protein in yeast. Two widely utilized yeasts for production of eukaryotic proteins are *Saccharomyces cerevisiae* and *Pichia pastoris*. Vectors, strains, and protocols for expression in *Saccharomyces* and *Pichia* are known in the art and available from commercial suppliers (e.g., Invitrogen). Suitable vectors usually have expression control sequences, such as promoters, including 3-phosphoglycerate kinase or alcohol oxidase, and an origin of replication, termination sequences and the like as desired.

A protein of the present invention, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates or the

pellets. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassay of other standard immunoassay techniques.

The sequences encoding proteins of the present invention can also be ligated to various expression vectors for use in transfecting cell cultures of, for instance, mammalian, insect, or plant origin. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. A number of suitable host cell lines capable of expressing intact proteins have been developed in the art, and include the HEK293, BHK21, and CHO cell lines. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (*e.g.*, the CMV promoter, a HSV *tk* promoter or *pgk* (phosphoglycerate kinase) promoter), an enhancer (Queen *et al.*, *Immunol. Rev.* 89: 49 (1986)), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites (*e.g.*, an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells useful for production of proteins of the present invention are available, for instance, from the American Type Culture Collection Catalogue of Cell Lines and Hybridomas (7th edition, 1992).

Appropriate vectors for expressing proteins of the present invention in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth, and *Drosophila* cell lines such as a Schneider cell line (See Schneider, *J. Embryol. Exp. Morphol.* 27: 353-365 (1987).

As with yeast, when higher animal or plant host cells are employed, polyadenylation or transcription terminator sequences are typically incorporated into the vector. An example of a terminator sequence is the polyadenylation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VP1 intron from SV40 (Sprague, *et al.*, *J. Virol.* 45: 773-781 (1983)). Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors. Saveria-Campo, M., Bovine Papilloma Virus DNA a Eukaryotic Cloning Vector in *DNA Cloning Vol. II a Practical Approach*, D.M. Glover, Ed., IRL Press, Arlington, Virginia pp. 213-238 (1985).

In addition, one of the genes for fumonisin esterase or the APAO or trAPAO placed in the appropriate plant expression vector can be used to transform plant cells. The

enzyme can then be isolated from plant callus or the transformed cells can be used to regenerate transgenic plants. Such transgenic plants can be harvested, and the appropriate tissues (seed or leaves, for example) can be subjected to large scale protein extraction and purification techniques, and the fumonisin degradation enzymes or APAO can be isolated for use in fumonisin and fumonisin hydrolysis product detoxification processes.

Plant Transformation Methods

Numerous methods for introducing foreign genes into plants are known and can be used to insert an APAO or trAPAO polynucleotide into a plant host, including biological and physical plant transformation protocols. See, for example, Miki et al., (1993), "Procedure for Introducing Foreign DNA into Plants", In: *Methods in Plant Molecular Biology and Biotechnology*, Glick and Thompson, eds., CRC Press, Inc., Boca Raton, pages 67-88. The methods chosen vary with the host plant, and include chemical transfection methods such as calcium phosphate, microorganism-mediated gene transfer such as *Agrobacterium* (Horsch, et al., (1985), *Science* 227:1229-31), electroporation, micro-injection, and biolistic bombardment.

Expression cassettes and vectors and *in vitro* culture methods for plant cell or tissue transformation and regeneration of plants are known and available. See, for example, Gruber, et al., (1993), "Vectors for Plant Transformation" In: *Methods in Plant Molecular Biology and Biotechnology*, Glick and Thompson, eds. CRC Press, Inc., Boca Raton, pages 89-119.

Agrobacterium-mediated Transformation

The most widely utilized method for introducing an expression vector into plants is based on the natural transformation system of *Agrobacterium*. *A. tumefaciens* and *A. rhizogenes* are plant pathogenic soil bacteria, which genetically transform plant cells. The Ti and Ri plasmids of *A. tumefaciens* and *A. rhizogenes*, respectively, carry genes responsible for genetic transformation of plants. See, for example, Kado, (1991), *Crit. Rev. Plant Sci.* 10:1. Descriptions of the *Agrobacterium* vector systems and methods for *Agrobacterium*-mediated gene transfer are provided in Gruber et al., *supra*; Miki, et al., *supra*; and Moloney et al., (1989), *Plant Cell Reports* 8:238.

Similarly, the gene can be inserted into the T-DNA region of a Ti or Ri plasmid derived from *A. tumefaciens* or *A. rhizogenes*, respectively. Thus, expression cassettes can be constructed as above, using these plasmids. Many control sequences are known which when coupled to a heterologous coding sequence and transformed into a host organism show fidelity in gene expression with respect to tissue/organ specificity of the original coding sequence. See, e.g., Benfey, P. N., and Chua, N. H. (1989) *Science* 244: 174-181. Particularly suitable control sequences for use in these plasmids are promoters for constitutive leaf-specific expression of the gene in the various target plants. Other useful control sequences include a promoter and terminator from the nopaline synthase gene (NOS). The NOS promoter and terminator are present in the plasmid pARC2, available from the American Type Culture Collection and designated ATCC 67238. If such a system is used, the virulence (*vir*) gene from either the Ti or Ri plasmid must also be present, either along with the T-DNA portion, or via a binary system where the *vir* gene is present on a separate vector. Such systems, vectors for use therein, and methods of transforming plant cells are described in US Pat. No. 4,658,082; US application Ser. No. 913,914, filed Oct. 1, 1986, as referenced in US Patent 5,262,306, issued November 16, 1993 to Robeson, et al.; and Simpson, R. B., et al. (1986) *Plant Mol. Biol.* 6: 403-415 (also referenced in the '306 patent); all incorporated by reference in their entirety.

Once constructed, these plasmids can be placed into *A. rhizogenes* or *A. tumefaciens* and these vectors used to transform cells of plant species, which are ordinarily susceptible to *Fusarium* or *Alternaria* infection. Several other transgenic plants are also contemplated by the present invention including but not limited to soybean, corn, sorghum, alfalfa, rice, clover, cabbage, banana, coffee, celery, tobacco, cowpea, cotton, melon and pepper. The selection of either *A. tumefaciens* or *A. rhizogenes* will depend on the plant being transformed thereby. In general *A. tumefaciens* is the preferred organism for transformation. Most dicotyledonous plants, some gymnosperms, and a few monocotyledonous plants (e.g. certain members of the *Liliales* and *Arales*) are susceptible to infection with *A. tumefaciens*. *A. rhizogenes* also has a wide host range, embracing most dicots and some gymnosperms, which includes members of the *Leguminosae*, *Compositae*, and *Chenopodiaceae*. Monocot plants can now be transformed with some success. European Patent Application Publication Number 604 662 A1 to Hiei *et al.* discloses a method for transforming monocots using *Agrobacterium*. Saito *et al.* discloses a method

for transforming monocots with *Agrobacterium* using the scutellum of immature embryos (European Application 672 752 A1). Ishida *et al.* discusses a method for transforming maize by exposing immature embryos to *A. tumefaciens* (Ishida *et al.*, *Nature Biotechnology*, 1996, 14:745-750).

Once transformed, these cells can be used to regenerate transgenic plants, capable of degrading fumonisin. For example, whole plants can be infected with these vectors by wounding the plant and then introducing the vector into the wound site. Any part of the plant can be wounded, including leaves, stems and roots. Alternatively, plant tissue, in the form of an explant, such as cotyledonary tissue or leaf disks, can be inoculated with these vectors, and cultured under conditions, which promote plant regeneration. Roots or shoots transformed by inoculation of plant tissue with *A. rhizogenes* or *A. tumefaciens*, containing the gene coding for the fumonisin degradation enzyme, can be used as a source of plant tissue to regenerate fumonisin-resistant transgenic plants, either via somatic embryogenesis or organogenesis. Examples of such methods for regenerating plant tissue are disclosed in Shahin, E. A. (1985) *Theor. Appl. Genet.* 69:235-240; US Pat. No. 4,658,082; Simpson, R. B., et al. (1986) *Plant Mol. Biol.* 6: 403-415; and U.S. patent applications Ser. Nos. 913,913 and 913,914, both filed Oct. 1, 1986, as referenced in U.S. Patent 5,262,306, issued November 16, 1993 to Robeson, et al.; the entire disclosures therein incorporated herein by reference.

Direct Gene Transfer

Despite the fact that the host range for *Agrobacterium*-mediated transformation is broad, some major cereal crop species and gymnosperms have generally been recalcitrant to this mode of gene transfer, even though some success has recently been achieved in rice (Hiei *et al.*, (1994), *The Plant Journal* 6:271-282). Several methods of plant transformation, collectively referred to as direct gene transfer, have been developed as an alternative to *Agrobacterium*-mediated transformation.

A generally applicable method of plant transformation is microprojectile-mediated transformation, where DNA is carried on the surface of microprojectiles measuring about 1 to 4 μm . The expression vector is introduced into plant tissues with a biolistic device that accelerates the microprojectiles to speeds of 300 to 600 m/s which is sufficient to penetrate the plant cell walls and membranes. (Sanford *et al.*, (1987), *Part. Sci. Technol.* 5:27;

Sanford, 1988, *Trends Biotech* 6:299; Sanford, (1990), *Physiol. Plant* 79:206; Klein et al., (1992), *Biotechnology* 10:268).

Another method for physical delivery of DNA to plants is sonication of target cells as described in Zang et al., (1991), *BioTechnology* 9:996. Alternatively, liposome or spheroplast fusions have been used to introduce expression vectors into plants. See, for example, Deshayes et al., (1985), *EMBO J.* 4:2731; and Christou et al., (1987), *PNAS USA* 84:3962. Direct uptake of DNA into protoplasts using CaCl_2 precipitation, polyvinyl alcohol, or poly-L-ornithine has also been reported. See, for example, Hain et al., (1985), *Mol. Gen. Genet.* 199:161; and Draper et al., (1982), *Plant Cell Physiol.* 23:451.

Electroporation of protoplasts and whole cells and tissues has also been described. See, for example, Donn et al., (1990), In: *Abstracts of the VIIth Int'l. Congress on Plant Cell and Tissue Culture IAPTC*, A2-38, page 53; D'Halluin et al., (1992), *Plant Cell* 4:1495-1505; and Spencer et al., (1994), *Plant Mol. Biol.* 24:51-61.

Thus, polynucleotide encoding a polypeptide able to degrade fumonisin or AP1 can be isolated and cloned in an appropriate vector and inserted into an organism normally sensitive to the *Fusarium* or its toxins. Furthermore, the polynucleotide imparting fumonisin or AP1 degradative activity can be transferred into a suitable plasmid, and transformed into a plant. Thus, a fumonisin or AP1 degrading transgenic plant can be produced. Organisms expressing the polynucleotide can be easily identified by their ability to degrade fumonisin or AP1. The protein capable of degrading fumonisin or AP1 can be isolated and characterized using techniques well known in the art.

APAO or trAPAO in a Transgenic Plant

Fumonisin esterase reduces but does not eliminate the toxicity of fumonisins. Therefore a second enzymatic modification to further reduce or abolish toxicity is desirable. The partially purified APAO enzyme from *Exophiala spinifera* has little or no activity on intact FB1, a form of fumonisin. However, recombinant APAO enzyme from *Exophiala spinifera*, expressed in *E. coli*, has significant but reduced activity on intact FB1 and other B-series fumonisins. APAO or trAPAO thus could potentially be used without fumonisin esterase since the amine group is the major target for detoxification. Alternatively, the two genes, fumoninsin esterase and APAO (or trAPAO) can be used together for degrading toxins.

APAO is predicted to be an enzyme that, when by itself or co-expressed in a heterologous expression system along with fumonisin esterase (either ESP1 or BEST1), will result in the production of 2-oxo-FB1 and/or 2-oxo pentol (2-OP) from fumonisin B1. The substrate range of recombinant, *E. coli*-expressed APAO is limited to fumonisins and their hydrolysis products and does not include amino acids, sphingolipid precursors such as phytosphingosine, or polyamines such as spermidine. Thus, APAO is highly specific for fumonisin-like amines, and thus would have little deleterious effect on other cellular metabolites. In addition, if it is extracellularly localized, it will limit any contact with biologically important amines that might also be substrates. The end result will be a more effective detoxification of fumonisins than can be achieved with esterase alone.

The oxidase activity of APAO is predicted to result in generation of hydrogen peroxide in stoichiometric amounts relative to AP1 or fumonisin oxidized. This may prove to be an additional benefit of this enzyme, since hydrogen peroxide is both antimicrobial and is thought to contribute to the onset of a defense response in plants (Przemylaw, *Biochem J.*, 322:681-692 (1997), Lamb, *et al.*, *Ann Rev Plant Physiol Plant Mol Bio* 48:251-275 (1997), and Alvarez, *et al.*, *Oxidative Stress and the Molecular Biology of Antioxidant Defenses*, Cold Spring Harbor Press, 815-839 (1997)).

Because one of the embodiments of the present invention is to have both a fumonisin esterase polynucleotide and an APAO or trAPAO polynucleotide present in a plant, there are several ways to introduce more than one polynucleotide in a plant. One way is to transform plant tissue with polynucleotides to both fumonisin esterase and APAO or trAPAO at the same time. In some tissue culture systems it is possible to transform callus with one polynucleotide and then after establishing a stable culture line containing the first polynucleotide, transform the callus a second time with the second polynucleotide. One could also transform plant tissue with one polynucleotide, regenerate whole plants, then transform the second polynucleotide into plant tissue and regenerate whole plants. The final step would then be to cross a plant containing the first polynucleotide with a plant containing the second polynucleotide and select for progeny containing both polynucleotides.

Another method is to create a fusion protein between esterase and APAO or trAPAO, preferably with a spacer region between the two polypeptides. Both enzymes would be

active although tethered to each other. In addition, an enzyme cleavage site engineered in the spacer region, would allow cleavage by an endogenous or introduced protease.

Transgenic plants containing both a fumonisin esterase enzyme and/or the APAO enzyme and thus able to degrade fumonisin or a structurally related mycotoxin would be able to reduce or eliminate the pathogenicity of any microorganism that uses fumonisin or a structurally related mycotoxin as a mode of entry to infect a plant. Fungal pathogens frequently use toxins to damage plants and weaken cell integrity in order to gain entry and expand infection in a plant. By preventing the damage induced by a toxin, a plant would be able to prevent the establishment of the pathogen and thereby become tolerant or resistant to the pathogen.

Another benefit of fumonisin degradation is the production of hydrogen peroxide. When fumonisin or AP1 is oxididatively deaminated at C-2, as occurs by exposure to APAO or trAPAO enzyme, hydrogen peroxide is produced as a by-product. Hydrogen peroxide production can trigger enhanced resistance responses in a number of ways. 1) Hydrogen peroxide has direct antimicrobial activity. 2) Hydrogen peroxide acts as a substrate for peroxidases associated with lignin polymerization and hence cell wall strengthening. 3) Via still to be determined mechanisms, hydrogen peroxide acts as a signal for activation of expression of defense related genes, including those that result in stimulation of salicylic acid accumulation. Salicylic acid is thought to act an endogenous signal molecule that triggers expression of genes coding for several classes of pathogenesis-related proteins. Moreover, salicylic acid may set up the oxidative burst and thus act in a feedback loop enhancing its own synthesis. Salicylic acid may also be involved in hypersensitive cell death by acting as an inhibitor of catalase, an enzyme that removes hydrogen peroxide. 4) Hydrogen peroxide may trigger production of additional defense compounds such as phytoalexins, antimicrobial low molecular weight compounds. For a review on the role of the oxidative burst and SA please see Lamb, C. and Dixon, R.A., *Ann. Rev. Plant Physiol. Plant Mol. Biol.*, 48: 251-275 (1997).

Detoxification of Harvested Grain, Silage, or Contaminated Food Crop

The present invention also relates to a method of detoxifying a fumonisin or a structurally related mycotoxin with an APAO enzyme during the processing of grain for animal or human food consumption, during the processing of plant material for silage, or

food crops contaminated with a toxin producing microbe, such as but not limited to, tomato. Since the atmospheric ammoniation of corn has proven to be an ineffective method of detoxification (see B. Fitch Haumann, *INFORM* 6:248-257 (1995)), such a methodology during processing is particularly critical where transgenic detoxification is not applicable.

In one embodiment of the present invention, fumonisin degradative enzymes are presented to grain, plant material for silage, or a contaminated food crop, or during the processing procedure, at the appropriate stages of the procedure and in amounts effective for detoxification of fumonisins and structurally related mycotoxins. Detoxification by the enzymes, microbial strains, or an engineered microorganism can occur not only during the processing, but also any time prior or during the feeding of the grain or plant material to an animal or incorporation of the grain or food crop into a human food product, or before or during ingestion of the food crop.

Another embodiment of the present invention is the engineering of a bacterium or fungus to express the detoxification enzymes and then using the bacterium or fungus rather than the enzyme itself. There are a number of microbes that could be engineered to express the polynucleotides of the present invention. One could also activate, either inducibly or constitutively, the endogenous genes for fumonisin esterase or APAO. By overexpressing the degradative enzymes and then treating plants, seed, or silage with the microorganism, it would be possible to degrade fumonisin *in situ*.

The polynucleotides of the invention can be introduced into microorganisms that multiply on plants (epiphytes) to deliver enzymes to potential target crops. Epiphytes can be gram-positive or gram-negative bacteria, for example.

The microorganisms that have been genetically altered to contain at least one degradative polynucleotide and resulting polypeptide may be used for protecting agricultural crops and products. In one aspect of the invention, whole, i.e. unlysed, cells of the transformed organism are treated with reagents that prolong the activity of the enzyme produced in the cell when the cell is applied to the environment of a target plant. A secretion leader may be used in combination with the gene of interest such that the resulting enzyme is secreted outside the host cell for presentation to the target plant.

The degradative enzymes can be fermented in a bacterial host and the resulting bacteria processed and used as a microbial spray. Any suitable microorganism can be used

for this purpose. See, for example, Gaertner, *et al.* (1993) in *Advanced Engineered Pesticides*, (ed. Kim, Marcel Dekker, New York).

The enzymes or microorganisms can be introduced during processing in appropriate manners, for example as a wash or spray, or in dried or lyophilized form or
5 powered form, depending upon the nature of the milling process and/or the stage of processing at which the enzymatic treatment is carried out. See generally, Hosney, R.C., *Principles of Cereal Science and Technology*, American Assn. of Cereal Chemists, Inc., 1990 (especially Chapters 5, 6 and 7); Jones, J.M., *Food Safety*, Eagan Press, St. Paul, MN, 1992 (especially Chapters 7 and 9); and Jelen, P., *Introduction to Food Processing*,
10 Restan Publ. Co., Reston, VA, 1985. Processed grain or silage to be used for animal feed can be treated with an effective amount of the enzymes in the form of an inoculant or probiotic additive, for example, or in any form recognized by those skilled in the art for use in animal feed. The enzymes of the present invention are expected to be particularly useful in detoxification during processing and/or in animal feed prior to its use, since the
15 enzymes display relatively broad ranges of pH activity. The esterase from *Exophiala spinifera*, ATCC 74269, showed a range of activity from about pH 3 to about pH 6, and the esterase from the bacterium of ATCC 55552 showed a range of activity from about pH 6 to about pH 9 (US patent no. 5,716,820, *supra*). The APAO enzyme from *Exophiala spinifera* (ATCC 74269) has a pH range of activity from pH 6 to pH 9.

Genetic Engineering of Ruminant Microorganisms

Ruminant microorganisms can be genetically engineered to contain and express either the fumonisin esterase enzymes or APAO, or a combination of the enzymes. The genetic engineering of microorganisms is now an art recognized technique, and ruminant
25 microorganisms so engineered can be added to feed in any art recognized manner, for example as a probiotic or inoculant. In addition, microorganisms capable of functioning as bioreactors can be engineered so as to be capable of mass producing either the fumonisin esterases or the APAO enzyme.

Use of the Fumonisin Esterase and APAO Enzymes for Detection of Reagents for Fumonisin and Related Compounds

Another embodiment of the present invention is the use of the enzymes of the present invention as detection reagents for fumonisins and related compounds. The enzymes of the present invention can be used as detection reagents because of the high specificity of the esterase and deaminase enzymes, and the fact that hydrolysis followed by amine oxidation can be monitored by detection of hydrogen peroxide or ammonia using standard reagents (analogous to a glucose detection assay using glucose oxidase). Hydrogen peroxide is often measured by linking a hydrogen peroxide-dependent peroxidase reaction to a colored or otherwise detectable peroxidase product (e.g. Demmano, *et al.*, *European Journal of Biochemistry* 238(3): 785-789 (1996)). Ammonia can be measured using ion-specific electrodes : Fritsche, *et al.*, *Analytica Chimica Acta* 244(2): 179-182 (1991); West, *et al.*, *Analytical Chemistry* 64(5): 533-540 (1992), and all herein incorporated by reference) or by GC or other chromatographic method.

For example, recombinant or non-recombinant, active fumonisin esterase (ESP1 or BEST) and APAO proteins are added in catalytic amounts to a sample tube containing an unknown amount of fumonisins (FB1, FB2, FB3, FB4, or partial or complete hydrolysis products of these). The tube is incubated under pH and temperature conditions sufficient to convert any fumonisin in the sample to AP1 or to 2-oxo-FB1, and correspondingly the AP1 to 2-OP, ammonia, and hydrogen peroxide. Alternatively, APAO or trAPAO is added in catalytic amounts to a sample tube containing an unknown amount of fumonisins (FB1, FB2, FB3, FB4, or partial or complete hydrolysis products of these). The tube is incubated under pH and temperature conditions sufficient to convert any fumonisin in the sample to 2-oxo FB1, ammonia, and hydrogen peroxide. Then suitable reagents are added for quantification of the hydrogen peroxide or ammonia that were generated stoichiometrically from fumonisins. By comparison with control tubes that received no esterase or APAO enzyme, the amount of fumonisin present can be calculated in direct molar proportion to the hydrogen peroxide or ammonia detected, relative to a standard curve.

This invention can be better understood by reference to the following non-limiting examples. It will be appreciated by those skilled in the art that other embodiments of the invention may be practiced without departing from the spirit and the scope of the invention as herein disclosed and claimed.

Example 1

Fungal and bacterial isolates. *Exophiala* isolates from maize were isolated as described in US patent no. 5,716,820, issued February 10, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997, and herein incorporated by reference.

Isolation methods. Direct isolation of black yeasts from seed was accomplished by plating 100 microliters of seed wash fluid onto YPD or Sabouraud agar augmented with cycloheximide (500 mg/liter) and chloramphenicol (50 mg/liter). Plates were incubated at room temperature for 7-14 days, and individual pigmented colonies that arose were counted and cultured for analysis of fumonisin-degrading ability as described in US patent no. 5,716,820, issued February 10, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997.

Analysis of fumonisins and metabolism products. Analytical thin-layer chromatography was carried out on 100% silanized C18 silica plates (Sigma #T-7020; 10 x 10 cm; 0.1 mm thick) by a modification of the published method of Rottinghaus (Rottinghaus, *et al.*, *J Vet Diagn Invest*, 4: 326 (1992), and herein incorporated by reference).

To analyze fumonisin esterase activity sample lanes were pre-wet with methanol to facilitate sample application. After application of from 0.1 to 2 μ l of aqueous sample, the plates were air-dried and developed in MeOH:4% KCl (3:2) or MeOH:0.2 M KOH (3:2) and then sprayed successively with 0.1 M sodium borate (pH 9.5) and fluorescamine (0.4 mg/ml in acetonitrile). Plates were air-dried and viewed under long wave UV.

For analysis of APAO activity, an alternative method was used. Equal volumes of sample and 14 C-API (1 mg/ml, pH 8, 50 mM sodium phosphate) were incubated at room temperature for one to six days. Analytical thin-layer chromatography was then carried out on C60 HPK silica gel plates (Whatman #4807-700; 10x10 cm; 0.2 mm thick). After application of from 0.1 to 2 μ l of aqueous sample, the plates were air-dried and developed in CHCl_3 :MeOH: CH_3COOH : H_2O (55:36:8:1). Plates were then air dried, and exposed to PhosphorImager screen (Molecular Dynamics) or autoradiographic film. A StormTM PhosphorImager (Molecular Dynamics) was used to scan the image produced on the screen.

Alkaline hydrolysis of FB1 to AP1. FB1 or crude fumonisin C₈ material was suspended in water at 10-100 mg/ml and added to an equal volume of 4 N NaOH in a screw-cap tube. The tube was sealed and incubated at 60°C for 1 hr. The hydrolysate was cooled to RT and mixed with an equal volume of ethyl acetate, centrifuged at 1000 RCF for 5 minute and the organic (upper) layer recovered. The pooled ethyl acetate layers from two successive extractions were dried under N₂ and resuspended in distilled H₂O. The resulting material (the aminopentol of FB1 or “AP1”) was analyzed by TLC.

Enzyme activity of culture filtrate and mycelium. *Exophiala spinifera* isolate 2141.10 was grown on YPD agar for 1 week, and conidia were harvested, suspended in sterile water, and used at 10⁵ conidia per ml to inoculate sterile Fries mineral salts medium containing 1 mg/ml purified FB1 (Sigma Chemical Co.). After 2 weeks incubation at 28° C in the dark, cultures were filtered through 0.45 micron cellulose acetate filters, and rinsed with Fries mineral salts. Fungal mycelium was suspended in 15 mL of 0.1% FB1, pH 5.2 + 1 mM EDTA + 3 µg/mL Pepstatin A + 1.5 µg/mL Leupeptin and disrupted in a Bead Beater™ using 0.1 mm beads and one minute pulses, with ice cooling. Hyphal pieces were collected by filtering through Spin X™ (0.22 µm), and both mycelial supernatant and original culture filtrates were assayed for fumonisin modification by methods outlined above.

Preparation of crude culture filtrate. Agar cultures grown as above were used to inoculate YPD broth cultures (500 ml) in conical flasks at a final concentration of 10⁵ conidia per ml culture. Cultures were incubated 5 days at 28°C without agitation and mycelia harvested by filtration through 0.45 micron filters under vacuum. The filtrate was discarded and the mycelial mat was washed and resuspended in sterile carbon-free, low mineral salts medium (1 g/liter NH₃NO₄; 1 g/liter NaH₂PO₄; 0.5 g/liter MgCl₂; 0.1 g/liter NaCl; 0.13 g/liter CaCl₂; 0.02 g/liter FeSO₄ · 7H₂O, pH 4.5) containing 0.5 mg/ml alkaline hydrolyzed crude FB1. After 3-5 days at 28°C in the dark with no agitation the cultures were filtered through low protein binding 0.45 micron filters to recover the culture filtrate. Phenylmethyl sulfonyl fluoride (PMSF) was added to a concentration of 2.5 mM and the culture filtrate was concentrated using an Amicon™ YM10 membrane in a stirred cell at room temperature, and resuspended in 50 mM sodium acetate, pH 5.2 containing 10 mM CaCl₂. The crude culture filtrate (approx. 200-fold concentrated) was stored at -20°C.

To obtain preparative amounts of enzyme-hydrolyzed fumonisin, 10 mg. of FB1 (Sigma) was dissolved in 20 mL of 50 mM sodium acetate at pH 5.2 + 10 mM CaCl₂, and 0.25 mL of 200x concentrated crude culture filtrate of 2141.10 was added. The solution was incubated at 37°C for 14 hours, and then cooled to room temperature. The reaction mixture was brought to approx. pH 9.5 by addition of 0.4 mL of 4 N KOH, and the mixture was extracted twice with 10 mL ethyl acetate. The combined organic layers were dried under N₂ and resuspended in dH₂O. 2.5 milligrams of organic extracted material were analyzed by Fast Atom Bombardment (FAB) mass spectrometry. The resulting mass spectrum showed a major ion at M/z (+1)=406 mass units, indicating the major product of enzymatic hydrolysis was AP1, which has a calculated molecular weight of 405.

Example 2

Preparation of AP1-induced and non-induced mycelium.

Liquid cultures of *Exophiala spinifera* isolate 2141.10 were prepared from YPD agar plates (Yeast Extract 10 gm, Bacto-Peptone 20 gm, Dextrose 0.5 gm, and Bacto-Agar 15 gm per liter of water). Aliquots (400-500 uL) of a water suspension of *E. spinifera* cells from YPD agar were spread uniformly onto 150 x 15 mm YPD agar plates with 4 mm sterile glass beads. The plates were incubated at room temperature for 6-7 days. The mycelia/conidia were transferred from the agar plates into Mineral Salts Medium (MSM) (Na₂HPO₄·7H₂O 0.2 gm, NH₄Cl 1.0 gm, CaCl₂·2H₂O 0.01 gm, FeSO₄·7H₂O 0.02 gm per liter of distilled water, pH 4.5) and centrifuged at 5000 x g, 4°C, 20 minutes to pellet the cells. The cell pellet was rinsed once in 40 ml MSM and recentrifuged. The rinsed cell pellet was used to inoculate MSM at a 1:19 ratio of packed cells: MSM. The culture to be induced was supplemented with AP1 to a final concentration of 0.5-1.0 mg/ml and incubated at 28°C, 100 rpm, in the dark to induce catabolic enzymes. The non-induced cultures did not receive AP1 but were grown on media containing 4-ABA at the same concentration as AP1. The supernatants were removed by filtration through 0.45 cellulose acetate. The remaining mycelial mat was washed with sterile MSM and then frozen in liquid nitrogen for storage.

Example 3

Effect of FB1 and AP1 on maize coleoptiles

Maize coleoptiles from 4 day dark-grown germinated maize seeds were excised above the growing point and placed in 96-well microtiter plates in the presence of 60 microliters of sterile distilled water containing FB1 or AP1 at approximately equimolar concentrations of 1.5, .5, .15, .05, .015, .005, .0015, or .0005 millimolar, along with water controls. After 2 days in the dark at 28° C the coleoptiles were placed in the light and incubated another 3 days. Injury or lack thereof was evaluated as follows:

	0	.0005	.0015	.005	.015	.05	.15	.5	1.5	mM
FB1	-	-	-	-	+/-	+	+	+	+	
AP1	-	-	-	-	-	-	-	-	+	

+ = brown necrotic discoloration of coleoptile

- = no symptoms (same as water control)

The results (see table above) indicate there is at least a 30-fold difference in toxicity between FB1 and AP1 to maize coleoptiles of this genotype. This is in general agreement with other studies where the toxicity of the two compounds was compared for plant tissues: In *Lemna* tissues, AP1 was approx. 40-fold less toxic (Vesonder *et al.*, "Arch Environ Contam Toxicol 23: 464-467 (1992).). Studies with both AAL toxin and FB1 in tomato also indicate the hydrolyzed version of the molecule is much less toxic (Gilchrist *et al.*, *Mycopathologia* 117: 57-64 (1992)). Lamprecht *et al.* also observed an approximate 100-fold reduction in toxicity to tomato by AP1 versus FB1 (Lamprecht *et al.*, *Phytopathology* 84: 383391 (1994))

Example 4

Effect of FB1 and AP1 on maize tissue cultured cells (Black Mexican Sweet, BMS)

FB1 or AP1 at various concentrations was added to suspensions of BMS cells growing in liquid culture medium in 96-well polystyrene plates. After 1 week the cell density in wells was observed under low power magnification and growth of toxin-treated wells was compared to control wells that received water. Growth of BMS cells was significantly inhibited at 0.4 micromolar FB1, but no inhibition was observed until 40 micromolar AP1. This represents an approximate 100-fold difference in toxicity to maize tissue cultured cells. Similarly Van Asch *et al.* (VanAsch *et al.*, *Phytopathology* 82: 1330-

1332 (1992)) observed significant inhibition of maize callus grown on solid medium at 1.4 micromolar FB1. AP1 was not tested in that study, however.

Example 5

APAO Activity

A cell-free extract that contains the deaminase activity was obtained by subjecting substrate-induced *Exophiala spinifera* cells to disruption using a Bead Beater™ in 50 mM Na-phosphate, pH 8.0, and recovering the cell-free supernatant by centrifugation and .45 micron filtration. Catabolic activity is assayed by incubating extracts with AP1 (hydrolyzed fumonisin B1 backbone) or ¹⁴C-labelled AP1 with the extract and evaluating by TLC on C18 or C60 silica. The product 2-OP has a lower R_f than AP1 and is detected either by radiolabel scan or by H₂SO₄ spray/charring of the TLC plate. 2-OP does not react with the amine reagent, fluorescamine, that is routinely used to detect AP1 on TLC plates, suggesting that the amine group is missing or chemically modified. Activity is greater at 37°C than at room temperature, but following 30 min. at 65°C or 100°C (no AP1 catabolic activity remained). Activity is maximal at pH 9. At pH 9, complete conversion to 2-OP occurred in 30 minutes. Activity is retained by 30,000 dalton molecular weight cutoff membrane, but only partially retained by 100,000 dalton molecular weight cutoff membrane. Other amine-containing substrates were tested for modification by the crude extract. Fumonisin, with tricarballic acids attached, is not modified by the extract, indicating that ester-hydrolysis must occur first for the APAO to be able to be effective in modifying FB1 (as noted below, the *E. coli*-expressed, recombinant APAO enzyme does in fact oxidize FB1 although at a lower rate than AP1). Other long-chain bases (sphingosine, sphinganine, and phytosphingosine) are apparently not modified by the crude APAO, suggesting the enzyme(s) is specific for the fumonisin backbone. Preparative amounts of the product, named 2-OP, have also been purified and analyzed by C13 nmr. The results indicate that 2-OP has a keto group at carbon 2 instead of an amine, consistent with an oxidative deamination by an amine oxidase. The C13 nmr data also indicate that 2-OP spontaneously forms an internal hemiketal between C-1 and C-5, resulting in a 5-membered ring with a new chiral center at C-2. All other carbon assignments are as in AP1, thus 2-OP is a compound of composition C₂₂H₄₄O₆, FW 404. The product of the

enzyme acting on hydrolyzed fumonisins would not be expected to display any significant toxicity.

Other enzymes were tested for their ability to modify AP1. All enzymes were assayed by radiolabeled TLC, as described above, under optimal conditions at 37° Celsius, overnight or longer. The results are as follows:

Deaminating	EC	Source	Result
Monoamine Oxidase	1.4.3.4	bovine plasma	negative
D-amino oxidase	1.4.3.3	porcine kidney; TypeX	negative
L-amino oxidase	1.4.3.2	C.adamanteus venom; TypeI	negative
Tyramine oxidase	1.4.3.4	Arthrobacter spp	negative
Methylamine dehydrogenase	1.4.99.3	Paracoccus denitrificans	negative
Alkyl amine dehydrogenase	1.4.99.4	Alcaligenes faecalis	negative
Phenylalanine ammonia lyase	4.3.1.5	Rhodotorula glutinis; TypeI	negative
Histidine ammonia lyase	4.3.1.3	Pseudomonas fluorescens	negative
L-aspartase	4.3.1.1	Hafnia alvei (Bacterium cadaveris)	negative
Tyrosine oxidase	1.14.18.1	mushroom	negative
Lysine oxidase	1.4.3.14	Trichoderma viride	negative
Diamine oxidase	1.4.3.6	porcine kidney	negative

The results were negative for each enzyme tested. Therefore isolates from the American Type Culture Collection (ATCC) were collected. The ATCC isolates selected were listed as containing amine-modifying enzymes or were capable of growth/utilization on amine-containing substrates. The isolates were tested to determine if they could grow on or utilize AP1 as the sole carbon source and if any could modify AP1 to a new compound(s). The nitrogen sources that were used in liquid cultures were AP1 0.1% (w/v), s-butylamine 0.1% (v/v), n-butylamine 0.1% (v/v), and ammonium nitrate 0.2% (w/v). These were prepared in Vogel's Minimal Media (without NH₄NO₃) containing 2% sucrose. The isolates were inoculated into the various media and monitored for growth over 2-3 weeks. They were also assayed with the ¹⁴C-radiolabeled TLC assay for AP1 modification. In summary, none of the isolates tested exhibited modification of AP1 *in vivo*. Clearly the APAO enzyme is unique and unusual in its ability to modify the AP1 toxin.

EXAMPLE 6

Isolation of the trAPAO Polynucleotide

The trAPAO polynucleotide was identified using a proprietary transcript imaging method that compares transcript patterns in two samples and allows cloning of differentially expressed fragments. This technology was developed by CuraGen® (New

Haven, CT). (see Published PCT patent application no. WO 97/15690, published May 1, 1997, and hereby incorporated by reference) Fluorescently-tagged, PCR amplified cDNA fragments representing expressed transcripts can be visualized as bands or peaks on a gel tracing, and the cDNA from differentially expressed (induced or suppressed) bands can be recovered from a duplicate gel, cloned and sequenced. Known cDNAs can be identified without the need for cloning, by matching the predicted size and partially known sequence of specific bands on the tracing.

In the present invention two RNA samples were obtained from cultures of *E. spinifera* grown for a specified period in a mineral salts medium containing either AP1 (induced condition), or gamma-aminobutyric acid (ABA; non-induced condition) as a sole carbon source. In the induced condition, fumonisin esterase and APAO enzyme activities are detected, whereas in the non-induced condition these activities are not detected. The methods used for induction of APAO and detection of activity are described earlier (see Example 2 and Example 5). RNA was extracted from induced mycelium by Tri-Reagent methods (Molecular Research Center Inc., Cincinnati, Ohio) only grinding a frozen slurry of tissue and Tri-Reagent with a mortar and pestle until almost melted and adding an additional extraction after the phase separation by extracting the aqueous phase one time with phenol, and two times with a phenol:chloroform:isoamyl alcohol mixture. The RNA's were submitted for CuraGen® transcript imaging to detect cDNA fragments that are induced specifically in the presence AP1. In the resulting gel tracing several bands were found which showed induction of at least 2-fold and up to 79-fold or even 100-fold or more in AP1. In the resulting gel tracing several bands were found which showed induction of at least 10-fold in AP1-grown cells as compared to cells grown in ABA. The sequence of two highly induced bands can be found in Table 1.

TABLE 1

Nucleotide sequence of two CuraGen® bands that were identified as strongly induced by AP1 in cultures of *Exophiala spinifera*.

>k0n0-395.5_b (SEQ ID NO: 1)
GGGCCCCGGCGTTCTCGTAGGCTGCGCGGAGTTGGTCCCAGACAGACTTTTGTTCGTACCTGCTTG
GACTGTTGGGACCACTTCCGTCCCGGGTCTCCGACCATGAAACAGGTAATGGACCATTGTTCGAT
CGACGTCGATGCTGGTATCTCTGGCAAATGAGATGGGGTCACAGCTCGATTGGAGGACGCCCCGA
GAAGCCTTGTTTCGCGCCACCACGGCTTGTCCTATACGAAGACTATCTTGCTATAGTAGCCAGG
ATAGAATTTTCCGCCAATGCTTGCTTCTCGGCGGGAAGAGGTGGTGAAAATGTCAAGGTGGGAT
ACAAGGTTGTTCGGTAACGAAACCANCACCTTTTGGCTTCGGAACACGGCGC

>r0c0-182.3_6 (SEQ ID NO: 2)
GAATTTTCCGCCAATGCTTGCTTCTCGGCGGGAAGAGGTGGTGAAAATGTCAAGGTGGGATACA
AGGTTGTTCGGTAACGAAACCACCACCTTTTTGCTTCGGAACACGGCGCCCGAGGCCGATCGTAC
TGTACAGCCGGATGCCGACTGCTCAATTCAGCGACGGGGGTGTTGAGGTGCAC

5

Two of the highly induced bands, k0n0-395.5, and r0c0-182.3 showed significant sequence homology to a family of enzymes, flavin-containing amine oxidases (EC 1.4.3.4), that oxidize primary amines to an aldehyde or ketone, releasing ammonia and hydrogen peroxide (Table 2).

10

TABLE 2

15 **Identification of a putative flavin amine oxidase from *E. spinifera*: AP1-induced transcript fragments with amine oxidase homology. BLAST 2.0 default parameters.**

Clone ID	Size	Best Hit	Best Hit Name, source	Prob	from	to	Likely function
k0n0-395.5	395 bp	P40974	putrescine oxidase, <i>Micrococcus rubens</i> , EC 1.4.3.10 Length = 478	8.0 e-07	276	333	oxidation of C-2 amine of AP1
r0c0-182.3 (contigs with k0n0-395)	182 bp	P12398	monoamine oxidase type A (MAO-A) [<i>Bos taurus</i>] Length = 527	0.0039	238	296	oxidation of C-2 amine of AP1

20 The chemical structure of the primary product of AP1 deamination is thought to be a 2-keto compound which cyclizes to a hemiketal at carbons 2 and 5. Therefore it is predicted that this induced enzyme is responsible for deamination of AP1.

Using sequence derived from k0n0-395.5, a partial cDNA was obtained by 3' and 5' RACE-PCR (Chenchik, *et al.*, *CLONTECHniques* X 1:5-8 (1995); Chenchik, *et al.*, A new method for full-length cDNA cloning by PCR. In *A Laboratory Guide to RNA: Isolation, Analysis, and Synthesis*. Ed. Krieg, P.A. (Wiley-Liss, Inc.), 273-321 (1996)). A RACE cloning kit from CLONTECH was used, to obtain the RACE amplicons. Briefly, poly A+ RNA is transcribed to make first strand cDNA using a "lock-docking" poly T, cDNA synthesis primer, the second strand is synthesized and the

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Marathon cDNA adaptor is ligated to both ends of the ds cDNA. Diluted template is then used with the Marathon adapter primer and in separate reactions either a 5' Gene Specific Primer (GSP) or a 3'GSP is used to produce the 3' or 5' RACE amplicon. After characterization of the RACE product(s) and sequencing, full-length cDNAs may be generated by 1) end-to-end PCR using distal 5' and 3' GSPs with the adapter-ligated ds cDNA as template, or 2) the cloned 5' and 3'-RACE fragments may be digested with a restriction enzyme that cuts uniquely in the region of overlap, the fragments isolated and ligated. Subsequently, the RACE-generated full-length cDNAs from 1) and 2) may be cloned into a suitable vector.

In combination with the supplied adapter primer the following gene specific primers were used: for 3' RACE the oligonucleotide N21965: 5'-TGGTTTCGTTACCGACAACCTTGTATCCC-3' (SEQ ID NO: 3) and for 5' race, the oligonucleotide N21968: 5'-GAGTTGGTCCCAGACAGACTTTTGTCGT-3' (SEQ ID NO: 4). The polynucleotide sequence of the trAPAO polynucleotide, k0n0-395_6.5, from *Exophiala spinifera* is shown in SEQ ID NO: 5. The polypeptide sequence of trAPAO is shown in SEQ ID NO: 6.

A second clone of APAO containing an unspliced intron was also found. The polynucleotide sequence of trAPAO-I polynucleotide, k0n0-395_5.4, the intron containing clone, from *Exophiala spinifera*, can be found in SEQ ID NO: 7. The polypeptide sequence of trAPAO-I with the intron spliced out is shown in SEQ ID NO: 8. The polypeptide sequence of trAPAO-I without the intron spliced out is shown in SEQ ID NO: 9.

EXAMPLE 7

Heterologous Expression of trAPAO

Protein alignments generated with PileUp (GCG) indicate that k0n0-395_6.5 (trAPAO) is similar in size to other flavin amine oxidases and is close to being full length with respect to the amino terminus of their class of proteins. The k0n0-395_6.5 sequence contains a complete β - α - β fold that is required for dinucleotide (FAD) binding, close to the amino end. The k0n0-395 sequence appears to lack only a variable amino terminal segment that varies in length from 5 amino acids in rat monoamine oxidases A & B to 40 amino acids in length in *Aspergillus* MAO-N. The function of these amino terminal extensions is not known; they are not recognizable as secretion signals. Based on the likely

localization of the *Exophiala* APAO outside the cell membrane, the prediction is that k0n0-395 would have a signal sequence similar to that of the fumonisin esterase cloned from the same organism (US patent no. 5,716,820, *supra*). Using GenomeWalker™, it is possible to clone the 5' end of the transcript and upstream genomic regulatory elements.

5 However, the signal sequence is not expected to be critical to the functionality of the enzyme; in fact, the preferred strategy for heterologous expression in maize and *Pichia pastoris* involves replacing the endogenous signal sequence (if present) with an optimized signal sequence for the organism, e.g. barley alpha amylase for maize and the yeast alpha factor secretion signal for *Pichia*. In maize transformed with fumonisin esterase, the
10 barley alpha amylase signal sequence gave higher amounts of functional protein than the native fungal signal, therefore replacement of the native fungal signal sequence is a logical optimization step. Since many of the amine oxidases have a positively charged amino acid near the N-terminus and upstream of the dinucleotide binding site, an additional optimization step included adding a codon for the lysine (K) to the N-terminus of the
15 trAPAO clone (k0n0-395_6.5, SEQ ID NO: 5). This clone is designated K:trAPAO and can be seen in SEQ ID NOS: 10 and 11. The extra lysine is at amino acid 1 and nucleotides 1-3.

EXAMPLE 8

Pichia Expression of trAPAO

20 For optimum expression of trAPAO in *Pichia pastoris* the alpha mating factor signal peptide was operably linked in-frame with K:trAPAO coding sequence and can be seen in SEQ ID NOS: 16 and 17. The nucleotide sequence of clone pPicZalphaA:K:trAPAO contains a PCR-amplified insert comprising the k0n0-395 open
25 reading frame with an additional lysine residue at the amino terminus, with a 5' EcoRI site and 3' NotI site for in-frame cloning into the alpha factor secretion vector pPicZalphaA. Nucleotides 1-267 contain the yeast α mating factor secretion signal. The amino acid sequence, shown in SEQ ID NO: 17, contains the trAPAO polypeptide produced from pPicZalphaA:K:trAPAO following transformation into *Pichia pastoris*.

30 For cloning into expression vectors, two cloning strategies were used. The cDNA k0n0-395_5.4 was generated by using end-to-end PCR using distal 5' and 3' GSPs with the adapter-ligated double stranded cDNA as a template. Each oligonucleotide primer was

designed with 5' restriction enzyme sites that contain a 23-25 bp of anchored gene sequence. The 3' primer also included the stop codon. The primer sequences are N23256: 5'-ggggaattcAAAGACAACGTTGCGGACGTGGTAG-3' (SEQ ID NO: 12) and N23259: 5'-ggggcgccgcCTATGCTGCTGGCACCAGGCTAG-3' (SEQ ID NO: 13). A second method was used to generate k0n0-395_6.5. 5' RACE and 3' RACE products using a distal primer containing the necessary restriction enzyme sites, stop codon, etc as described above and paired with a "medial" GSP. The "medial primers" N21965: 5'-TGGTTTCGTTACCGACAACCTTGTATCCC-3' (SEQ ID NO: 14) for 3' RACE and for 5' race, the oligonucleotide N21968: 5'-GAGTTGGTCCCAGACAGACTTTTGTCGT-3' (SEQ ID NO: 15). Adapter-ligated double stranded cDNA was used as template. The isolated 5' and 3'-RACE fragments were digested with a restriction enzyme that cuts uniquely in the region of overlap, in this case Bgl I, isolated and ligated into the expression vector. The digestible restriction sites allow cloning of the inserts in-frame into EcoRI/NotI digested pPicZalphaA. pPicZalphaA is an *E. coli* compatible *Pichia* expression vector containing a functional yeast alpha factor secretion signal and peptide processing sites, allowing high efficiency, inducible secretion into the culture medium of *Pichia*. The resulting 1.4 kb bands were cloned into EcoRI/NotI digested pPicZalphaA plasmid.

SEQ ID NO: 16 contains the polynucleotide sequence of clone pPicZalphaA:K:trAPAO, a PCR-amplified insert that comprises the k0n0-395 open reading frame with an additional lysine residue at the amino terminus, and a 5' EcoRI site and 3' NotI site for in-frame cloning into the alpha factor secretion vector pPicZalphaA. SEQ ID NO: 17 contains the amino acid sequence of the trAPAO polypeptide produced from pPicZalphaA:K:trAPAO following transformation into *Pichia pastoris*. The alpha factor secretion signal and a lysine are added.

Pichia was transformed as described in Invitrogen Manual, Easy Select™ *Pichia* Expression Kit, Version B, #161219, with the trAPAO polynucleotide as described above with either an intron (trAPAO-I, negative control, no expression of active trAPAO since *Pichia* does not splice introns very efficiently) or without an intron (capable of making an active APAO protein). The *Pichia* culture fluids and pellets were assayed for APAO activity as described earlier.

The set of frozen six day *Pichia* culture cell pellets contained two samples with intron (SEQ ID NO: 7) in gene construct, # 11, # 14, and two samples without intron in

gene construct (SEQ ID NO: 5), #6, # 52. The six day culture fluids from the same cultures were used to spike with crude fungal enzyme for positive controls.

The 50 µl cell pellets were resuspended in 150 µl cold 50mM Na-phosphate, pH 8.0, and divided into two fresh 500 µl tubes. One tube was kept on ice with no treatment, the pellet suspension, and one tube was used for lysis. An equal volume of 0.1 mm zirconia-silica beads was added to each tube. The tubes were BeadBeat™ for 15 seconds then cooled on ice 5 minutes. This was repeated three times. The crude lysate was then transferred to another tube for assay or lysate suspension.

The TLC assays were performed as follows, the samples are 1) pellet suspensions; 10 µl; 2) lysate suspensions; 10 µl; 3) media controls-mixed 5 µl media with 5 µl crude fungal enzyme; 10 µl; 4) positive control-used crude fungal enzyme undiluted; 10 µl; 5) substrate control-used 50mM Na-phosphate, pH8.0; 10 µl. Ten microliters of each sample plus 10 µl of ¹⁴C-AP1 (1 mg/ml, 50 mM Na-phosphate, pH 8) was incubated at room temperature for 6 days. One microliter of the sample was spotted onto C18 and C60 TLC plates. The C18 plates were developed in MeOH:4% KCl (3:2). The C60 plates were developed in CHCl₃:MeOH:CH₃COOH:H₂O (55:36:8:1). The plates were then air dried and then exposed to a PhosphorScreen™ for 2-3 days. A Storm™ PhosphorImager was used to develop the images.

A positive TLC result is obtained if an additional radioactive spot appears at a lower R_f of the produced AP1 modification earlier identified as 2-OP, a deaminated product of AP1. In samples # 6 and # 52 (without intron) the AP1-modifying enzyme activity (conversion of AP1 to 2-OP) was detected in pellet suspensions and pellet lysates, although the majority of activity was associated with the pellet suspensions. In samples #11 and #14 (with intron) a minimal amount of AP1-modifying enzyme activity was detectable in the pellet lysate of # 14 only, which indicates *Pichia* cannot process the intron efficiently.

This experiment verified APAO activity can be detected in *Pichia* transformants, which verifies that trAPAO as described functions correctly in degrading AP1. The activity is associated with cell suspensions, which show higher activity than pellet lysates. Pellet lysates may show less activity due to release of endogenous proteases during lysis of the cells.

EXAMPLE 9

Expression of trAPAO or APAO in *E. coli*

The vector for expressing K:trAPAO in *E. coli* is pGEX-4T-1. This vector is a prokaryotic glutathione S-transferase (GST) fusion vector for inducible, high-level intracellular expression of genes or gene fragments as fusions with *Schistosoma japonicum* GST. GST gene fusion vectors include the following features, a lac promoter for inducible, high-level expression; an internal lac Iq gene for use in any *E. coli* host; and the thrombin factor Xa or PreScission Protease recognition sites for cleaving the desired protein from the fusion product. The insert of interest, k0n0-395_6.5 (K:trAPAO) or APAO, was subcloned into the 5' EcoRI site and a 3' NotI site allowing in-frame expression of the GST:K:trAPAO or GST:APAO fusion peptide.

The polynucleotide sequence of the GST:K:trAPAO fusion can be found in SEQ ID NO: 18. The GST fusion with polylinker can be found at nucleotides 1 to 687. The K:trAPAO can be found at nucleotides 688 to 2076. The resulting polypeptide for the GST:K:trAPAO fusion can be seen at SEQ ID NO: 19. Amino acids 1 to 229 represent the GST fusion plus polylinker and amino acids 230 to 692 represent the K:trAPAO portion of the fusion.

E. coli was transformed with the pGEX-4T-1 vector containing K:trAPAO or APAO as described in BRL catalogue, Life Technologies, Inc. catalogue; Hanahan, D., *J. Mol. Biol.* 166:557 (1983) Jessee, J. *Focus* 6:4 (1984); King, P.V. and Blakesley, R., *Focus* 8:1, 1 (1986), and hereby incorporated by reference. The transformed *E. coli* was induced by addition of IPTG (isopropyl b-D-thiogalactopyranoside). Four samples of soluble extract and four samples of insoluble inclusion bodies were tested for trAPAO or APAO activity as described in Example 8. APAO activity was present in all soluble samples and two insoluble samples. Highest activity was found at 10 uM IPTG induction. Thus the pGEX-4T-1 vector containing K:trAPAO or APAO is capable of producing active APAO enzyme in *E. coli*.

EXAMPLE 10

The Complete Nucleotide Sequence of the *Exophiala* APAO Gene

Using Genome Walker, the complete nucleotide sequence of the *Exophiala* APAO gene was recovered. The nucleotide sequence described in SEQ ID NO: 5 is missing a

portion of the 5' end of the native gene. The missing portion of the 5' end of the native gene is not necessary for expression of an active APAO enzyme, as can be seen in Examples 8 and 9. The complete nucleotide sequence of APAO can be seen in SEQ ID NO: 22. The translation of SEQ ID NO: 22 can be found in SEQ ID NO: 23.

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EXAMPLE 11

Expression of APAO and ESP1 in transgenic maize callus

One of the preferred constructs for expression in maize is the nucleotide sequence of the trAPAO operably linked to the barley alpha amylase signal sequence. The nucleotide sequence of K:trAPAO translational fusion with barley alpha amylase signal sequence, for expression and secretion of the mature trAPAO in maize can be seen in SEQ ID NO: 20. Nucleotides 1-72, represent the barley alpha amylase signal sequence; nucleotides 73-75, represent the added lysine residue; and nucleotides 76 -1464 , represent the trAPAO cDNA. The amino acid sequence translation of SEQ ID NO: 20 can be found in SEQ ID NO: 21. Amino acids 1 to 24 represent the barley alpha amylase signal sequence and amino acids 25 to 463 is the sequence of K:trAPAO.

Maize embryos were transformed with linear DNA (insert, lacking a bacterial antibiotic resistance marker), derived from constructs containing three transcription units: 1) a PAT selectable marker gene (Wohlleben *et al.*, *Gene* 70, 25-37 (1988)), 2) fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence, and 3) full length APAO without or with an amino-terminal barley alpha amylase signal sequence, (P13603, comprising a PAT selectable marker operably linked to a 35S promoter, fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence and the ubiquitin promoter, and APAO operably linked to the ubiquitin promoter and P13611, comprising a PAT selectable marker operably linked to the 35S promoter, fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence and the ubiquitin promoter and APAO operably linked to a barley alpha amylase signal sequence and the ubiquitin promoter). In these constructs both ESP1 and APAO were linked to the maize ubiquitin promoter and first intron. In a third construct, the same three transcriptional units were cloned into an Agrobacterium T1 vector (P15258, the construct comprises a PAT selectable marker, fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence and APAO). Stably transformed callus or T0 plants regenerated from callus were tested for ESP1 and APAO activity in buffer extracts of leaf tissue, using

radiolabeled FB1 and/or AP1 and C18 thin-layer chromatography. Positive controls consist of non-transformed tissue spiked with *E coli*-expressed recombinant ESP1 or APAO. The results indicate that both ESP1 and APAO activities can be detected in transgenic maize callus and plants.

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Expression of ESP1 and APAO in transgenic callus

Construct	Sample ID Number	ESP1 activity (TLC)	APAO activity (TLC)
13603	3065.031-2	+	+
13603	3065.034-3	+	+
13603	3065.1117-3	+	+
13603	3065.11s7-n13	+	+
13603	3065.117-2	+	+
13603	3065.1115-2	+	+
13603	3065.1115-6	+	+
13603	3065.1112-1	+	+
13603	3065.118-6	+	+
13603	3065.11s3-1	+	+
13603	3065.11s1-13	+	+
13603	2805.762-2	+	+
13603	3065.1110-2	+	+
13603	3065.039-2	+	+
13611	3065.293-3	+	+
13611	3065.263-1	+	+
13611	3070.24.2.3	+	+

Transgenic plants were regenerated from the transgenic callus positive for both ESP1 and APAO activity by standard methods known in the art. Enzyme activity was tested as described previously. As can be seen below transgenic maize plants can successfully express both ESP1 and APAO enzymes.

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Expression of APAO and ESP1 in transgenic maize plants (T0)

Construct	Sample ID Number	ESP1 activity (TLC)	APAO activity (TLC)
13603	910080	+	+
13603	910081	+	+
13603	917065	+	+

Another preferred construct for expression of APAO in a plant is targeting the APAO to the peroxisome. Maize embryos were bombarded with insert containing APAO

operably linked to ubiquitin promoter and a peroxisomal targeting sequence (Gould, *et al.*, *J Cell Biol* 108:1657-1664 (1989)); ESP1 operably linked to ubiquitin promoter and the barley alpha amylase signal sequence; and a selectable marker of PAT operably linked to the 35S promoter (construct number I14952). Negative controls were unbombarded embryos/callus. Positive controls were unbombarded embryos/callus spiked with purified enzyme. Transformed callus was then tested for ESP1 or APAO activity as previously described. Out of 67 samples tested 18 samples contained both ESP1 activity and APAO activity. Peroxisomally targeted APAO and apoplast targeted fumonisin esterase can both be successfully expressed in a plant cell.

Another preferred construct for expression of APAO in a plant is targeting the APAO to the mitochondrial membrane. A C-terminal extension is required for targeting monoamine oxidases MAO-A and MAO-B to mammalian outer mitochondrial membranes. A MAO-A, MAO-B, or functionally similar C-terminal extension can be ligated in-frame to APAO or trAPAO to facilitate localization of this enzyme to the mitochondrial membrane of maize or other transformed species.

EXAMPLE 12

Comparison of APAO Sequence With Other Sequences

The *Exophiala* cDNA APAO (SEQ ID NO: 22) contains an 1800 bp open reading frame coding for a 600 amino acid polypeptide (SEQ ID NO: 23) with divergent homology to two classes of proteins. The carboxy three-fourths of APAO (amino acids 137 to 593) is strongly homologous to flavin amine oxidases, a group of enzymes catalyzing the oxidative deamination of primary amines at carbon 1. The amine oxidase function of the carboxy terminal domain was confirmed by expression of a truncated APAO polypeptide (from 137 to 600) in both *Pichia pastoris* and *E. coli*, using AP1 as a substrate (see Example 9). The amino terminal portion of APAO, in contrast, (from approx. 5 to 134) shows significant homology to a group of small deduced open reading frames (ORFs) reported in several bacteria and blue-green algae, as well as several higher organisms. These ORFs code for small proteins of unknown function, ranging in size from 14 to 17 kDA. The juxtaposition of these divergent homologies in a single polypeptide has not been reported previously.

Flavin amine oxidases (E.C. 4.1.4.3) are a group of flavoenzymes found in both higher and lower organisms, and serve a variety of functions in catabolism. They catalyze the oxidative deamination of primary amino groups located at the C-1 position of a variety of substrates, resulting in an aldehyde product plus ammonia and hydrogen peroxide. The

5 APAO enzymes of the present invention are the first flavin amine oxidase known to attack a primary amine not located at C-1 (i.e. C-2 of AP1) and resulting in a keto rather than aldehydic product. However, amino acid oxidases, while not closely related to flavin amine oxidases, are flavoenzymes that oxidize a C-2 amine adjacent to a C-1 carboxyl group.

10 The monoamine oxidases MAO A & B, (from human, bovine, and trout), are localized in the mitochondrial outer membrane of higher organisms and regulate the level of neurotransmitters. Microbial examples include a fungal amine oxidase (*Aspergillus niger* (*niger*) MAO-N) involved in amine catabolism, and a bacterial putrescine oxidase from a gram (+) bacterium (*Micrococcus rubens*.). The primary polypeptides vary in

15 length from 478 to 527 amino acids, and share regions of high amino acid sequence conservation at the 5' end as well as at various points through the coding region. Protein alignments generated with PileUp (GCG) indicate that trAPAO contains all conserved domains found in this class of proteins including those near the 5' end.

The amine oxidase domain of trAPAO contains several key features shared by this

20 class of enzymes, including an amino-terminal dinucleotide (ADP) binding region characterized by a beta-alpha-beta stretch containing three invariant glycines (G -X-G-X-X-G) in the beta-alpha turn. In trAPAO, this sequence is (DVVVVGAGLSG). This region is involved in FAD binding. Absent are several features unique to the mammalian amine oxidases, including several important cysteine residues (Wu *et al.*, *Mol Pharm*

25 43:888 (1993)), one of which (Cys-406 of MAO-A) is involved in covalent binding of FAD, and a carboxy-terminal extension that has been demonstrated to be involved in transporting to and anchoring the MAO in the outer mitochondrial membrane. The *Aspergillus* enzyme MAO-N has been demonstrated to contain non-covalent FAD, and also lacks the conserved cysteine. Therefore it is possible that the APAO enzyme has a

30 non-covalent FAD. The *Aspergillus* MAO-N has a carboxy-terminal tripeptide Ala-Arg-Leu that is involved in peroxisomal targeting and localization; this sequence is absent from *Exophiala* MAO.

The amine oxidase domain of trAPAO contains a total of seven cysteines, compared to ten for the *Aspergillus* enzyme and only two for the *Micrococcus* enzyme. The mammalian MAO enzymes contain variable numbers of cysteines (at least ten), some of which are highly conserved (including the FAD binding residue mentioned above). The trAPAO sequence also has two putative glycosylation sites (NDS, NQS) towards the amino end.

The purpose of the amino-terminal extension of APAO and the basis for its homology to a group of 14-17 kDa proteins is not clear. In *Synechocystis*, a similar polypeptide ORF is located immediately upstream of the NADP-dependent glutamine dehydrogenase (gdhA) and has been shown to be required for functional expression of gdhA (Chavez et al, 1995). However, in trAPAO the domain is clearly not necessary for enzymatic activity, as shown by the results of the expression experiments using the truncated APAO. An interesting clue comes from the frequent association of this small ORF with gene clusters involved in oxidoreductase activity in bacteria, or induced by heat stress in mice, suggesting a possible role in redox protection. A byproduct of amine oxidase activity is hydrogen peroxide. Flavoenzymes and other redox enzymes are often susceptible to inactivation by hydrogen peroxide (Schrader *et al.*, *App Microb Biotechnol* 45:458; Aguirre, *et al.*, *J Bacteriol* 171:6243 (1989)), and it is possible that this protein has a protective role against oxidants such as hydrogen peroxide. Alternatively, this domain could be involved in enzyme function, localization or association of the enzyme with other structures. No signal peptide region can be detected in this amino terminal region.

In multiple sequence alignment using GCG PileUp, trAPAO is most similar to putrescine oxidase of *Micrococcus rubens*, Swissprot accession number P40974, (30% identical amino acids, 40% similar). Homology with several mammalian monoamine oxidases A and B, Swissprot accession numbers P21397 (*Homo Sapiens* mao a), P19643 (*Rattus norvegicus* mao b), P21396 (*Rattus norvegicus* mao a), and P21398 (*Bos taurus* mao a), is somewhat less, ranging from 25 to 28% identity and 36 to 40% similarity. Homology to the only other fungal flavin amine oxidase known, MAO-N from *Aspergillus niger* (Swissprot accession number P46882), is somewhat lower (24% identical, 34% similar). The microbial enzymes are considerably divergent from each other, while the mammalian monoamine oxidases share 65 to 87% identity.

The amino terminal domain (ATD) of APAO also shows homology to a 14.5 kD protein from human and rat phagocytes that shows translational inhibition activity *in vitro* (Swissprot accession # P52758, P52759) Schmiedeknecht, *et al.*, *Eur J Biochem* 242 (2), 339-351 (1996)), and includes a heat-responsive protein from mouse (Samuel, *et al.*, *Hepatology* 25 (5), 1213-1222 (1997)). This suggests that this family of proteins is involved in regulating cellular metabolism. No example exists in which this domain is fused to a larger protein domain, however, making APAO unique. Without intending to be limited by theory, all of this suggests, that this domain plays a regulatory role in APAO gene expression, possibly to prevent translation of the message when it is not needed. This raises the question of how translation of the message is restored when active enzyme is required by the *Exophiala* cell. Possibly there are alternative start sites that begin downstream of the inhibitor domain; or proteolysis, complexing, degradation, or phosphorylation/ dephosphorylation of the inhibitor domain when it is not needed. The first possibility is less likely because there are no other ATG codons prior to the ATG at 122-124 that constitutes the predicted start site of APAO. The second possibility cannot be easily tested, although there is a casein kinase site in the ATD. Alternative roles for the ATD include oligomerization of the APAO protein, or anchoring the protein to some intracellular site, such as the membrane.

A parallel example of regulatory control over another flavoenzyme, human flavin monooxygenase 4 (FMO-4), by a C-terminal extension has been reported (Itagaki, *et al.*, *J of Biol Chem* 271(33): 20102-20107 (1996)). In this case the introduction of a stop codon prior to the 81 base C-terminal extension allowed expression of active enzyme in heterologous systems. The role of the C-terminal portion was not elucidated, however. In another example, alternative splicing led to a shorter gene product that complexed with and interfered with the function of the normally spliced version (Quinet, *et al.*, *J of Biol Chem* 268(23): 16891-16894 (1993)). In another case, an alternative splicing-generated insert in another protein led to inhibition of cell growth (Bhat, *et al.*, *Protein Engineering* 9(8): 713-718 (1996)). In yet another variation, fas/Apo1 splicing variants prevent apoptosis, apparently through a 49 amino acid domain shared by all variants ((Papoff, *et al.*, *J of Immunology* 156(12): 4622-4630 (1996)).

EXAMPLE 13

Making a Chimera Protein Containing Fumonisin Esterase and APAO activity in the Same Polypeptide.

5 The enzyme activities of fumonisin esterase and APAO can be combined in a single polypeptide by using the open reading frames together either with or without a spacer region between the two polypeptides. This creates a hybrid protein with dual enzyme activities that can be exported as a unit to the apoplast, and will allow both enzyme activities to be conveniently localized to the same area of the cell wall. The two cDNAs
10 can be combined in either order, but the preferred method is to link them in the order NH₃-Esterase:APAO-COOH. The spacer, if present, may consist of a short stretch of amino acids such as GGGSGGGS, or a set of amino acids that comprises a protease cleavage site that can be acted on by an apoplastic protease. This would result in the production of stoichiometric amounts of both esterase and APAO enzymes in the apoplast.
15 Alternatively, a polycistronic message could be engineered which is capable of direct translation of a downstream sequence, for example inclusion of an IRES sequence in the spacer region or a polynucleotide spacer region containing a polynucleotide cleavage site that can be recognized by RNase or is a self-cleaving ribozyme. The length of the splice site could be of any length that ensures proper translation of the polynucleotide.

20 The esterase-APAO ligated protein can be made with any fumonisin esterase, including but not limited to, the fumonisin esterase from *E. spinifera* (ESP1) or fumonisin esterase from bacterium (BEST1). Since the pH range for maximum activity of BEST1 is similar to that of APAO (range 6.0 to 8.0), these may present the most effective combination in fusion form. In addition, any of the polynucleotides of the present
25 invention, including APAO mutated to improve expression, may be used for an esterase-APAO ligation. As described in previous examples these fusion sequences can be placed in the appropriate expression vectors and used to express proteins in either bacteria or plants.

The nucleotide sequence of ESP1 contains three nucleotide differences and three
30 corresponding amino acid differences for the ESP1 sequence disclosed in pending US application no. 08/888,950, filed July 7, 1997 and US patent no. 6,025,188, issued February 15, 2000. Both the sequences disclosed in the present application and the sequences disclosed in the pending US applications contain functional fumonisin esterase

genes. For the purposes of the present invention, either the original ESP1 sequences or the ESP1 sequences may be used in combination with the APAO sequences or in fusion sequences. The nucleotide sequence of a BAA:ESP1:trAPAO construct for plant expression can be found in SEQ ID NO: 24 and the translation in SEQ ID NO: 25. The nucleotide sequence for a BAA:BEST1:K:trAPAO construct for plant expression can be found in SEQ ID NO: 26 and the translation in SEQ ID NO: 27. The nucleotide sequence of a GST:ESP1:K:trAPAO fusion for bacterial expression in a pGEX-4T-1 or similar vector can be found in SEQ ID NO: 28 and the translation in SEQ ID NO: 29. The nucleotide sequence for a GST:BEST1:K:trAPAO fusion for bacterial expression in a pGEX-4T-1 or similar vector can be seen in SEQ ID NO: 30 and the translation in SEQ ID NO: 31.

EXAMPLE 14

APAO Substrate Studies

The following assay was used to determine the substrate specificity of the APAO enzyme. Reaction mix: 436 μ l of 200 mM Na-phosphate, pH8.0; 50 μ l substrate (10 mM); 2 μ l Amplex Red (1 mg in 200 μ l DMSO); and 2 μ l of Peroxidase (5000 U/ml). The APAO enzyme was recombinant enzyme produced as GST fusion in *E. coli*, purified over a glutathione affinity column and cleaved with thrombin to remove the GST. All components were mixed at room temperature. The initial rate was determined in a spectrophotometer at 572 nm over one minute by absorbance units/second (BLANK). Ten microliters of APAO at 70 ug/ml was added and mixed. The initial rate was again determined at 572 nm over one minute in absorbance units/second (SAMPLE). The rates were converted to absorbance units/minute. The BLANK value was subtracted from the SAMPLE value. The absorbance units were converted to μ M H₂O₂ wherein 1 μ M H₂O₂ equals 0.138 absorbance units at pH 8.0.

SUBSTRATES FOR APAO

SUBSTRATE	RATE $\mu\text{M H}_2\text{O}_2/\text{min}$
1 mM Fumonisin B1	0.1429
1 mM AP1	0.8876
0.5mg/mL Fumonisin B2	0.3058
1 mM Fumonisin B3	0.1449
0.5mg/mL Fumonisin B4	0.1728
1 mM norepinephrine	0.0087
1 mM epinephrine	0.0071
1 mM dopamine	0.0040
1 mM spermine	0.0002

NOT SUBSTRATES FOR APAO (defined as compounds resulting in less than 1% conversion to hydrogen peroxide by APAO relative to AP1 under similar conditions of time, pH, temperature, and substrate concentration): 2-phenylethylamine, spermidine, EDTA- Na_2 , tryptamine, putrescine, benzamidine, serotonin, cadaverine, Pefabloc SC, tyramine, 1,3-diaminopropane, leupeptin, histamine, hydroxylamine, aprotinin, deprenyl, Fumonisin C4, isoniazid, sphingosine, phenelzine, sphinganine, phytosphingosine, D-alanine, DL-alanine, L-arginine, L-asparagine, L-aspartic acid, D-aspartic acid, L-cysteine, L-glutamine, L-glutamic acid, glycine, L-histidine, L-isoleucine, L-leucine, L-lysine, DL-lysine, L-methionine, DL-methionine, L-phenylalanine, L-proline, L-threonine, L-tryptophan, L-tyrosine, L-valine.

EXAMPLE 15

Sites on APAO for Possible Mutagenesis

Some cytosolic enzymes, when engineered for secretion by fusion with a heterologous signal peptide, lack function due to glycosylation at one or more potential glycosylation sites (amino acid consensus sequence N-X-S/T) that are not normally glycosylated in the native environment (Farrell *et al.*, *Plant Mol Biol* 15(6):821-5 (1990)). Since APAO lacks a recognizable signal sequence, it may be cytoplasmically localized in *Exophiala spinifera*, although secretion by some other method not involving a signal peptide cannot be ruled out. APAO contains two potential glycosylation sites, which may be glycosylated when APAO is secreted in a plant or other eukaryotic cell. Other modifications to APAO can be made to improve its expression in a plant system, including

site-directed mutagenesis to remove selected cysteine residues, which may be detrimental to proper folding when the protein is secreted into the endomembrane system for delivery to the apoplast.

Knowledge of the 3-dimensional structure of APAO would help to evaluate the likelihood that particular amino acids could contribute to misfolding, and increase the odds of making rational changes in the APAO sequence for successful secretion. To this end a 3-dimensional model of APAO was developed based on the crystal structure of a related amine oxidase from maize, maize polyamine oxidase or MPAO (Binda *et al.*, *Structure* 7:265-276 (1999)). The model was derived by automated modeling using the program *Modeler* (Molecular Simulations, Inc., San Diego, CA) and the resulting 3-D structure showed excellent fit based on an RMS deviation of 0.68 Å for the backbone coordinates of the two structures. The 3-D model of APAO based on MPAO is shown in Figures 1 and 2. Some of the possible mutations of APAO, which would result in removal of glycosylation sites or removal of cysteine residues can be seen below and in Figure 1.

Table of site-directed mutagenesis vectors and enzyme assay results.

Residue number	1	2	3	4	5	6	7	8	9	Glyc Site 1a,b	Glyc Site 2a,b	<i>E. coli</i> expression vector, APAO or trAPAO activity		Maize expression vector, APAO or trAPAO Activity	
Residue position in APAO or trAPAO	C64	C109	C167	C292	C351	C359	C387	C461	C482	N201	S203	N204	S206		
Construct	Amino acid substitution											Plasmid	Act	Plasmid	Act
Wild type APAO-1												PHP13367	+	PHP	-
Glyc(-) 1a2a APAO									A		A	PHP16284	-	n/a	
Glyc(-) 1a2b APAO									A		A	PHP16285	-	n/a	
Glyc(-) 1b2a APAO										A	A	PHP16286	-	n/a	
Glyc(-) 1b2b APAO										A	A	PHP16287	-	n/a	
Glyc(-)2a N204A APAO											A	PHP16589	+/-	n/a	
Glyc(-)2b S206A APAO											A	PHP16590	+	PHP16711	-
Cys(-)#8 trAPAO								S				PHP16737	+	n/a	
Cys(-)#6,8 trAPAO						S		S				PHP16738	+		
Cys(-)#3,6,8 trAPAO			S			S		S				PHP17089	+ ¹		
Cys(-)#1,2,7 APAO	A	A					A								

A= alanine

S= serine

1 - activity against FB1 equals wild type; activity against AP1 was reduced.

APAO and trAPAO polypeptide sequence, annotated. (SEQ ID NO: 47)

The amino terminal domain is italicized. Cysteines and residues involved in putative glycosylation sites are underlined. Boxed residues represent amino acids that were successfully altered without complete loss of activity as *E. coli*-expressed protein.

*MALAPSYINPPNVAS*PAGYSHVGVGPDGGGRYVTLAGQIGQDASGVTDPAYEKQVAQAFANLRACLAAVGATSNDVTKLNYIVDYAPSKLTAIGDGLKATFALDRLPPCTLVPVSALSSP

EYLF²⁰⁶EV²⁰⁷D²⁰⁸A²⁰⁹T²¹⁰A²¹¹L²¹²V²¹³P²¹⁴G²¹⁵H²¹⁶T²¹⁷P²¹⁸D²¹⁹N²²⁰V²²¹A²²²D²²³V²²⁴V²²⁵V²²⁶G²²⁷A²²⁸G²²⁹L²³⁰S²³¹G²³²L²³³E²³⁴T²³⁵A²³⁶R²³⁷K²³⁸V²³⁹Q²⁴⁰A²⁴¹A²⁴²G²⁴³L²⁴⁴S²⁴⁵²⁰⁶Q²⁴⁶L²⁴⁷V²⁴⁸L²⁴⁹E²⁵⁰A²⁵¹M²⁵²D²⁵³
RVGGK²⁵⁴T²⁵⁵L²⁵⁶S²⁵⁷V²⁵⁸Q²⁵⁹S²⁶⁰G²⁶¹P²⁶²G²⁶³R²⁶⁴T²⁶⁵T²⁶⁶I²⁶⁷N²⁶⁸D²⁶⁹L²⁷⁰G²⁷¹A²⁷²A²⁷³W²⁷⁴I²⁷⁵N²⁷⁶D²⁷⁷S²⁷⁸N²⁷⁹Q²⁸⁰S²⁸¹E²⁸²V²⁸³S²⁸⁴R²⁸⁵L²⁸⁶F²⁸⁷E²⁸⁸R²⁸⁹F²⁹⁰H²⁹¹L²⁹²E²⁹³G²⁹⁴E²⁹⁵L²⁹⁶Q²⁹⁷R²⁹⁸T²⁹⁹T³⁰⁰G³⁰¹N³⁰²S³⁰³I³⁰⁴H³⁰⁵
QAQD³⁰⁶G³⁰⁷T³⁰⁸T³⁰⁹T³¹⁰T³¹¹A³¹²P³¹³Y³¹⁴G³¹⁵D³¹⁶S³¹⁷L³¹⁸L³¹⁹S³²⁰E³²¹E³²²V³²³A³²⁴S³²⁵A³²⁶L³²⁷A³²⁸E³²⁹L³³⁰L³³¹P³³²V³³³W³³⁴S³³⁵Q³³⁶L³³⁷I³³⁸E³³⁹E³⁴⁰H³⁴¹S³⁴²L³⁴³Q³⁴⁴D³⁴⁵L³⁴⁶K³⁴⁷A³⁴⁸S³⁴⁹P³⁵⁰Q³⁵¹A³⁵²K³⁵³R³⁵⁴L³⁵⁵D³⁵⁶S³⁵⁷V³⁵⁸
SFAHY³⁵⁹C³⁶⁰E³⁶¹K³⁶²E³⁶³L³⁶⁴N³⁶⁵L³⁶⁶P³⁶⁷A³⁶⁸V³⁶⁹L³⁷⁰G³⁷¹V³⁷²A³⁷³N³⁷⁴Q³⁷⁵I³⁷⁶T³⁷⁷R³⁷⁸A³⁷⁹L³⁸⁰L³⁸¹G³⁸²V³⁸³E³⁸⁴A³⁸⁵H³⁸⁶E³⁸⁷I³⁸⁸S³⁸⁹M³⁹⁰L³⁹¹F³⁹²L³⁹³T³⁹⁴D³⁹⁵Y³⁹⁶I³⁹⁷K³⁹⁸S³⁹⁹A⁴⁰⁰T⁴⁰¹G⁴⁰²L⁴⁰³S⁴⁰⁴N⁴⁰⁵I⁴⁰⁶F⁴⁰⁷S⁴⁰⁸D⁴⁰⁹K⁴¹⁰K⁴¹¹
5 DGGQYMR⁴¹²C⁴¹³K⁴¹⁴T⁴¹⁵G⁴¹⁶M⁴¹⁷Q⁴¹⁸S⁴¹⁹I⁴²⁰C⁴²¹H⁴²²A⁴²³M⁴²⁴S⁴²⁵K⁴²⁶E⁴²⁷L⁴²⁸V⁴²⁹P⁴³⁰G⁴³¹S⁴³²V⁴³³H⁴³⁴L⁴³⁵N⁴³⁶T⁴³⁷P⁴³⁸V⁴³⁹A⁴⁴⁰E⁴⁴¹I⁴⁴²E⁴⁴³Q⁴⁴⁴S⁴⁴⁵A⁴⁴⁶S⁴⁴⁷G⁴⁴⁸C⁴⁴⁹T⁴⁵⁰V⁴⁵¹R⁴⁵²S⁴⁵³A⁴⁵⁴S⁴⁵⁵G⁴⁵⁶A⁴⁵⁷V⁴⁵⁸F⁴⁵⁹
RSKKV⁴⁶⁰V⁴⁶¹V⁴⁶²S⁴⁶³L⁴⁶⁴P⁴⁶⁵T⁴⁶⁶T⁴⁶⁷L⁴⁶⁸Y⁴⁶⁹P⁴⁷⁰T⁴⁷¹L⁴⁷²T⁴⁷³F⁴⁷⁴S⁴⁷⁵P⁴⁷⁶P⁴⁷⁷L⁴⁷⁸P⁴⁷⁹A⁴⁸⁰E⁴⁸¹K⁴⁸²Q⁴⁸³A⁴⁸⁴L⁴⁸⁵A⁴⁸⁶E⁴⁸⁷N⁴⁸⁸S⁴⁸⁹I⁴⁹⁰L⁴⁹¹G⁴⁹²Y⁴⁹³Y⁴⁹⁴S⁴⁹⁵K⁴⁹⁶I⁴⁹⁷V⁴⁹⁸F⁴⁹⁹V⁵⁰⁰W⁵⁰¹D⁵⁰²K⁵⁰³P⁵⁰⁴W⁵⁰⁵W⁵⁰⁶R⁵⁰⁷E⁵⁰⁸Q⁵⁰⁹G⁵¹⁰F⁵¹¹
SGVL⁵¹²Q⁵¹³S⁵¹⁴S⁵¹⁵C⁵¹⁶D⁵¹⁷P⁵¹⁸I⁵¹⁹S⁵²⁰F⁵²¹A⁵²²R⁵²³D⁵²⁴T⁵²⁵S⁵²⁶I⁵²⁷D⁵²⁸V⁵²⁹D⁵³⁰R⁵³¹Q⁵³²W⁵³³S⁵³⁴I⁵³⁵T⁵³⁶C⁵³⁷F⁵³⁸M⁵³⁹V⁵⁴⁰G⁵⁴¹D⁵⁴²P⁵⁴³G⁵⁴⁴R⁵⁴⁵K⁵⁴⁶W⁵⁴⁷S⁵⁴⁸Q⁵⁴⁹Q⁵⁵⁰S⁵⁵¹K⁵⁵²Q⁵⁵³V⁵⁵⁴R⁵⁵⁵Q⁵⁵⁶K⁵⁵⁷S⁵⁵⁸V⁵⁵⁹W⁵⁶⁰D⁵⁶¹Q⁵⁶²L⁵⁶³
RAAYENAGA⁵⁶⁴Q⁵⁶⁵V⁵⁶⁶PE⁵⁶⁷PAN⁵⁶⁸V⁵⁶⁹LE⁵⁷⁰IE⁵⁷¹W⁵⁷²S⁵⁷³K⁵⁷⁴Q⁵⁷⁵Q⁵⁷⁶Y⁵⁷⁷F⁵⁷⁸Q⁵⁷⁹G⁵⁸⁰A⁵⁸¹P⁵⁸²S⁵⁸³A⁵⁸⁴V⁵⁸⁵Y⁵⁸⁶G⁵⁸⁷L⁵⁸⁸N⁵⁸⁹D⁵⁹⁰L⁵⁹¹I⁵⁹²T⁵⁹³L⁵⁹⁴G⁵⁹⁵S⁵⁹⁶A⁵⁹⁷L⁵⁹⁸R⁵⁹⁹T⁶⁰⁰P⁶⁰¹F⁶⁰²K⁶⁰³S⁶⁰⁴V⁶⁰⁵
HFVGTETSLVWKG⁶⁰⁶YMEGAIRSGQ⁶⁰⁷RGA⁶⁰⁸AEV⁶⁰⁹VASLVPAA

APAO enzyme activity is maintained when a serine residue at position 206 is mutated to alanine, eliminating a potential glycosylation site (N204 – S206) close to the putative substrate binding site. Please see the tables entitled “Table of site-directed mutagenesis vectors and enzyme assay results” and “Glyc(-) APAO lysates from *E. coli*.”

The polynucleotide sequence of APAO mutated to alter the serine at position 206 to an alanine (S206A) can be seen in SEQ ID NO: 32. The resulting polypeptide is shown in SEQ ID NO: 33.

Glyc(-) APAO lysates from *E. coli*

Sample (lysate)	Substrate	M H ₂ O ₂ /min ¹	Conclusion
WT APAO	AP1	1.92	Active (wild type)
	FB1	0.12	Slightly active (wt)
N204A	AP1	0.09	Slightly active
	FB1	0.04	Slightly active
S206A	AP1	0.85	Partially Active
	FB1	0.07	Slightly active

However, in transient expression assays in maize, expression of S206A resulted in no detectable enzyme activity. Please see the table above entitled “Table of site-directed mutagenesis vectors and enzyme assay results.” Thus, elimination of this glycosylation site is not in itself sufficient to have an active protein upon secretion. This could be due to glycosylation occurring at a second adjacent site (N201 - S203). However, no active APAO was recovered when either N201 or S203 is mutated along with S206. Please see the table entitled “Table of site-directed mutagenesis vectors and enzyme assay results.”

While not to be limited by theory, the molecule may be inactive because both N201 and S203 are buried within the tertiary structure of APAO, and any modification of side chains disrupts proper folding or conformation, or FAD binding. This is backed up by predicted solvent accessibility numbers for these residues in the 3-D model based on the

maize amine oxidase. Please see the table below entitled “Solvent accessibility for cysteine residues of truncated APAO.” The elimination of APAO glycosylation site at amino acids 204 to 206 is not sufficient to allow APAO to be secreted from the cell and retain full enzyme activity, but elimination of this site may improve chances for obtaining a fully active enzyme once the other roadblock(s) to secretability have been resolved. In other words, elimination of this site may be necessary but not sufficient to produce active secretable APAO.

APAO also contains nine cysteine residues, which are likely to be unpaired in the reducing environment of the cytosol but which may crosslink unfavorably upon secretion. Cysteines are present at residues 64, 109, 167, 292, 351, 359, 387, 461, and 482. The 3-D model helps predict the relative location of each amino acid in the structure, and whether it is solvent accessible or buried. Buried residues are more difficult to mutate without destroying structural integrity.

Solvent accessibility for cysteine residues of truncated APAO

APAO Position ¹		Position ²	Cys# ³	aa MPAO		-1	0	1	average	Conclusion
Cys	26	167	3	Leu	32	0.675	0.253	0.24	0.389333	maybe partially exposed
Cys	151	292	4	Asn	147	0.069	0.122	0.147	0.112667	buried
Cys	210	351	5	Tyr	211	0.184	0.244	0.03	0.152667	buried
Cys	218	359	6	Thr	219	0.633	0.319	0.447	0.466333	maybe partially exposed
Cys	246	387	7	Val	247	0.145	0.046	0.366	0.185667	buried
Cys	320	461	8	Ser	324	0.199	0.789	0.643	0.543667	exposed
Cys	341	482	9	Leu	346	0.152	0.071	0.052	0.091667	buried

1. Relative to amino acid 1 of truncated APAO
2. Relative to amino acid 1 of full length APAO
3. Cysteine number relative to full length APAO

Proteins that are secreted to the apoplast are folded to their mature form in the highly oxidizing environment of the ER/Golgi. Among other things this promotes crosslinking of cysteine residues often found in secreted proteins. Unpaired cysteines that are solvent-accessible are rare in secreted proteins, since they would rapidly be oxidized by other cysteine residues of the same protein or another protein. Although not to be limited by theory, it is possible that APAO is normally a cytosolic protein, and thus the presence of nine cysteine residues would not be unusual even though they may not be crosslinked in

the mature protein. In fact, the 3-D model predicts that they would not be crosslinked because the intermolecular distances predicted would be too great. Therefore it is possible that secretion of APAO to the apoplast results in an improper folding and crosslinking of cysteines in the Golgi, and results in inactive enzyme. Using the solvent accessibility
5 tables from APAO modeled against MPAO, the three most solvent-exposed cysteines were identified and then eliminated by site-directed mutagenesis of the APAO cDNA. The sequence of APAO mutated at cysteine 461 and used for expression in bacteria can be seen in SEQ ID NO: 48. The resulting protein is shown in SEQ ID NO: 49. The polynucleotide and resulting polypeptide sequence of APAO mutated at both cysteines 359 and 461 and
10 used for in the bacterial expression system can be seen in SEQ ID NOS: 50 and 51. The polynucleotide and resulting polypeptide sequence of APAO mutated at cysteines 169, 359, and 461 can be seen in SEQ ID NOS: 52 and 53.

The APAO molecules mutated at specific cysteines were tested in a bacterial expression system using the previously described Amplex Red assay. The results can be
15 seen in the table below entitled "Cys(-) APAO lysates from *E. coli*." The mutated APAO molecules can then be tested in maize, linked to a signal peptide, as previously described. Either one of the cysteines or two or three together could be mutated to serines without any measured loss in APAO enzyme activity of the *E coli*-expressed enzyme. In fact, one of the *E coli*-expressed clones (C359S + C461S; PHI16738) had more APAO activity in
20 crude lysates than wild type enzyme and may represent a catalytic improvement. A triply Cys-mutated version of APAO does not show catalytic improvement but retains full activity of the wild type enzyme against FB1, although AP1 activity was somewhat reduced. The mutated versions of APAO operably linked to a signal sequence, which retain
25 additional stability or foldability when expressed in plants or other secretion expression systems.

Cys(-) APAO lysates from *E coli*

Sample (lysate)	Substrate	M H ₂ O ₂ /min ¹	Conclusion
WT APAO	AP1	2.14	Active (wild type)
	FB1	0.11	Slightly active (wt)
C461S	AP1	2.25	Fully Active
	FB1	0.14	Slightly active
C359S, C461S	AP1	3.90	Fully/More Active
	FB1	0.16	Slightly active
C167S, C359S, C461S	AP1	0.27	Slightly active
	FB1	0.25	Slightly active

Triple Cys(-) APAO lysates from *E coli*

Sample (lysate)	Substrate	M H ₂ O ₂ /min ¹	Conclusion
WT APAO	AP1	1.16	Active (wild type)
	FB1	0.27	Slightly active (wt)
C167S, C359S, C461S	AP1	0.27	Slightly Active
	FB1	0.26	Slightly active

It is expected that the S206A mutations will contribute to the functionality of secreted APAO by reducing the degree of glycosylation and the C167S, C359S, and C461S mutations (or combinations thereof) will improve the functionality of secreted APAO by reducing chances for spurious disulfide formation on folding.

To determine expression of a mutated APAO in maize, three APAO constructs were introduced into maize embryos by Agrobacterium-mediated transformation (Zhao et al, 1999, US Patent 5,981,840). The three constructs were PHP17105 (Ubi:BAA:Cys(-)K-trAPAO (C359S, C461S):PinII), PHP17108 (Ubi:Cys(-)K-trAPAO (C359S, C461S):PinII), and PHP17110 (Ubi:APAO:PinII). In addition, PHP16543 (NOS:CRC:PinII-Ubi:MO-PAT:T35) was introduced as a negative control and PHP15258 (Ubi:APAO:PinII-Ubi:BAA:ESP1:PinII-P35S:PAT:T35S) was introduced as a non-targeted positive control. One experiment with two replications was performed. Samples were assayed for both APAO activity by TLC as described previously and by Enzyme Linked ImmunoSorbent Assay (ELISA). For a discussion of ELISA methods, please see, for example, *Current Protocols in Molecular Biology*, 2:11.1.1-11.3.4, John Wiley & Sons, Inc. (Ausubel, et al., eds. 1994). The APAO ELISA is a capture format assay for the quantitative determination of APAO protein in the presence of extracted maize tissue protein. It was performed by co-incubation of biotinylated antibody with an extract prepared from leaf, seed, or callus in phosphate buffered saline with 0.5% Tween-20®. The detection of the antibody complex was accomplished through the added incubation of

streptavidin-alkaline phosphatase (Bio-Rad Life Sciences Products #19542-018), followed by the addition of substrate (pNPP tablets, Sigma #104-105). The resultant color intensity was quantified by determining optical density and was directly proportional to the amount of APAO protein present in the sample extracts. The assay has no matrix effects at 1 µg/well or below for maize leaf, seed, or callus protein. The standard curve was spiked with wild type extract at levels above 1.0 µg/well. The transient testing results are summarized in the table below.

Transient Testing of APAO Constructs (6-8-2000)

Experiment	Rep	Construct	APAO-TLC	APAO-ELISA (ppm)
negative control	none	none	0	-2
4350.08.01	1	php16543, as a (-) control	0	-4
4350.08.02		php15258, non-targeted APAO as a (+) control	3	out high
4350.08.03		php17105, UBI-BAA::CYS(-)K-TR-APAO (C359S, C461S)	1	107
4350.08.04		php17108, UBI-CYS(-)K-TR-APAO (C359S, C461S)	3	270
4350.08.05		php17110, UBI-APAO	3	out high
4350.08.06	2	php16543	0	-5
4350.08.07		php15258	3	313
4350.08.08		php17105	0	52
4350.08.09		php17108	2	143
4350.08.10		php17110	2	123
3477.27.01	transformed callus lines	php15258 as postive controls	1	118
3477.27.02			2	141
3477.27.03			2	187
3477.27.04			2	184

As can be seen in the Table above, the BAA-targeted APAO (PHP17105) did not accumulate as much APAO as the non-BAA targeted counterpart (PHP17108). Although not to be limited by theory, the lack of APAO protein accumulation rather than APAO function may play a role in the lack of detectable APAO activity with the BAA-targeted APAO construct. It appears that only when the APAO concentration exceeds 100 ppm can APAO activity be seen by TLC. Nevertheless, the double Cys(-) mutant is active in maize when expressed either cytosolically or extracellularly.

EXAMPLE 16

Other APAO Polynucleotides From *Exophiala spinifera* and *Rhinocladiella atrovirens*

Using primers designed from the APAO isolated from *Exophiala spinifera*, ATCC 74269 (Table 15), three new APAO polynucleotides were isolated from *Exophiala spinifera* (isolates ESP002 and ESP003), designated ESP002_C2, ESP002_C3 and ESP003_C12

and three new APAO polynucleotides from *Rhinocladia atrovirens* (isolate RAT011) designated RAT011_C1, RAT011_C2, RAT011_C4. The strains used to isolate the polynucleotides are described below.

5

Isolate	Genus species	Source	FB1 degrader	APAO homologs isolated
ESP002	<i>Exophiala spinifera</i>	Palm, ATCC 26089	Yes	ESP002_c2 in pGEX4T1 ESP002_c3 in pGEX4T1
ESP003	<i>Exophiala spinifera</i>	Maize seed	Yes	ESP003_c12 in pGEX4T1
RAT011	<i>Rhinocladia atrovirens</i>	Maize seed	Yes	RAT011_c1 in pGEM11Zf+ RAT011_c2 in pGEX4T1 RAT011_c4 in pGEM11Zf+

Growth conditions and production of culture material

1. Streak 150 x 15 mm YPD plates with a glycerol aliquot of the above fungal isolates.
2. Grow at 28° C in the dark until there is sufficient growth for inoculating liquid medium usually at least two weeks.
3. Mycelia and spores were scraped from the plates or agar cubes used to inoculate 50 mls YPD broth in 250 ml baffled flasks.
4. Flasks of culture material were grown at 28° C in the dark at ~125 rpm.
5. After sufficient growth was obtained the cultures were harvested by pelleting the culture in 50 ml centrifuge tubes at 3400 rpm for 15 min.
6. The supernatant was discarded and the pellets were frozen at -20° C.

YPD broth and agar medium

Amount per liter:	Yeast Extract	10 g	
	Bactopeptone	20 g	
	Dextrose	0.5 g	
	Bactoagar	15 g	(for agar media only)

DNA Isolation,

The DNA was isolated according to a modified version of a plant CTAB DNA extraction protocol (Saghai-Maroo MA, *et al.*, *Proc Natl Acad Sci, USA*, 81:8014-8018 (1984)) as follows.

1. Place 0.2-0.5 g (dry weight) lyophilized fungal mycelium in a 50 ml disposable centrifuge tube, break up mat with a spatula or glass rod. Shake briefly.
2. Add 10 ml (per 0.5 g mat) of CTAB extraction buffer. Gently mix to wet all the powdered mat.

3. Place in 65° C water bath for 30 minutes.
4. Cool. Add an equal volume of phenol:chloroform. Shake briefly to mix.
5. Centrifuge 20 minutes at 3400 rpm.
6. To the aqueous phase add an equal volume of chloroform:isoamyl alcohol (24:1). Shake briefly to mix.
7. Centrifuge 15 minutes at 3400 rpm.
8. To aqueous phase add an equal volume of isopropanol.
9. Centrifuge for 30 minutes at 3400 rpm to pellet precipitated DNA.
10. Rinse DNA pellet with 70% ethanol.
11. Air dry pellet.
12. Resuspend pellet in 1-5 ml TE containing 20 ug/ml RNase A.

CTAB Extraction Buffer

- 0.1 M Tris, pH 7.5
- 1% CTAB (mixed hexadecyl trimethyl ammonium bromide)
- 0.7 M NaCl
- 10 mM EDTA
- 1% 2-mercaptoethanol
- Add proteinase K to a final concentration of 0.3 mg/ml prior to use.

Primer Design

Primers used were gene specific primers based on APAO polynucleotide sequence (SEQ ID NO: 22) with restriction enzymes sites for cloning. The 5'-primer, 26194, contained the restriction enzyme recognition site, EcoRI. The complementary 3'-primer, 23259, contained the restriction enzyme recognition site, NotI.

26194
5' ggggaattcATGGCACTTGCACCGAGCTACATCAATC 3' , 37-mer (SEQ ID NO: 34)

23259
5' gggGCGGCCCGCCTATGCTGCTGGCACCAGGCTAG 3' , 34-mer (SEQ ID NO: 13)

PCR conditions

1. The PCR cocktail:

10 mM dNTPs	1 ul
10X Advantage polymerase buffer	5 ul
HPLC water	38 ul
10 uM primer 26194	2 ul
10 uM primer 23259	2 ul
50 X Advantage polymerase mix	1 ul

(Clontech)

Template, genomic DNA, 50 ng/ul 1 ul

2. Thermocycling conditions:
MJ PTC-100 AgV Thermocycler:

Step	1	95°	2 minutes
	2	95°	30 seconds
	3	60°	1 minute
	4	72°	1 minute 30 seconds
	5	Go to step 2, 34X more	
	6	72°	5 minutes
	7	4°	Hold
	8	End	

3. PCR products were analyzed on a 1% LE-agarose, TAE plus ethidium bromide gel.
Bands of about 1900 bp were seen on the gel. The band was not present in the no DNA control reaction.

Cloning Protocols

1. DNA was extracted from excised gel fragments using a QIAGEN Gel Extraction Kit (Catalog number 28704, QIAGEN, Santa Clara, CA).
2. PCR fragments were digested with EcoRI and Not I to free up the sites for cloning into EcoRI and Not I digested vector, either pGEX4T1 (Pharmacia) or pGEM11Zf+ (Promega).
3. Digests were cleaned up and desalted using a QIAquick PCR Purification Kit (Catalog number 28104).
4. Isolated fragment was quantified and checked for purity on a 1% LE-agarose, TAE + ethidium bromide gel.
5. Fragments were ligated into compatible sites in either pGEX4T1 (Pharmacia) or pGEM11Zf+ (Promega).
6. After heat inactivation Library efficiency DH5 competent *E. coli* were transformed with a small amount of the ligation reaction.
7. LB + carbenicillin, 50 ug/ml, plates were spread with an aliquot of the transformation mix, grown overnight at 37° C.
8. Colonies were screened for full-length insert using a PCR miniprep method utilizing vector primers flanking the multiple cloning region.

9. Positive clones were identified and overnight cultures grown for plasmid isolation and verification by sequencing.
10. Positive clones are identified as follows:
 - DH5 :pGEX4T1:ESP002FL_c2 (from palm tree isolate)
 - DH5 :pGEX4T1:ESP002FL_c3 (from palm tree isolate)
 - DH5 :pGEX4T1:ESP003FL_c12 (from maize isolate)
 - DH5 :pGEM11Zf+:RAT011FL_c1 (from maize isolate)
 - DH5 :pGEM11Zf+:RAT011FL_c4 (from maize isolate)
 - DH5 :pGEX4T1:RAT011FL_c2 (from maize isolate)

**Important note: These are genomic clones containing two introns

Sequence Results

Three APAO polynucleotides and related polypeptides were isolated from *Exophiala spinifera* (isolates ESP002 and ESP003), designated ESP002_C2, (SEQ ID NOS: 35 and 36) ESP002_C3 (SEQ ID NOS: 37 and 38) and ESP003_C12 (SEQ ID NOS: 39 and 40). Three APAO polynucleotides were isolated from *Rhinocladia atrovirens* (isolate RAT011) designated RAT011_C1 (SEQ ID NOS: 41 and 42), RAT011_C2 (SEQ ID NOS: 43 and 44), and RAT011_C4 (SEQ ID NOS: 45 and 46). Introns were detected by comparison of the genomic sequence with the cDNA sequence of APAO from *E. spinifera* 2141.10 (SEQ ID NO: 22), and by identifying putative intron splice junctions in the gap domains (Shah, *et al.*, *Journal of Molecular and Applied Genetics* 2:111-126 (1983)).

Plasmids containing the polynucleotide sequences of the invention were deposited with American Type Culture Collection (ATCC), Manassas, Virginia, and assigned Accession No. 98812, 98813, 98814, 98815, 98816, (all deposited on July 15, 1998) and PTA-32 (deposited on May 7, 1999). The deposits will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. The deposits were made merely as a convenience for those of skill in the art and are not an admission that a deposit is required under 35 U.S.C. § 112.

Preliminary sequence results were entered into GCG, and nucleotide and protein alignments were done in a pileup using a software program called Genedoc for shading and homology comparisons (Nicholas, *et al.*, *EMBNW.NEWS* 4:14 (1997); or at the Internet site <http://www.cris.com/~Ketchup/genedoc.shtml>). The first APAO (SEQ ID NO: 22) sequence was included for comparison. Comparing the reference sequence SEQ ID NO:

22 to the other homologs sequence identities range from 96 to 99% (identities are lower since APAO introns were not included). Homologies are slightly higher comparing *Exophiala* genes sequences. At the amino acid sequence level the comparison of the reference sequence (SEQ ID NO: 23) to the other homologs yielded sequence identities of approximately 97%.

All publications and patent applications in this specification are indicative of the level of ordinary skill in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated by reference.

The invention has been described with reference to various specific and preferred embodiments and techniques. However, it should be understood that many variations and modifications may be made while remaining within the spirit and scope of the invention.

WHAT IS CLAIMED IS:

1. An isolated polynucleotide comprising an APAO encoding polynucleotide linked to a fumonisin esterase encoding polynucleotide, wherein the APAO encoding polynucleotide comprises a member selected from:
 - a) a polynucleotide encoding a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53;
 - b) a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 33, 35, 37, 39, 41, 43, 45, 48, 50 and 52; and
 - c) a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52.
2. A recombinant expression cassette comprising a polynucleotide of claim 1 operably linked to a promoter.
3. The recombinant expression cassette of claim 2 wherein the polynucleotide is operably linked to a plant signal sequence.
4. A vector comprising the recombinant expression cassette of claim 2.
5. A host cell comprising the recombinant expression cassette of claim 2.
6. The host cell of claim 5 wherein the cell is a plant cell.
7. The host cell of claim 6 wherein the plant cell is selected from the group consisting of maize, sorghum, wheat, tomato, soybean, alfalfa, sunflower, canola, cotton, barley, millet, and rice.
8. A plant comprising a polynucleotide of claim 1.
9. A seed from a plant of claim 7.

10. An isolated polypeptide comprising a member selected from:
- a) a polypeptide comprising at least 70% sequence identity to a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53;
 - b) a polypeptide encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52; and
 - c) a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53.
11. The polynucleotide of claim 1 wherein the fumonisin esterase encoding polynucleotide is ESP1.
12. The polynucleotide of claim 11 wherein the polynucleotide is set forth in SEQ ID NO: 24.
13. The polynucleotide of claim 1 wherein the fumonisin esterase encoding polynucleotide is BEST1.
14. The polynucleotide of claim 13 wherein the polynucleotide is set forth in SEQ ID NO: 26.
15. A method of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a breakdown product of a structurally related mycotoxin comprising the steps of:
- a) applying an APAO enzyme as a spray or wash; and
 - b) under degradation conditions allowing sufficient time for the polypeptide to degrade the fumonisin, the structurally related mycotoxin, the fumonisin breakdown product, or the breakdown product of a structurally related mycotoxin.
16. The method of claim 15 wherein the fumonisin or structurally related mycotoxin is present in harvested grain.

17. The method of claim 15 wherein degradation occurs during processing of the harvested grain.
18. The method of claim 17 wherein the harvested grain is to be used as animal feed.
19. The method of claim 15 wherein degradation occurs in silage.
20. The method of claim 15 wherein fumonisin esterase is also added at or before step (a).
21. The method of claim 15 wherein the APAO enzyme is selected from:
- a) a polypeptide comprising at least 70% sequence identity to a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53;
 - b) a polypeptide encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52; and
 - c) a polypeptide selected from SEQ ID NOS: 6, 11, 23, 36, 38, 40, 42, 44, 46, 49, 51 and 53.
22. A method of identifying transformed plant cells comprising the steps of:
- a) introducing into a plant cell at least one copy of an expression cassette comprising an APAO encoding polynucleotide;
 - b) placing the plant cell on culture media containing an AP1 or a phytotoxic analog; and
 - c) identifying transformed cells as the surviving cells in the culture.
23. The method of claim 22 wherein the APAO encoding polynucleotide comprises a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, and 50.
24. The method of claim 22 wherein a fumonisin esterase encoding polynucleotide is also introduced into the plant cell.

25. A method of detecting fumonisins or structurally related toxins, the method comprising:

- a) adding APAO enzymes to a sample containing fumonisin or a structurally related toxin;
- b) reacting the sample under conditions of time and temperature sufficient to convert the toxin to the corresponding oxidized or deaminated toxin; and
- c) detecting the hydrogen peroxide or ammonia produced.

26. The method of claim 25 wherein the APAO enzyme is encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, and 50.

27. The method of claim 25 wherein fumonisin esterase is added at or before step (a).

28. A method of producing a plant capable of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a breakdown product of a structurally related mycotoxin comprising the steps of:

- a) introducing into a plant cell at least one copy of an expression cassette comprising a polynucleotide encoding an APAO enzyme operably linked to a promoter; and
- b) under degradation conditions expressing the APAO enzyme for a time sufficient to degrade the fumonisin, the fumonisin breakdown product, the structurally related mycotoxin, AP1, or a breakdown product of a structurally related mycotoxin.

29. The method of claim 28 wherein a polynucleotide encoding a fumonisin esterase is also introduced.

30. The method of claim 28 wherein the APAO enzyme is encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52.

31. The method of claim 28 wherein the plant cell is regenerated into a plant.
32. The method of claim 28 wherein a fumonisin esterase encoding polynucleotide is also introduced.
33. A host cell comprising an APAO encoding polynucleotide and a fumonisin esterase encoding polynucleotide.
34. The host cell of claim 33 wherein the APAO encoding polynucleotide comprises a polynucleotide having at least 70% identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, and 50.
35. The host cell of claim 33 wherein the fumonisin esterase encoding polynucleotide is selected from ESP1 and BEST1.
36. The host cell of claim 33 wherein the cell is a plant cell.
37. The host cell of claim 36 wherein the cell is selected from maize, sorghum, wheat, tomato, soybean, alfalfa, sunflower, canola, cotton, and rice.
38. The host cell of claim 37 wherein the plant cell is regenerated into a plant.
39. A method of predicting possible mutagenesis sites on APAO comprising the steps of:
- a) developing a 3-dimensional model of APAO; and
 - b) identifying sites on APAO to mutate by evaluating the likelihood that particular amino acids could contribute to misfolding.
40. A 3-dimensional model of APAO generated by an automated modeling program.
41. The model of claim 40 wherein the automated modeling program is *Modeler*.

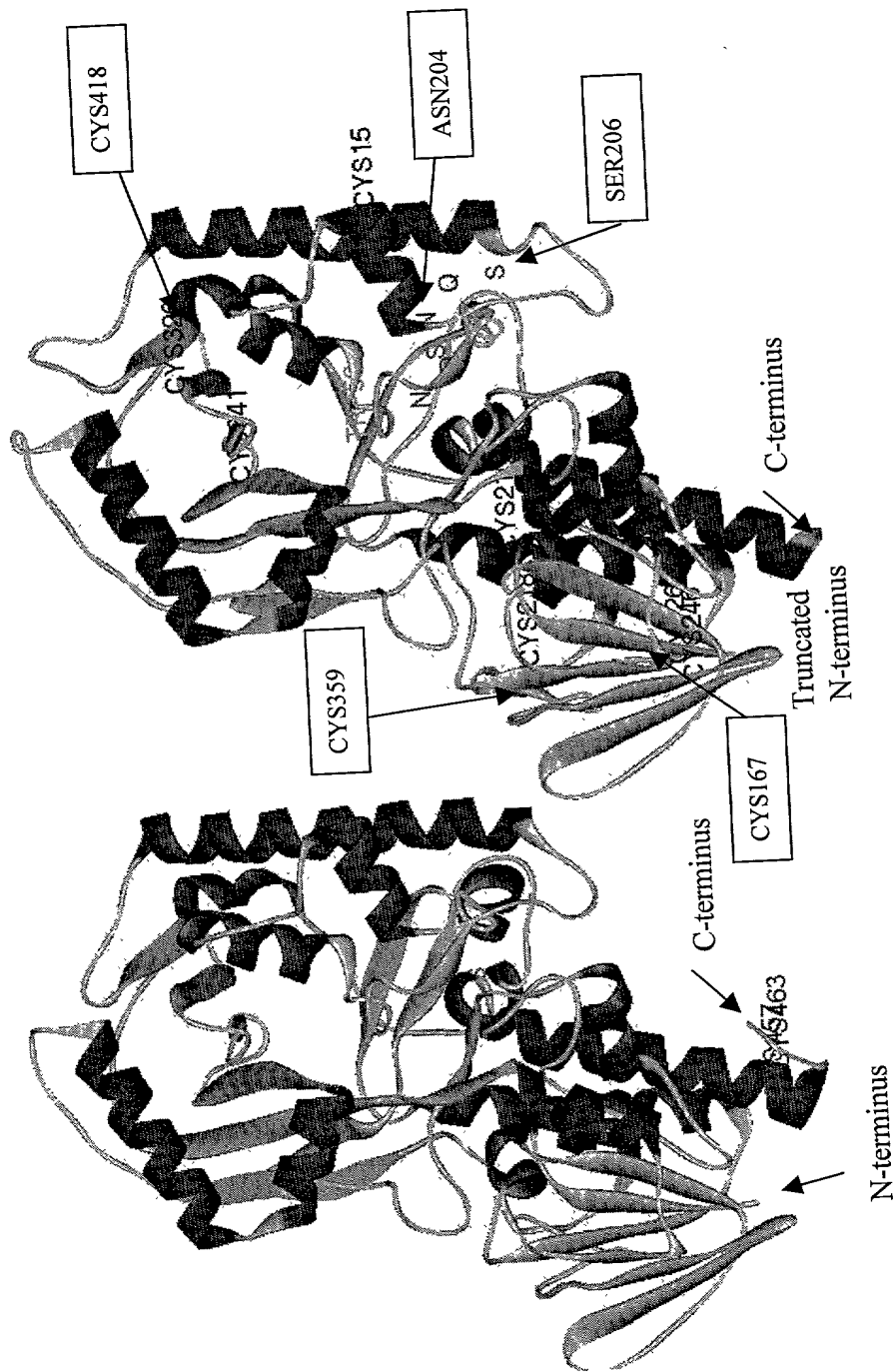
AMINO POLYOL AMINE OXIDASE POLYNUCLEOTIDES AND RELATED POLYPEPTIDES AND METHODS OF USE

5 **Abstract**

 The present invention provides polynucleotides and related polypeptides of the enzyme APAO isolated from *Exophiala spinifera* and *Rhinocladiella atrovirens*. The polynucleotides may be mutated to remove glycosylation sites and cysteine residues. Additionally, the present invention provides recombinant expression cassettes, host cells, transgenic plants, and transgenic seed. The present invention also provides for polynucleotides containing both APAO and a fumonisin esterase. In addition, the present invention provides methods for producing the APAO enzyme in both prokaryotic and eukaryotic systems, methods for detecting fumonisins, and methods for identifying transformed plant cells. Methods for degrading fungal toxins in plants, grain, grain processing, silage, food crops and in animal feed are also disclosed.

10

15



B

A

Figure 1

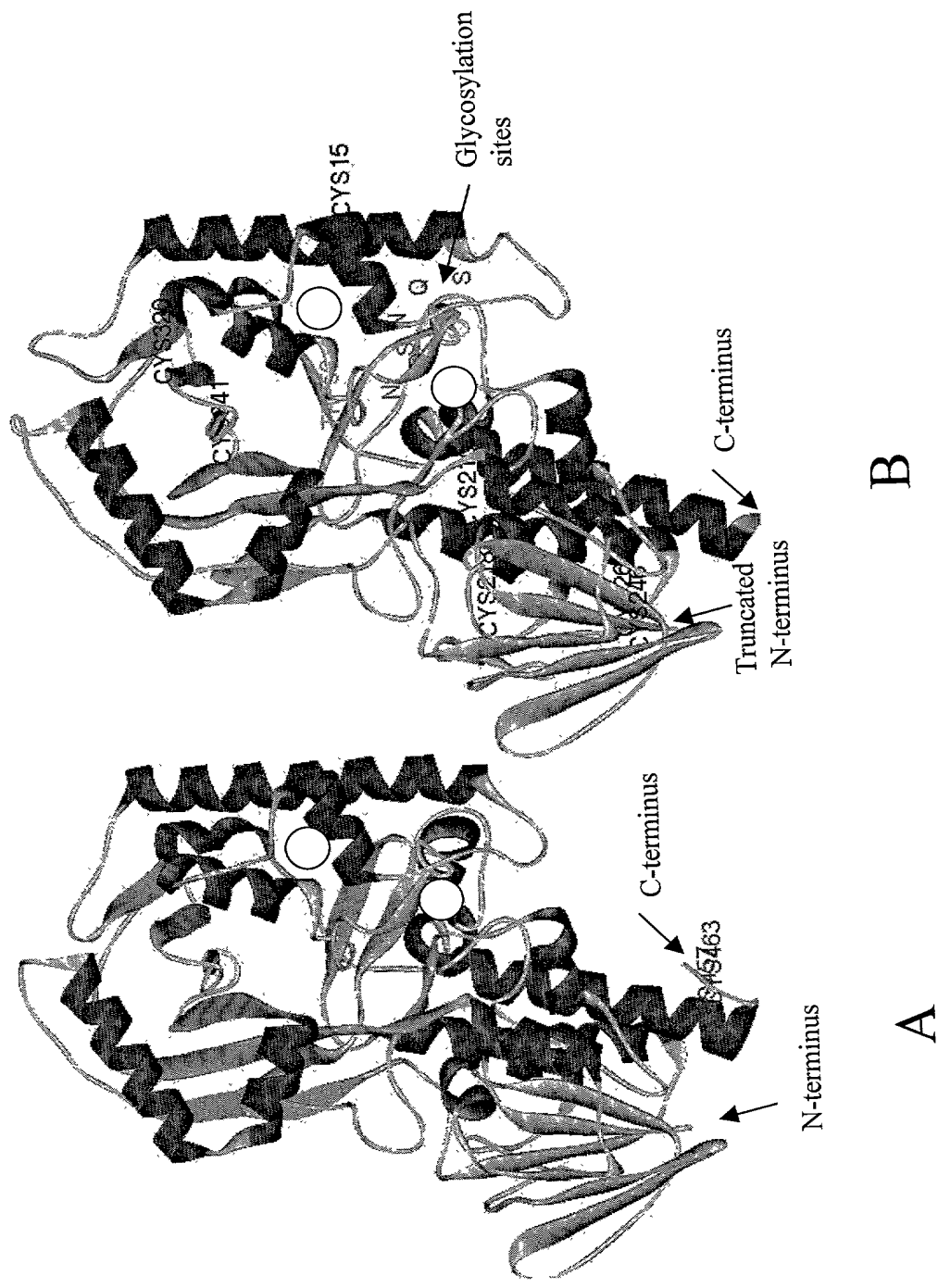


Figure 2

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Polynucleotides and Related Polypeptides and Methods of Use

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100105110																
gag Glu	gtt Val	gca Ala	agt Ser 115	gca Ala	ctt Leu	gcg Ala	gaa Glu 120	ctc Leu	ctc Leu	ccc Pro	gta Val	tgg Trp	tct Ser	cag Gln	ctg Leu	384
115120125																
atc Ile	gaa Glu 130	gag Glu	cat His	agc Ser	ctt Leu	caa Gln 135	gac Asp	ctc Leu	aag Lys	gcg Ala	agc Ser 140	cct Pro	cag Gln	gcg Ala	aag Lys	432
130135140																
cgg Arg 145	ctc Leu	gac Asp	agt Ser	gtg Val	agc Ser	ttc Phe	gcg Ala	cac His	tac Tyr	tgt Cys	gag Glu	aag Lys	gaa Glu	cta Leu	aac Asn 160	480
145150155160																
ttg Leu	cct Pro	gct Ala	gtt Val	ctc Leu 165	ggc Gly	gta Val	gca Ala	aac Asn	cag Gln	atc Ile	aca Thr	gcg Arg	gct Ala	ctg Leu	ctc Leu	528
165170175																
ggc Gly	gtg Val	gaa Glu	gcc Ala 180	cac His	gag Glu	atc Ile	agc Ser	atg Met 185	ctt Leu	ttt Phe	ctc Leu	acc Thr	gac Asp	tac Tyr	atc Ile	576
180185190																
aag Lys	agt Ser	gcc Ala	acc Thr 195	ggc Gly	ctc Leu	agt Ser	aat Asn 200	att Ile	ttc Phe	tcg Ser	gac Asp	aag Lys	aaa Lys	gac Asp	ggc Gly	624
195200205																
ggg Gly	cag Gln	tat Tyr	atg Met	cga Arg	tgc Cys	aaa Lys 215	aca Thr	ggc Gly	atg Met	cag Gln	tcg Ser	att Ile	tgc Cys	cat His	gcc Ala	672
210215220																
atg Met 225	tca Ser	aag Lys	gaa Glu	ctt Leu	gtt Val	cca Pro	ggc Gly	tca Ser	gtg Val	cac His	ctc Leu	aac Asn	acc Thr	ccc Pro	gtc Val 240	720
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gct gaa att gag cag tcg gca tcc ggc tgt aca gta cga tcg gcc tcg Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser 245 250 255	768
ggc gcc gtg ttc cga agc aaa aag gtg gtg gtt tcg tta ccg aca acc Gly Ala Val Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr 260 265 270	816
ttg tat ccc acc ttg aca ttt tca cca cct ctt ccc gcc gag aag caa Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln 275 280 285	864
gca ttg gcg gaa aat tct atc ctg ggc tac tat agc aag ata gtc ttc Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe 290 295 300	912
gta tgg gac aag ccg tgg tgg cgc gaa caa ggc ttc tcg ggc gtc ctc Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu 305 310 315 320	960
caa tcg agc tgt gac ccc atc tca ttt gcc aga gat acc agc atc gac Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp 325 330 335	1008
gtc gat cga caa tgg tcc att acc tgt ttc atg gtc gga gac ccg gga Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly 340 345 350	1056
cgg aag tgg tcc caa cag tcc aag cag gta cga caa aag tct gtc tgg Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp 355 360 365	1104
gac caa ctc cgc gca gcc tac gag aac gcc ggg gcc caa gtc cca gag Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu 370 375 380	1152
ccg gcc aac gtg ctc gaa atc gag tgg tcg aag cag cag tat ttc caa Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln 385 390 395 400	1200
gga gct ccg agc gcc gtc tat ggg ctg aac gat ctc atc aca ctg ggt Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly 405 410 415	1248
tcg gcg ctc aga acg ccg ttc aag agt gtt cat ttc gtt gga acg gag Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu 420 425 430	1296
acg tct tta gtt tgg aaa ggg tat atg gaa ggg gcc ata cga tcg ggt Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly 435 440 445	1344
caa cga ggt gct gca gaa gtt gtg gct agc ctg gtg cca gca gca Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala 450 455 460	1389
tag	1392

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 <212> PRT
 <213> Exophiala spinifera

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Leu	Glu	Thr	Ala 20	Arg	Lys	Val	Gln	Ala 25	Ala	Gly	Leu	Ser 30	Cys	Leu	Val
Leu	Glu	Ala 35	Met	Asp	Arg	Val	Gly 40	Gly	Lys	Thr	Leu 45	Val	Gln	Ser	
Gly	Pro 50	Gly	Arg	Thr	Thr	Ile 55	Asn	Asp	Leu	Gly 60	Ala	Ala	Trp	Ile	Asn
Asp 65	Ser	Asn	Gln	Ser	Glu 70	Val	Ser	Arg	Leu 75	Phe	Glu	Arg	Phe	His	Leu 80
Glu	Gly	Glu	Leu	Gln 85	Arg	Thr	Thr	Gly 90	Asn	Ser	Ile	His	Gln 95	Ala	Gln
Asp	Gly	Thr	Thr 100	Thr	Ala	Pro	Tyr 105	Gly	Asp	Ser	Leu 110	Leu	Ser	Glu	
Glu	Val	Ala 115	Ser	Ala	Leu	Ala	Glu 120	Leu	Leu	Pro	Val	Trp 125	Ser	Gln	Leu
Ile	Glu	Glu	His	Ser	Leu	Gln	Asp 130	Leu	Lys	Ala 140	Ser	Pro	Gln	Ala	Lys
Arg 145	Leu	Asp	Ser	Val	Ser 150	Phe	Ala	His	Tyr	Cys 155	Glu	Lys	Glu	Leu	Asn 160
Leu	Pro	Ala	Val	Leu 165	Gly	Val	Ala	Asn	Gln	Ile 170	Thr	Arg	Ala	Leu	Leu 175
Gly	Val	Glu	Ala 180	His	Glu	Ile	Ser 185	Met	Leu	Phe	Leu	Thr 190	Asp	Tyr	Ile
Lys	Ser	Ala 195	Thr	Gly	Leu	Ser	Asn 200	Ile	Phe	Ser	Asp 205	Lys	Lys	Asp	Gly
Gly	Gln	Tyr	Met	Arg	Cys	Lys	Thr 210	Gly	Met	Gln	Ser 220	Ile	Cys	His	Ala
Met 225	Ser	Lys	Glu	Leu	Val 230	Pro	Gly	Ser	Val	His 235	Leu	Asn	Thr	Pro	Val 240
Ala	Glu	Ile	Glu	Gln 245	Ser	Ala	Ser	Gly	Cys	Thr	Val	Arg	Ser	Ala	Ser 255
Gly	Ala	Val	Phe 260	Arg	Ser	Lys	Lys	Val 265	Val	Val	Ser	Leu	Pro 270	Thr	Thr
Leu	Tyr	Pro	Thr 275	Leu	Thr	Phe	Ser 280	Pro	Pro	Leu	Pro	Ala	Glu	Lys	Gln
Ala	Leu	Ala	Glu 290	Asn	Ser	Ile	Leu 295	Gly	Tyr	Tyr	Ser 300	Lys	Ile	Val	Phe
Val	Trp	Asp	Lys	Pro	Trp	Trp	Arg	Glu	Gln	Gly	Phe	Ser	Gly	Val	Leu 320
Gln	Ser	Ser	Cys 325	Asp	Pro	Ile	Ser	Phe	Ala	Arg	Asp	Thr	Ser	Ile	Asp 335
Val	Asp	Arg	Gln 340	Trp	Ser	Ile	Thr	Cys 345	Phe	Met	Val	Gly	Asp	Pro	Gly
Arg	Lys	Trp	Ser 355	Gln	Gln	Ser	Lys 360	Gln	Val	Arg	Gln	Lys	Ser	Val	Trp
Asp	Gln	Leu	Arg	Ala	Ala	Tyr	Glu 370	Asn	Ala	Gly	Ala 380	Gln	Val	Pro	Glu
Pro	Ala	Asn	Val	Leu	Glu	Ile	Glu 385	Trp	Ser	Lys	Gln	Gln	Tyr	Phe	Gln 400
Gly	Ala	Pro	Ser	Ala 405	Val	Tyr	Gly	Leu	Asn	Asp	Leu	Ile	Thr	Leu	Gly
Ser	Ala	Leu	Arg	Thr 420	Pro	Phe	Lys	Ser	Val	His	Phe	Val	Gly	Thr	Glu
Thr	Ser	Leu	Val	Trp 435	Lys	Gly	Tyr 440	Met	Glu	Gly	Ala	Ile	Arg	Ser	Gly
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							455				460				

12

<211> 34
<212> DNA
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<220>
<223> Primer sequence designed for cloning DNA into
expression vectors, N23256

<400> 12
ggggaattca aagacaacgt tgcggacgtg gtag 34

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<211> 34
<212> DNA
<213> Artificial Sequence

<220>
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expression vectors, N23259

<400> 13
ggggcgcccg cctatgctgc tggcaccagg ctag 34

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<211> 29
<212> DNA
<213> Artificial Sequence

<220>
<223> Designed oligonucleotide for 3' RACE, N21965

<400> 14
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<211> 28
<212> DNA
<213> Artificial Sequence

<220>
<223> Designed oligonucleotide for 5' race, N21968

<400> 15
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<210> 16
<211> 1673
<212> DNA
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<220>
<221> sig_peptide
<222> (1)...(267)
<223> yeast alpha mating factor secretion signal.

<221> CDS
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<400> 16
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Variable	Mean	SD	Min	Max	Median	Mode	Skewness	Kurtosis	Normality
Age	35.5	10.5	20	65	35	35	0.1	3.0	0.95
Gender	1.5	0.5	1	2	1	1	0.0	0.0	0.99
Marital Status	2.5	1.0	1	4	2	2	0.2	1.5	0.98
Education	12.5	2.0	10	16	12	12	0.1	3.0	0.95
Income	1500	500	500	3000	1200	1000	0.3	2.5	0.97
Occupation	1.5	0.5	1	2	1	1	0.0	0.0	0.99
Health Status	2.5	1.0	1	4	2	2	0.2	1.5	0.98
Stress Level	3.5	1.5	1	5	3	3	0.1	3.0	0.95
Life Satisfaction	4.5	1.0	3	5	4	4	0.0	0.0	0.99
Resilience	3.0	1.0	1	5	3	3	0.1	3.0	0.95
Optimism	4.0	1.0	3	5	4	4	0.0	0.0	0.99
Emotional Stability	3.5	1.0	1	5	3	3	0.1	3.0	0.95
Self-Esteem	4.0	1.0	3	5	4	4	0.0	0.0	0.99
Life Satisfaction	4.5	1.0	3	5	4	4	0.0	0.0	0.99
Resilience	3.0	1.0	1	5	3	3	0.1	3.0	0.95
Optimism	4.0	1.0	3	5	4	4	0.0	0.0	0.99
Emotional Stability	3.5	1.0	1	5	3	3	0.1	3.0	0.95
Self-Esteem	4.0	1.0	3	5	4	4	0.0	0.0	0.99

ggc Gly	gta Val	gca Ala 170	aac Asn	cag Gln	atc Ile	aca Thr	cgc Arg 175	gct Ala	ctg Leu	ctc Leu	ggg Gly	gtg Val 180	gaa Glu	gcc Ala	cac His	816
gag Glu	atc Ile 185	agc Ser	atg Met	ctt Leu	ttt Phe	ctc Leu 190	acc Thr	gac Asp	tac Tyr	atc Ile	aag Lys 195	agt Ser	gcc Ala	acc Thr	ggg Gly	864
ctc Leu 200	agt Ser	aat Asn	att Ile	ttc Phe	tgc Ser 205	gac Asp	aag Lys	aaa Lys	gac Asp	ggc Gly 210	ggg Gly	cag Gln	tat Tyr	atg Met	cga Arg 215	912
tgc Cys	aaa Lys	aca Thr	ggg Gly	atg Met 220	cag Gln	tgc Ser	att Ile	tgc Cys	cat His 225	gcc Ala	atg Met	tca Ser	aag Lys	gaa Glu 230	ctt Leu	960
gtt Val	cca Pro	ggc Gly	tca Ser 235	gtg Val	cac His	ctc Leu	aac Asn	acc Thr 240	ccc Pro	gtc Val	gct Ala	gaa Glu 245	att Ile	gag Glu	cag Gln	1008
tgc Ser	gca Ala	tcc Ser 250	ggc Gly	tgt Cys	aca Thr	gta Val	cga Arg 255	tgc Ser	gcc Ala	tgc Ser	ggc Gly	gcc Ala 260	gtg Val	ttc Phe	cga Arg	1056
agc Ser	aaa Lys 265	aag Lys	gtg Val	gtg Val	gtt Val	tgc Ser 270	tta Leu	ccg Pro	aca Thr	acc Thr	ttg Leu 275	tat Tyr	ccc Pro	acc Thr	ttg Leu	1104
aca Thr 280	ttt Phe	tca Ser	cca Pro	cct Pro	ctt Leu 285	ccc Pro	gcc Ala	gag Glu	aag Lys	caa Gln 290	gca Ala	ttg Leu	gcg Ala	gaa Glu	aat Asn 295	1152
tct Ser	atc Ile	ctg Leu	ggc Gly	tac Tyr 300	tat Tyr	agc Ser	aag Lys	ata Ile	gtc Val 305	ttc Phe	gta Val	tgg Trp	gac Asp	aag Lys	ccg Pro 310	1200
tgg Trp	tgg Trp	cgc Arg 315	gaa Glu	caa Gln	ggc Gly	ttc Phe	tgc Ser 320	ggc Gly	gtc Val	ctc Leu	caa Gln	tgc Ser 325	agc Ser	tgt Cys	gac Asp	1248
ccc Pro	atc Ile	tca Ser 330	ttt Phe	gcc Ala	aga Arg	gat Asp	acc Thr 335	agc Ser	atc Ile	gac Asp	gtc Val 340	gat Asp	cga Arg	caa Gln	tgg Trp	1296
tcc Ser	att Ile 345	acc Thr	tgt Cys	ttc Phe	atg Met	gtc Val 350	gga Gly	gac Asp	ccg Pro	gga Gly	cgg Arg 355	aag Lys	tgg Trp	tcc Ser	caa Gln	1344
cag Gln 360	tcc Ser	aag Lys	cag Gln	gta Val	cga Arg 365	caa Gln	aag Lys	tct Ser	gtc Val	tgg Trp 370	gac Asp	caa Gln	ctc Leu	cgc Arg	gca Ala 375	1392
gcc Ala	tac Tyr	gag Glu	aac Asn	gcc Ala 380	ggg Gly	gcc Ala	caa Gln	gtc Val 385	cca Pro	gag Glu	ccg Pro	gcc Ala	aac Asn	gtg Val 390	ctc Leu	1440
gaa Glu	atc Ile	gag Glu	tgg Trp 395	tgc Ser	aag Lys	cag Gln	cag Gln	tat Tyr 400	ttc Phe	caa Gln	gga Gly	gct Ala 405	ccg Pro	agc Ser	gcc Ala	1488
gtc	tat	ggg	ctg	aac	gat	ctc	atc	aca	ctg	ggg	tgc	gcg	ctc	aga	acg	1536

Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr
410 415 420

ccg ttc aag agt gtt cat ttc gtt gga acg gag acg tct tta gtt tgg 1584
Pro Phe Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp
425 430 435

aaa ggg tat atg gaa ggg gcc ata cga tcg ggt caa cga ggt gct gca 1632
Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala
440 445 450 455

gaa gtt gtg gct agc ctg gtg cca gca gca taggcggccg c 1673
Glu Val Val Ala Ser Leu Val Pro Ala Ala
460 465

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<213> Exophiala spinifera

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<221> SIGNAL
<222> (1) ... (89)

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-70 -65 -60
Ile Pro Ala Glu Ala Val Ile Gly Tyr Ser Asp Leu Glu Gly Asp Phe
-55 -50 -45
Asp Val Ala Val Leu Pro Phe Ser Asn Ser Thr Asn Asn Gly Leu Leu
-40 -35 -30
Phe Ile Asn Thr Thr Ile Ala Ser Ile Ala Ala Lys Glu Glu Gly Val
-25 -20 -15 -10
Ser Leu Glu Lys Arg Glu Ala Glu Ala Glu Phe Lys Asp Asn Val Ala
-5 1 5
Asp Val Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg
10 15 20
Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp
25 30 35
Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr
40 45 50 55
Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser
60 65 70
Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln
75 80 85
Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr
90 95 100
Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala
105 110 115
Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser
120 125 130 135
Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val
140 145 150
Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu
155 160 165
Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His
170 175 180
Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly
185 190 195

Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg
 200 205 210 215
 Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu
 220 225 230
 Val Pro Gly Ser Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln
 235 240 245
 Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg
 250 255 260
 Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu
 265 270 275
 Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn
 280 285 290 295
 Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro
 300 305 310
 Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp
 315 320 325
 Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp
 330 335 340
 Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln
 345 350 355
 Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala
 360 365 370 375
 Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu
 380 385 390
 Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala
 395 400 405
 Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr
 410 415 420
 Pro Phe Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp
 425 430 435
 Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala
 440 445 450 455
 Glu Val Val Ala Ser Leu Val Pro Ala Ala
 460 465

<210> 18
 <211> 2079
 <212> DNA
 <213> Unknown

<220>
 <223> GST:K:trAPAO 2079 nt. Translation starting at nt 1
 - 687, gst fusion + polylinker, 688-2076,
 K:trAPAO, extra lysine underlined; 2077-2079, stop
 codon. For bacterial expression.

<221> CDS
 <222> (1)...(2076)
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 <222> (1)...(687)
 <223> gst fusion + polylinker

<221> misc_feature
 <222> (688)...(2076)
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 <223> Extra lysine

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ctg	tcc	tgc	ctc	gtt	ctt	gag	gcg	atg	gat	cgt	gta	ggg	gga	aag	act	816	
Leu	Ser	Cys	Leu	Val	Leu	Glu	Ala	Met	Asp	Arg	Val	Gly	Gly	Lys	Thr		
				260					265					270			
ctg	agc	gta	caa	tcg	ggt	ccc	ggc	agg	acg	act	atc	aac	gac	ctc	ggc	864	
Leu	Ser	Val	Gln	Ser	Gly	Pro	Gly	Arg	Thr	Thr	Ile	Asn	Asp	Leu	Gly		
				275					280					285			
gct	gcg	tgg	atc	aat	gac	agc	aac	caa	agc	gaa	gta	tcc	aga	ttg	ttt	912	
Ala	Ala	Trp	Ile	Asn	Asp	Ser	Asn	Gln	Ser	Glu	Val	Ser	Arg	Leu	Phe		
				290					295					300			
gaa	aga	ttt	cat	ttg	gag	ggc	gag	ctc	cag	agg	acg	act	gga	aat	tca	960	
Glu	Arg	Phe	His	Leu	Glu	Gly	Glu	Leu	Gln	Arg	Thr	Thr	Gly	Asn	Ser		
				310					315					320			
atc	cat	caa	gca	caa	gac	ggt	aca	acc	act	aca	gct	cct	tat	ggt	gac	1008	
Ile	His	Gln	Ala	Gln	Asp	Gly	Thr	Thr	Thr	Thr	Ala	Pro	Tyr	Gly	Asp		
				325					330					335			
tcc	ttg	ctg	agc	gag	gag	gtt	gca	agt	gca	ctt	gcg	gaa	ctc	ctc	ccc	1056	
Ser	Leu	Leu	Ser	Glu	Glu	Val	Ala	Ser	Ala	Leu	Ala	Glu	Leu	Leu	Pro		
				340					345					350			
gta	tgg	tct	cag	ctg	atc	gaa	gag	cat	agc	ctt	caa	gac	ctc	aag	gcg	1104	
Val	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Gln	Asp	Leu	Lys	Ala		
				355					360					365			
agc	cct	cag	gcg	aag	cgg	ctc	gac	agt	gtg	agc	ttc	gcg	cac	tac	tgt	1152	
Ser	Pro	Gln	Ala	Lys	Arg	Leu	Asp	Ser	Val	Ser	Phe	Ala	His	Tyr	Cys		
				370					375					380			
gag	aag	gaa	cta	aac	ttg	cct	gct	gtt	ctc	ggc	gta	gca	aac	cag	atc	1200	
Glu	Lys	Glu	Leu	Asn	Leu	Pro	Ala	Val	Leu	Gly	Val	Ala	Asn	Gln	Ile		
				385					390					395			400
aca	cgc	gct	ctg	ctc	ggt	gtg	gaa	gcc	cac	gag	atc	agc	atg	ctt	ttt	1248	
Thr	Arg	Ala	Leu	Leu	Gly	Val	Glu	Ala	His	Glu	Ile	Ser	Met	Leu	Phe		
				405					410					415			
ctc	acc	gac	tac	atc	aag	agt	gcc	acc	ggt	ctc	agt	aat	att	ttc	tcg	1296	
Leu	Thr	Asp	Tyr	Ile	Lys	Ser	Ala	Thr	Gly	Leu	Ser	Asn	Ile	Phe	Ser		
				420					425					430			
gac	aag	aaa	gac	ggc	ggg	cag	tat	atg	cga	tgc	aaa	aca	ggt	atg	cag	1344	
Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys	Thr	Gly	Met	Gln		
				435					440					445			
tcg	att	tgc	cat	gcc	atg	tca	aag	gaa	ctt	gtt	cca	ggc	tca	gtg	cac	1392	
Ser	Ile	Cys	His	Ala	Met	Ser	Lys	Glu	Leu	Val	Pro	Gly	Ser	Val	His		
				450					455					460			
ctc	aac	acc	ccc	gtc	gct	gaa	att	gag	cag	tcg	gca	tcc	ggc	tgt	aca	1440	
Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	Glu	Gln	Ser	Ala						

Thr	Arg	Leu	Leu	Leu	Glu	Tyr	Leu	Glu	Glu	Lys	Tyr	Glu	Glu	His	Leu
			20					25					30		
Tyr	Glu	Arg	Asp	Glu	Gly	Asp	Lys	Trp	Arg	Asn	Lys	Lys	Phe	Glu	Leu
		35					40					45			
Gly	Leu	Glu	Phe	Pro	Asn	Leu	Pro	Tyr	Tyr	Ile	Asp	Gly	Asp	Val	Lys
	50					55					60				
Leu	Thr	Gln	Ser	Met	Ala	Ile	Ile	Arg	Tyr	Ile	Ala	Asp	Lys	His	Asn
65					70					75				80	
Met	Leu	Gly	Gly	Cys	Pro	Lys	Glu	Arg	Ala	Glu	Ile	Ser	Met	Leu	Glu
				85					90					95	
Gly	Ala	Val	Leu	Asp	Ile	Arg	Tyr	Gly	Val	Ser	Arg	Ile	Ala	Tyr	Ser
			100					105					110		
Lys	Asp	Phe	Glu	Thr	Leu	Lys	Val	Asp	Phe	Leu	Ser	Lys	Leu	Pro	Glu
		115					120					125			
Met	Leu	Lys	Met	Phe	Glu	Asp	Arg	Leu	Cys	His	Lys	Thr	Tyr	Leu	Asn
	130					135					140				
Gly	Asp	His	Val	Thr	His	Pro	Asp	Phe	Met	Leu	Tyr	Asp	Ala	Leu	Asp
145					150					155					160
Val	Val	Leu	Tyr	Met	Asp	Pro	Met	Cys	Leu	Asp	Ala	Phe	Pro	Lys	Leu
				165					170					175	
Val	Cys	Phe	Lys	Lys	Arg	Ile	Glu	Ala	Ile	Pro	Gln	Ile	Asp	Lys	Tyr
			180					185					190		
Leu	Lys	Ser	Ser	Lys	Tyr	Ile	Ala	Trp	Pro	Leu	Gln	Gly	Trp	Gln	Ala
		195					200					205			
Thr	Phe	Gly	Gly	Gly	Asp	His	Pro	Pro	Lys	Ser	Asp	Leu	Val	Pro	Arg
	210				215						220				
Gly	Ser	Pro	Glu	Phe	Lys	Asp	Asn	Val	Ala	Asp	Val	Val	Val	Val	Gly
225					230					235					240
Ala	Gly	Leu	Ser	Gly	Leu	Glu	Thr	Ala	Arg	Lys	Val	Gln	Ala	Ala	Gly
				245					250					255	
Leu	Ser	Cys	Leu	Val	Leu	Glu	Ala	Met	Asp	Arg	Val	Gly	Gly	Lys	Thr
			260					265					270		
Leu	Ser	Val	Gln	Ser	Gly	Pro	Gly	Arg	Thr	Thr	Ile	Asn	Asp	Leu	Gly
		275					280					285			
Ala	Ala	Trp	Ile	Asn	Asp	Ser	Asn	Gln	Ser	Glu	Val	Ser	Arg	Leu	Phe
	290					295					300				
Glu	Arg	Phe	His	Leu	Glu	Gly	Glu	Leu	Gln	Arg	Thr	Thr	Gly	Asn	Ser
305					310					315					320
Ile	His	Gln	Ala	Gln	Asp	Gly	Thr	Thr	Thr	Thr	Ala	Pro	Tyr	Gly	Asp
				325					330					335	
Ser	Leu	Leu	Ser	Glu	Glu	Val	Ala	Ser	Ala	Leu	Ala	Glu	Leu	Leu	Pro
			340					345					350		
Val	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Gln	Asp	Leu	Lys	Ala
		355					360					365			
Ser	Pro	Gln	Ala												

Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr
515 520 525
Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly
530 535 540
Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg
545 550 555 560
Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met
565 570 575
Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg
580 585 590
Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly
595 600 605
Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys
610 615 620
Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp
625 630 635 640
Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His
645 650 655
Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly
660 665 670
Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu
675 680 685
Val Pro Ala Ala
690

<210> 20
<211> 1464
<212> DNA
<213> Unknown

<220>
<223> Nucleotide sequence of K:trAPAO translational fusion with barley alpha amylase signal sequence, for expression and secretion of the mature trAPAO in maize. Nucleotides 1-72, barley alpha amylase signal sequence, nucleotides 73-75, added lysine residue; nucleotides 76 -1464 , trAPAO cDNA.

<221> sig_peptide
<222> (1)...(72)
<223> Barley alpha amylase signal sequence

<221> misc_feature
<222> (73)...(1464)
<223> K:trAPAOcDNA

<221> CDS
<222> (1)...(1461)

<221> misc_feature
<222> (73)...(75)
<223> Added lysine residue

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Met Ala Asn Lys His Leu Ser Leu Ser Leu Phe Leu Val Leu Leu Gly
-20 -15 -10
ctc tcc gcc tcc ctc gcc agc ggc aaa gac aac gtt gcg gac gtg gta 96
Leu Ser Ala Ser Leu Ala Ser Gly Lys Asp Asn Val Ala Asp Val Val
-5 1 5

gtg Val	gtg Val	ggc Gly	gct Ala	ggc Gly	ttg Leu	agc Ser	ggg Gly	ttg Leu	gag Glu	acg Thr	gca Ala	cgc Arg	aaa Lys	gtc Val	cag Gln	144			
10					15					20									
gcc Ala	gcc Ala	ggt Gly	ctg Leu	tcc Ser	tgc Cys	ctc Leu	gtt Val	ctt Leu	gag Glu	gcg Ala	atg Met	gat Asp	cgt Arg	gta Val	ggg Gly	192			
25					30					35					40				
gga Gly	aag Lys	act Thr	ctg Leu	agc Ser	gta Val	caa Gln	tcg Ser	ggg Gly	ccc Pro	ggc Gly	agg Arg	acg Thr	act Thr	atc Ile	aac Asn	240			
45					50					55									
gac Asp	ctc Leu	ggc Gly	gct Ala	gcg Ala	tgg Trp	atc Ile	aat Asn	gac Asp	agc Ser	aac Asn	caa Gln	agc Ser	gaa Glu	gta Val	tcc Ser	288			
60					65					70									
aga Arg	ttg Leu	ttt Phe	gaa Glu	aga Arg	ttt Phe	cat His	ttg Leu	gag Glu	ggc Gly	gag Glu	ctc Leu	cag Gln	agg Arg	acg Thr	act Thr	336			
75					80					85									
gga Gly	aat Asn	tca Ser	atc Ile	cat His	caa Gln	gca Ala	caa Gln	gac Asp	ggg Gly	aca Thr	acc Thr	act Thr	aca Thr	gct Ala	cct Pro	384			
90					95					100									
tat Tyr	ggg Gly	gac Asp	tcc Ser	ttg Leu	ctg Leu	agc Ser	gag Glu	gag Glu	gtt Val	gca Ala	agt Ser	gca Ala	ctt Leu	gcg Ala	gaa Glu	432			
105					110					115					120				
ctc Leu	ctc Leu	ccc Pro	gta Val	tgg Trp	tct Ser	cag Gln	ctg Leu	atc Ile	gaa Glu	gag Glu	cat His	agc Ser	ctt Leu	caa Gln	gac Asp	480			
125					130					135									
ctc Leu	aag Lys	gcg Ala	agc Ser	cct Pro	cag Gln	gcg Ala	aag Lys	cgg Arg	ctc Leu	gac Asp	agt Ser	gtg Val	agc Ser	ttc Phe	gcg Ala	528			
140					145					150									
cac His	tac Tyr	tgt Cys	gag Glu	aag Lys	gaa Glu	cta Leu	aac Asn	ttg Leu	cct Pro	gct Ala	gtt Val	ctc Leu	ggc Gly	gta Val	gca Ala	576			
155					160					165									
aac Asn	cag Gln	atc Ile	aca Thr	cgc Arg	gct Ala	ctg Leu	ctc Leu	ggg Gly	gtg Val	gaa Glu	gcc Ala	cac His	gag Glu	atc Ile	agc Ser	624			
170					175					180									
atg Met	ctt Leu	ttt Phe	ctc Leu	acc Thr	gac Asp	tac Tyr	atc Ile	aag Lys	agt Ser	gcc Ala	acc Thr	ggg Gly	ctc Leu	agt Ser	aat Asn	672			
185					190					195					200				
att Ile	ttc Phe	tcg Ser	gac Asp	aag Lys	aaa Lys	gac Asp	ggc Gly	ggg Gly	cag Gln	tat Tyr	atg Met	cga Arg	tgc Cys	aaa Lys	aca Thr	720			
205					210					215									
ggg Gly	atg Met	cag Gln	tcg Ser	att Ile	tgc Cys	cat His	gcc Ala	atg Met	tca Ser	aag Lys	gaa Glu	ctt Leu	gtt Val	cca Pro	ggc Gly	768			
220					225					230									
tca Ser	gtg Val	cac His	ctc Leu	aac Asn	acc Thr	ccc Pro	gtc Val	gct Ala	gaa Glu	att Ile	gag Glu	cag Gln	tcg Ser	gca Ala	tcc Ser	816			
235					240					245									
ggc Gly	tgt Cys	aca Glu	gta Val	cga Arg	tcg Ser	gcc Ala	tcg Ser	ggc Gly	gcc Ala	gtg Val	ttc Glu	cga Arg	agc Ser	aaa Lys	aag Glu	864			

[illegible]

<220>
 <221> SIGNAL
 <222> (1)...(24)

<400> 21

Met	Ala	Asn	Lys	His	Leu	Ser	Leu	Ser	Leu	Phe	Leu	Val	Leu	Leu	Gly	
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Leu	Ser	Ala	Ser	Leu	Ala	Ser	Gly	Lys	Asp	Asn	Val	Ala	Asp	Val	Val	
			-5					1				5				
Val	Val	Gly	Ala	Gly	Leu	Ser	Gly	Leu	Glu	Thr	Ala	Arg	Lys	Val	Gln	
	10				15						20					
Ala	Ala	Gly	Leu	Ser	Cys	Leu	Val	Leu	Glu	Ala	Met	Asp	Arg	Val	Gly	
25					30					35					40	
Gly	Lys	Thr	Leu	Ser	Val	Gln	Ser	Gly	Pro	Gly	Arg	Thr	Thr	Ile	Asn	
				45					50					55		
Asp	Leu	Gly	Ala	Ala	Trp	Ile	Asn	Asp	Ser	Asn	Gln	Ser	Glu	Val	Ser	
			60					65					70			
Arg	Leu	Phe	Glu	Arg	Phe	His	Leu	Glu	Gly	Glu	Leu	Gln	Arg	Thr	Thr	
		75				80						85				
Gly	Asn	Ser	Ile	His	Gln	Ala	Gln	Asp	Gly	Thr	Thr	Thr	Thr	Ala	Pro	
90					95						100					
Tyr	Gly	Asp	Ser	Leu	Leu	Ser	Glu	Glu	Val	Ala	Ser	Ala	Leu	Ala	Glu	
105					110					115					120	
Leu	Leu	Pro	Val	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Gln	Asp	
				125					130					135		
Leu	Lys	Ala	Ser	Pro	Gln	Ala	Lys	Arg	Leu	Asp	Ser	Val	Ser	Phe	Ala	
			140				145					150				
His	Tyr	Cys	Glu	Lys	Glu	Leu	Asn	Leu	Pro	Ala	Val	Leu	Gly	Val	Ala	
		155				160					165					
Asn	Gln	Ile	Thr	Arg	Ala	Leu	Leu	Gly	Val	Glu	Ala	His	Glu	Ile	Ser	
170					175					180						
Met	Leu	Phe	Leu	Thr	Asp	Tyr	Ile	Lys	Ser	Ala	Thr	Gly	Leu	Ser	Asn	
185				190						195					200	
Ile	Phe	Ser	Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys	Thr	
				205					210					215		
Gly	Met	Gln	Ser	Ile	Cys	His	Ala	Met	Ser	Lys	Glu	Leu	Val	Pro	Gly	
			220					225					230			
Ser	Val	His	Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	Glu	Gln	Ser	Ala	Ser	
		235				240						245				
Gly	Cys	Thr	Val	Arg	Ser	Ala	Ser	Gly	Ala	Val	Phe	Arg	Ser	Lys	Lys	
	250					255				260						
Val	Val	Val	Ser	Leu	Pro	Thr	Thr	Leu	Tyr	Pro	Thr	Leu	Thr	Phe	Ser	
265				270					275						280	
Pro	Pro	Leu	Pro	Ala	Glu	Lys	Gln	Ala	Leu	Ala	Glu	Asn	Ser	Ile	Leu	
				285				290						295		
Gly	Tyr	Tyr	Ser	Lys	Ile	Val	Phe	Val	Trp	Asp	Lys	Pro	Trp	Trp	Arg	
			300				305						310			
Glu	Gln	Gly	Phe	Ser	Gly	Val	Leu	Gln	Ser	Ser	Cys	Asp	Pro	Ile	Ser	
		315				320						325				
Phe	Ala	Arg	Asp	Thr	Ser	Ile	Asp	Val	Asp	Arg	Gln	Trp	Ser	Ile	Thr	
		330			335						340					
Cys	Phe	Met	Val	Gly	Asp	Pro	Gly	Arg	Lys	Trp	Ser	Gln	Gln	Ser	Lys	
345				350				355							360	
Gln	Val	Arg	Gln	Lys	Ser	Val	Trp	Asp	Gln	Leu	Arg	Ala	Ala	Tyr	Glu	
				365				370						375		
Asn	Ala	Gly	Ala	Gln	Val	Pro	Glu	Pro	Ala	Asn	Val	Leu	Glu	Ile	Glu	
			380				385						390			
Trp	Ser	Lys	Gln	Gln	Tyr	Phe	Gln	Gly	Ala	Pro	Ser	Ala	Val	Tyr	Gly	
		395				400						405				
Leu	Asn	Asp	Leu	Ile	Thr	Leu	Gly	Ser	Ala	Leu	Arg	Thr	Pro	Phe	Lys	
	410				415					420						
Ser	Val	His	Phe	Val	Gly	Thr	Glu	Thr	Ser	Leu	Val	Trp	Lys	Gly	Tyr	

			180				185				190							
aac	gac	ctc	ggc	gct	gcg	tgg	atc	aat	gac	agc	aac	caa	agc	gaa	gta	624		
Asn	Asp	Leu	Gly	Ala	Ala	Trp	Ile	Asn	Asp	Ser	Asn	Gln	Ser	Glu	Val			
			195				200				205							
tcc	aga	ttg	ttt	gaa	aga	ttt	cat	ttg	gag	ggc	gag	ctc	cag	agg	acg	672		
Ser	Arg	Leu	Phe	Glu	Arg	Phe	His	Leu	Glu	Gly	Glu	Leu	Gln	Arg	Thr			
			210				215				220							
act	gga	aat	tca	atc	cat	caa	gca	caa	gac	ggc	aca	acc	act	aca	gct	720		
Thr	Gly	Asn	Ser	Ile	His	Gln	Ala	Gln	Asp	Gly	Thr	Thr	Thr	Thr	Ala			
			225				230				235				240			
cct	tat	ggc	gac	tcc	ttg	ctg	agc	gag	gag	ggt	gca	agt	gca	ctt	gcg	768		
Pro	Tyr	Gly	Asp	Ser	Leu	Leu	Ser	Glu	Glu	Val	Ala	Ser	Ala	Leu	Ala			
			245				250				255							
gaa	ctc	ctc	ccc	gta	tgg	tct	cag	ctg	atc	gaa	gag	cat	agc	ctt	caa	816		
Glu	Leu	Leu	Pro	Val	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Gln			
			260				265				270							
gac	ctc	aag	gcg	agc	cct	cag	gcg	aag	cgg	ctc	gac	agt	gtg	agc	ttc	864		
Asp	Leu	Lys	Ala	Ser	Pro	Gln	Ala	Lys	Arg	Leu	Asp	Ser	Val	Ser	Phe			
			275				280				285							
gcg	cac	tac	tgt	gag	aag	gaa	cta	aac	ttg	cct	gct	ggt	ctc	ggc	gta	912		
Ala	His	Tyr	Cys	Glu	Lys	Glu	Leu	Asn	Leu	Pro	Ala	Val	Leu	Gly	Val			
			290				295				300							
gca	aac	cag	atc	aca	cgc	gct	ctg	ctc	ggc	gtg	gaa	gcc	cac	gag	atc	960		
Ala	Asn	Gln	Ile	Thr	Arg	Ala	Leu	Leu	Gly	Val	Glu	Ala	His	Glu	Ile			
			305				310				315				320			
agc	atg	ctt	ttt	ctc	acc	gac	tac	atc	aag	agt	gcc	acc	ggc	ctc	agt	1008		
Ser	Met	Leu	Phe	Leu	Thr	Asp	Tyr	Ile	Lys	Ser	Ala	Thr	Gly	Leu	Ser			
			325				330				335							
aat	att	ttc	tcg	gac	aag	aaa	gac	ggc	ggg	cag	tat	atg	cga	tgc	aaa	1056		
Asn	Ile	Phe	Ser	Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys			
			340				345				350							
aca	ggc	atg	cag	tcg	att	tgc	cat	gcc	atg	tca	aag	gaa	ctt	ggt	cca	1104		
Thr	Gly	Met	Gln	Ser	Ile	Cys	His	Ala	Met	Ser	Lys	Glu	Leu	Val	Pro			
			355				360				365							
ggc	tca	gtg	cac	ctc	aac	acc	ccc	gtc	gct	gaa	att	gag	cag	tcg	gca	1152		
Gly	Ser	Val	His	Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	Glu	Gln	Ser	Ala			
			370				375				380							
tcc	ggc	tgt	aca	gta	cga	tcg	gcc	tcg	ggc	gcc	gtg	ttc	cga	agc	aaa	1200		
Ser	Gly	Cys	Thr	Val	Arg	Ser	Ala	Ser	Gly	Ala	Val	Phe	Arg	Ser	Lys			
			385				390				395				400			
aag	gtg	gtg	ggt	tcg	tta	ccg	aca	acc	ttg	tat	ccc	acc	ttg	aca	ttt	1248		
Lys	Val	Val	Val	Ser	Leu	Pro	Thr	Thr	Leu	Tyr	Pro	Thr	Leu	Thr	Phe			
			405				4											

ctg ggc tac tat agc aag ata gtc ttc gta tgg gac aag ccg tgg tgg	1344
Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp	
435 440 445	
cgc gaa caa ggc ttc tcg ggc gtc ctc caa tcg agc tgt gac ccc atc	1392
Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile	
450 455 460	
tca ttt gcc aga gat acc agc atc gac gtc gat cga caa tgg tcc att	1440
Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile	
465 470 475 480	
acc tgt ttc atg gtc gga gac ccg gga cgg aag tgg tcc caa cag tcc	1488
Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser	
485 490 495	
aag cag gta cga caa aag tct gtc tgg gac caa ctc cgc gca gcc tac	1536
Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr	
500 505 510	
gag aac gcc ggg gcc caa gtc cca gag ccg gcc aac gtg ctc gaa atc	1584
Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile	
515 520 525	
gag tgg tcg aag cag cag tat ttc caa gga gct ccg agc gcc gtc tat	1632
Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr	
530 535 540	
ggg ctg aac gat ctc atc aca ctg ggt tcg gcg ctc aga acg ccg ttc	1680
Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe	
545 550 555 560	
aag agt gtt cat ttc gtt gga acg gag acg tct tta gtt tgg aaa ggg	1728
Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly	
565 570 575	
tat atg gaa ggg gcc ata cga tcg ggt caa cga ggt gct gca gaa gtt	1776
Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val	
580 585 590	
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Val Ala Ser Leu Val Pro Ala Ala	
595 600	

<210> 23

<211> 600

<212> PRT

<213> *Exophiala spinifera*

<400> 23

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Ala	Gly	Tyr	Ser	His	Val	Gly	Val	Gly	Pro	Asp	Gly	Gly	Arg	Tyr	Val
			20					25					30		
Thr	Ile	Ala	Gly	Gln	Ile	Gly	Gln	Asp	Ala	Ser	Gly	Val	Thr	Asp	Pro
		35				40						45			
Ala	Tyr	Glu	Lys	Gln	Val	Ala	Gln	Ala	Phe	Ala	Asn	Leu	Arg	Ala	Cys
		50				55				60					
Leu	Ala	Ala	Val	Gly	Ala	Thr	Ser	Asn	Asp	Val	Thr	Lys	Leu	Asn	Tyr
65					70					75				80	
Tyr	Ile	Val	Asp	Tyr	Ala	Pro	Ser	Lys	Leu	Thr	Ala	Ile	Gly	Asp	Gly

Val Ala Ser Leu Val Pro Ala Ala
 580 585 590
 595 600

<210> 24
 <211> 3003
 <212> DNA
 <213> Unknown

<220>
 <223> Sequence is barley alpha amylase signal sequence:
 esp1 mat: an artificial spacer sequence and
 K:trAPAO

<221> sig_peptide
 <222> (1)...(72)
 <223> Barley alpha amylase signal sequence

<221> misc_feature
 <222> (73)...(1575)
 <223> esp1 mat

<221> misc_feature
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 <223> spacer sequence

<221> misc_feature
 <222> (1612)...(3000)
 <223> K:trAPAO

<221> CDS
 <222> (1)...(3000)

<221> misc_feature
 <222> (1612)...(1614)
 <223> Extra lysine

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 -20 -15 -10
 ctc tcc gcc tcc ctc gcc agc ggc gct cct act gtc aag att gat gct 96
 Leu Ser Ala Ser Leu Ala Ser Gly Ala Pro Thr Val Lys Ile Asp Ala
 -5 1 5
 ggg atg gtg gtc ggc acg act act act gtc ccc ggc acc act gcg acc 144
 Gly Met Val Val Gly Thr Thr Thr Thr Val Pro Gly Thr Thr Ala Thr
 10 15 20
 gtc agc gag ttc ttg ggc gtt cct ttt gcc gcc tct ccg aca cga ttt 192
 Val Ser Glu Phe Leu Gly Val Pro Phe Ala Ala Ser Pro Thr Arg Phe
 25 30 35 40
 gcg cct cct act cgt ccc gtg cct tgg tca acg cct ttg caa gcc act 240
 Ala Pro Pro Thr Arg Pro Val Pro Trp Ser Thr Pro Leu Gln Ala Thr
 45 50 55
 gca tat ggt cca gca tgc cct caa caa ttc aat tac ccc gaa gaa ctc 288
 Ala Tyr Gly Pro Ala Cys Pro Gln Gln Phe Asn Tyr Pro Glu Glu Leu
 60 65 70

cgt gag att acg atg gcc tgg ttc aat aca ccg ccc ccg tca gct ggt	336
Arg Glu Ile Thr Met Ala Trp Phe Asn Thr Pro Pro Pro Ser Ala Gly	
75 80 85	
gaa agt gag gac tgc ctg aac ctc aac atc tac gtc cca gga act gag	384
Glu Ser Glu Asp Cys Leu Asn Leu Asn Ile Tyr Val Pro Gly Thr Glu	
90 95 100	
aac aca aac aaa gcc gtc atg gtt tgg ata tac ggt gga gcg ctg gaa	432
Asn Thr Asn Lys Ala Val Met Val Trp Ile Tyr Gly Gly Ala Leu Glu	
105 110 115 120	
tat ggt tgg aat tca ttc cac ctt tac gac ggg gct agt ttc gca gcc	480
Tyr Gly Trp Asn Ser Phe His Leu Tyr Asp Gly Ala Ser Phe Ala Ala	
125 130 135	
aat cag gat gtc atc gcc gtg acc atc aac tac aga acg aac att ctg	528
Asn Gln Asp Val Ile Ala Val Thr Ile Asn Tyr Arg Thr Asn Ile Leu	
140 145 150	
ggg ttc cct gct gcc cct cag ctt cca ata aca cag cga aat ctg ggg	576
Gly Phe Pro Ala Ala Pro Gln Leu Pro Ile Thr Gln Arg Asn Leu Gly	
155 160 165	
ttc cta gac caa agg ttt gct ttg gat tgg gta cag cgg aac atc gca	624
Phe Leu Asp Gln Arg Phe Ala Leu Asp Trp Val Gln Arg Asn Ile Ala	
170 175 180	
gcc ttt ggc ggt gat cct cga aag gtc aca ata ttt ggg cag agt gcg	672
Ala Phe Gly Gly Asp Pro Arg Lys Val Thr Ile Phe Gly Gln Ser Ala	
185 190 195 200	
ggg ggc aga agt gtc gac gtc ctc ttg acg tct atg cca cac aac cca	720
Gly Gly Arg Ser Val Asp Val Leu Leu Thr Ser Met Pro His Asn Pro	
205 210 215	
ccc ttc cga gca gca atc atg gag tcc ggt gtg gct aac tac aac ttc	768
Pro Phe Arg Ala Ala Ile Met Glu Ser Gly Val Ala Asn Tyr Asn Phe	
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Tyr Thr Leu Asp Asn Val Thr Ala Val Tyr Arg Ser Glu Thr Ala Arg	
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gcc tat tct acg ggc aag tcg acc atg aca ttc ggt ccc gag ggc cgc Ala Tyr Ser Thr Gly Lys Ser Thr Met Thr Phe Gly Pro Glu Gly Arg 460 465 470	1488
gcg gcg gtg gtg tcg ccc gga cct tcc atc ccc cct tgc gcg gat ggc Ala Ala Val Val Ser Pro Gly Pro Ser Ile Pro Pro Cys Ala Asp Gly 475 480 485	1536
gcc aag gcg ggg ggc gga ggc agc ggc gga ggc agc ggc gga ggc agc Ala Lys Ala Gly Gly Gly Gly Ser Gly Gly Gly Ser Gly Gly Ser 490 495 500	1584
aaa gac aac gtt gcg gac gtg gta gtg gtg ggc gct ggc ttg agc ggt Lys Asp Asn Val Ala Asp Val Val Val Val Gly Ala Gly Leu Ser Gly 505 510 515 520	1632
ttg gag acg gca cgc aaa gtc cag gcc gcc ggt ctg tcc tgc ctc gtt Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val 525 530 535	1680
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ggt ccc ggc agg acg act atc aac gac ctc ggc gct gcg tgg atc aat Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn 555 560 565	1776
gac agc aac caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu 570 575 580	1824
gag ggc gag ctc cag agg acg act gga aat tca atc cat caa gca caa Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln	1872

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gac Asp	ggt Gly	aca Thr	acc Thr	act Thr	aca Thr	gct Ala	cct Pro	tat Tyr	ggt Gly	gac Asp	tcc Ser	ttg Leu	ctg Leu	agc Ser	gag Glu	1920	
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gag Glu	gtt Val	gca Ala	agt Ser	gca Ala	ctt Leu	gcg Ala	gaa Glu	ctc Leu	ctc Leu	ccc Pro	gta Val	tgg Trp	tct Ser	cag Gln	ctg Leu	1968	
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atc Ile	gaa Glu	gag Glu	cat His	agc Ser	ctt Leu	caa Gln	gac Asp	ctc Leu	aag Lys	gcg Ala	agc Ser	cct Pro	cag Gln	gcg Ala	aag Lys	2016	
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cgg Arg	ctc Leu	gac Asp	agt Ser	gtg Val	agc Ser	ttc Phe	gcg Ala	cac His	tac Tyr	tgt Cys	gag Glu	aag Lys	gaa Glu	cta Leu	aac Asn	2064	
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ggt Gly	gtg Val	gaa Glu	gcc Ala	cac His	gag Glu	atc Ile	agc Ser	atg Met	ctt Leu	ttt Phe	ctc Leu	acc Thr	gac Asp	tac Tyr	atc Ile	2160	
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aag Lys	agt Ser	gcc Ala	acc Thr	ggt Gly	ctc Leu	agt Ser	aat Asn	att Ile	ttc Phe	tcg Ser	gac Asp	aag Lys	aaa Lys	gac Asp	ggc Gly	2208	
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ggg Gly	cag Gln	tat Tyr	atg Met	cga Arg	tgc Cys	aaa Lys	aca Thr	ggt Gly	atg Met	cag Gln	tcg Ser	att Ile	tgc Cys	cat His	gcc Ala	2256	
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atg Met	tca Ser	aag Lys	gaa Glu	ctt Leu	gtt Val	cca Pro	ggc Gly	tca Ser	gtg Val	cac His	ctc Leu	aac Asn	acc Thr	ccc Pro	gtc Val	2304	
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gct Ala	gaa Glu	att Ile	gag Glu	cag Gln	tcg Ser	gca Ala	tcc Ser	ggc Gly	tgt Cys	aca Thr	gta Val	cga Arg	tcg Ser	gcc Ala	tcg Ser	2352	
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ggc Gly	gcc Ala	gtg Val	ttc Phe	cga Arg	agc Ser	aaa Lys	aag Lys	gtg Val	gtg Val	gtt Val	tcg Ser	tta Leu	ccg Pro	aca Thr	acc Thr	2400	
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ttg Leu	tat Tyr	ccc Pro	acc Thr	ttg Leu	aca Thr	ttt Phe	tca Ser	cca Pro	cct Pro	ctt Leu	ccc Pro	gcc Ala	gag Glu	aag Lys	caa Gln	2448	
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gca Ala	ttg Leu	gcg Ala	gaa Glu	aat Asn	tct Ser	atc Ile	ctg Leu	ggc Gly	tac Tyr	tat Tyr	agc Ser	aag Lys	ata Ile	gtc Val	ttc Phe	2496	
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caa Gln	tcg Ser	agc Ser	tgt Cys	gac Asp	ccc Pro	atc Ile	tca Ser	ttt Phe	gcc Ala	aga Arg	gat Asp	acc Thr	agc Ser	atc Ile	gac Asp	2592	
				825				830				835				840	

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Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly	
845 850 855	
cgg aag tgg tcc caa cag tcc aag cag gta cga caa aag tct gtc tgg	2688
Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp	
860 865 870	
gac caa ctc cgc gca gcc tac gag aac gcc ggg gcc caa gtc cca gag	2736
Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu	
875 880 885	
ccg gcc aac gtg ctc gaa atc gag tgg tcg aag cag cag tat ttc caa	2784
Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln	
890 895 900	
gga gct ccg agc gcc gtc tat ggg ctg aac gat ctc atc aca ctg ggt	2832
Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly	
905 910 915 920	
tcg gcg ctc aga acg ccg ttc aag agt gtt cat ttc gtt gga acg gag	2880
Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu	
925 930 935	
acg tct tta gtt tgg aaa ggg tat atg gaa ggg gcc ata cga tcg ggt	2928
Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly	
940 945 950	
caa cga ggt gct gca gaa gtt gtg gct agc ctg gtg cca gca gca	2973
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Gly Ile Pro Tyr Ala Ala Pro Pro Val Gly Gly Leu Arg Trp Lys Pro	
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Pro Gln His Ala Arg Pro Trp Ala Gly Val Arg Pro Ala Thr Gln Phe	
45 50 55	
Gly Ser Asp Cys Phe Gly Ala Ala Tyr Leu Arg Lys Gly Ser Leu Ala	
60 65 70	
Pro Gly Val Ser Glu Asp Cys Leu Tyr Leu Asn Val Trp Ala Pro Ser	
75 80 85	
Gly Ala Lys Pro Gly Gln Tyr Pro Val Met Val Trp Val Tyr Gly Gly	
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<223> esp1 mat

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<223> spacer sequence

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<223> K:trAPAO

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<222> (2227)...(2229)

<223> Extra lysine

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act	cga	ctt	ctt	ttg	gaa	tat	ctt	gaa	gaa	aaa	tat	gaa	gag	cat	ttg	96
Thr	Arg	Leu	Leu	Leu	Glu	Tyr	Leu	Glu	Glu	Lys	Tyr	Glu	Glu	His	Leu	
			20					25					30			

tat	gag	cgc	gat	gaa	ggt	gat	aaa	tgg	cga	aac	aaa	aag	ttt	gaa	ttg	144
Tyr	Glu	Arg	Asp	Glu	Gly	Asp	Lys	Trp	Arg	Asn	Lys	Lys	Phe	Glu	Leu	
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ggt	ttg	gag	ttt	ccc	aat	ctt	cct	tat	tat	att	gat	ggt	gat	ggt	aaa	192
Gly	Leu	Glu	Phe	Pro	Asn	Leu	Pro	Tyr	Tyr	Ile	Asp	Gly	Asp	Val	Lys	
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Leu	Thr	Gln	Ser	Met	Ala	Ile	Ile	Arg	Tyr	Ile	Ala	Asp	Lys	His	Asn	
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atg	ttg	ggt	ggt	tgt	cca	aaa	gag	cgt	gca	gag	att	tca	atg	ctt	gaa	288
Met	Leu	Gly	Gly	Cys	Pro	Lys	Glu	Arg	Ala	Glu	Ile	Ser	Met	Leu	Glu	
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gga	gcg	ggt	ttg	gat	att	aga	tac	ggt	ggt	tcg	aga	att	gca	tat	agt	336
Gly	Ala	Val	Leu	Asp	Ile	Arg	Tyr	Gly	Val	Ser	Arg	Ile	Ala	Tyr	Ser	
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aaa	gac	ttt	gaa	act	ctc	aaa	ggt	gat	ttt	ctt	agc	aag	cta	cct	gaa	384
Lys	Asp	Phe	Glu	Thr	Leu	Lys	Val	Asp	Phe	Leu	Ser	Lys	Leu	Pro	Glu	
		115					120					125				

atg	ctg	aaa	atg	ttc	gaa	gat	cgt	tta	tgt	cat	aaa	aca	tat	tta	aat	432
Met	Leu	Lys	Met	Phe	Glu	Asp	Arg	Leu	Cys	His	Lys	Thr	Tyr	Leu	Asn	
	130					135					140					

ggt	gat	cat	gta	acc	cat	cct	gac	ttc	atg	ttg	tat	gac	gct	ctt	gat	480
Gly	Asp	His	Val	Thr	His	Pro	Asp	Phe	Met	Leu	Tyr	Asp	Ala	Leu	Asp	
145					150					155					160	

ggt	ggt	tta	tac	atg	gac	cca	atg	tgc	ctg	gat	gcg	ttc	cca	aaa	tta	528
Val	Val	Leu	Tyr	Met	Asp	Pro	Met	Cys	Leu	Asp	Ala	Phe	Pro	Lys	Leu	
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gtt tgt ttt aaa aaa cgt att gaa gct atc cca caa att gat aag tac	576
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr	
180 185 190	
ttg aaa tcc agc aag tat ata gca tgg cct ttg cag ggc tgg caa gcc	624
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala	
195 200 205	
acg ttt ggt ggt ggc gac cat cct cca aaa tcg gat ctg gtt ccg cgt	672
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg	
210 215 220	
gga tcc ccg gaa ttc gct cct act gtc aag att gat gct ggg atg gtg	720
Gly Ser Pro Glu Phe Ala Pro Thr Val Lys Ile Asp Ala Gly Met Val	
225 230 235 240	
gtc ggc acg act act act gtc ccc ggc acc act gcg acc gtc agc gag	768
Val Gly Thr Thr Thr Thr Val Pro Gly Thr Thr Ala Thr Val Ser Glu	
245 250 255	
ttc ttg ggc gtt cct ttt gcc gcc tct ccg aca cga ttt gcg cct cct	816
Phe Leu Gly Val Pro Phe Ala Ala Ser Pro Thr Arg Phe Ala Pro Pro	
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act cgt ccc gtg cct tgg tca acg cct ttg caa gcc act gca tat ggt	864
Thr Arg Pro Val Pro Trp Ser Thr Pro Leu Gln Ala Thr Ala Tyr Gly	
275 280 285	
cca gca tgc cct caa caa ttc aat tac ccc gaa gaa ctc cgt gag att	912
Pro Ala Cys Pro Gln Gln Phe Asn Tyr Pro Glu Glu Leu Arg Glu Ile	
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acg atg gcc tgg ttc aat aca ccg ccc ccg tca gct ggt gaa agt gag	960
Thr Met Ala Trp Phe Asn Thr Pro Pro Pro Ser Ala Gly Glu Ser Glu	
305 310 315 320	
gac tgc ctg aac ctc aac atc tac gtc cca gga act gag aac aca aac	1008
Asp Cys Leu Asn Leu Asn Ile Tyr Val Pro Gly Thr Glu Asn Thr Asn	
325 330 335	
aaa gcc gtc atg gtt tgg ata tac ggt gga gcg ctg gaa tat ggt tgg	1056
Lys Ala Val Met Val Trp Ile Tyr Gly Gly Ala Leu Glu Tyr Gly Trp	
340 345 350	
aat tca ttc cac ctt tac gac ggg gct agt ttc gca gcc aat cag gat	1104
Asn Ser Phe His Leu Tyr Asp Gly Ala Ser Phe Ala Ala Asn Gln Asp	
355 360 365	
gtc atc gcc gtg acc atc aac tac aga acg aac att ctg ggg ttc cct	1152
Val Ile Ala Val Thr Ile Asn Tyr Arg Thr Asn Ile Leu Gly Phe Pro	
370 375 380	
gct gcc cct cag ctt cca ata aca cag cga aat ctg ggg ttc cta gac	1200
Ala Ala Pro Gln Leu Pro Ile Thr Gln Arg Asn Leu Gly Phe Leu Asp	
385 390 395 400	
caa agg ttt gct ttg gat tgg gta cag cgg aac atc gca gcc ttt ggc	1248
Gln Arg Phe Ala Leu Asp Trp Val Gln Arg Asn Ile Ala Ala Phe Gly	
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[illegible]

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Phe	Leu	Thr	Asp	Tyr	Ile	Lys	Ser	Ala	Thr	Gly	Leu	Ser	Asn	Ile	Phe	
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tcg	gac	aag	aaa	gac	ggc	ggg	cag	tat	atg	cga	tgc	aaa	aca	ggg	atg	2880
Ser	Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys	Thr	Gly	Met	
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cag	tcg	att	tgc	cat	gcc	atg	tca	aag	gaa	ctt	gtt	cca	ggc	tca	gtg	2928
Gln	Ser	Ile	Cys	His	Ala	Met	Ser	Lys	Glu	Leu	Val	Pro	Gly	Ser	Val	
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cac	ctc	aac	acc	ccc	gtc	gct	gaa	att	gag	cag	tcg	gca	tcc	ggc	tgt	2976
His	Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	Glu	Gln	Ser	Ala	Ser	Gly	Cys	
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aca	gta	cga	tcg	gcc	tcg	ggc	gcc	gtg	ttc	cga	agc	aaa	aag	gtg	gtg	3024
Thr	Val	Arg	Ser	Ala	Ser	Gly	Ala	Val	Phe	Arg	Ser	Lys	Lys	Val	Val	
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gtt	tcg	tta	ccg	aca	acc	ttg	tat	ccc	acc	ttg	aca	ttt	tca	cca	cct	3072
Val	Ser	Leu	Pro	Thr	Thr	Leu	Tyr	Pro	Thr	Leu	Thr	Phe	Ser	Pro	Pro	
	1010					1015					1020					
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Leu	Pro	Ala	Glu	Lys	Gln	Ala	Leu	Ala	Glu	Asn	Ser	Ile	Leu	Gly	Tyr	
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tat	agc	aag	ata	gtc	ttc	gta	tgg	gac	aag	ccg	tgg	tgg	cgc	gaa	caa	3168
Tyr	Ser	Lys	Ile	Val	Phe	Val	Trp	Asp	Lys	Pro	Trp	Trp	Arg	Glu	Gln	
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Gly	Phe	Ser	Gly	Val	Leu	Gln	Ser	Ser	Cys	Asp	Pro	Ile	Ser	Phe	Ala	
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aga	gat	acc	agc	atc	gac	gtc	gat	cga	caa	tgg	tcc	att	acc	tgt	ttc	3264
Arg	Asp	Thr	Ser	Ile	Asp	Val	Asp	Arg	Gln	Trp	Ser	Ile	Thr	Cys	Phe	
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atg	gtc	gga	gac	ccg	gga	cgg	aag	tgg	tcc	caa	cag	tcc	aag	cag	gta	3312
Met	Val	Gly	Asp	Pro	Gly	Arg	Lys	Trp	Ser	Gln	Gln	Ser	Lys	Gln	Val	
	1090					1095				1100						
cga	caa	aag	tct	gtc	tgg	gac	caa	ctc	cgc	gca	gcc	tac	gag	aac	gcc	3360
Arg	Gln	Lys	Ser	Val	Trp	Asp	Gln	Leu	Arg	Ala	Ala	Tyr	Glu	Asn	Ala	
1105					1110					1115					1120	
ggg	gcc	caa	gtc	cca	gag	ccg	gcc	aac	gtg	ctc	gaa	atc	gag	tgg	tcg	3408
Gly	Ala	Gln	Val	Pro	Glu	Pro	Ala	Asn	Val	Leu	Glu	Ile	Glu	Trp	Ser	
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Lys	Gln	Gln	Tyr	Phe	Gln	Gly	Ala	Pro	Ser	Ala	Val	Tyr	Gly	Leu	Asn	
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gat	ctc	atc	aca	ctg	ggt	tcg	gcg	ctc	aga	acg	ccg	ttc	aag	agt	gtt	3504
Asp	Leu	Ile	Thr	Leu	Gly	Ser	Ala	Leu	Arg	Thr	Pro	Phe	Lys	Ser	Val	
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His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr Met Glu	
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Leu Val Pro Ala Ala	
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Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys	
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Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn	
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Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser	
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Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu	
115 120 125	
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn	
130 135 140	
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp	
145 150 155 160	
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu	
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Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr	
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Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg	
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Gly Ser Pro Glu Phe Ala Pro Thr Val Lys Ile Asp Ala Gly Met Val	
225 230 235 240	
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245 250 255	
Phe Leu Gly Val Pro Phe Ala Ala Ser Pro Thr Arg Phe Ala Pro Pro	
260 265 270	
Thr Arg Pro Val Pro Trp Ser Thr Pro Leu Gln Ala Thr Ala Tyr Gly	
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Pro Ala Cys Pro Gln Gln Phe Asn Tyr Pro Glu Glu Leu Arg Glu Ile	
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Val	Ile	Ala	Val	Thr	Ile	Asn	Tyr	Arg	Thr	Asn	Ile	Leu	Gly	Phe	Pro
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Gln	Arg	Phe	Ala	Leu	Asp	Trp	Val	Gln	Arg	Asn	Ile	Ala	Ala	Phe	Gly
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Gly	Asp	Pro	Arg	Lys	Val	Thr	Ile	Phe	Gly	Gln	Ser	Ala	Gly	Gly	Arg
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Leu	Met	Asn	Thr	Ile	Glu	Gln	Leu	Gly	Leu	Gly	Phe	Glu	Tyr	Thr	Leu
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Leu	Leu	Phe	Val	Leu	Gly	Glu	Asn	Asp	Thr	Gln	Ala	Tyr	Leu	Glu	Glu
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Pro	Ile	Gly	Ser	Pro	Gly	Ile	Gly	Ser	Pro	Gln	Asp	Gln	Ile	Ala	Ala
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Ile	Glu	Thr	Glu	Val	Arg	Phe	Gln	Cys	Pro	Ser	Ala	Ile	Val	Ala	Gln
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Asp	Ser	Arg	Asn	Arg	Gly	Ile	Pro	Ser	Trp	Arg	Tyr	Tyr	Tyr	Asn	Ala
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Thr	Phe	Glu	Asn	Leu	Glu	Leu	Phe	Pro	Gly	Ser	Glu	Val	Tyr	His	Ser
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Ser	Glu	Val	Gly	Met	Val	Phe	Gly	Thr	Tyr	Pro	Val	Ala	Ser	Ala	Thr
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Ala	Leu	Glu	Ala	Gln	Thr	Ser	Lys	Tyr	Met	Gln	Gly	Ala	Trp	Ala	Ala
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Phe	Ala	Lys	Asn	Pro	Met	Asn	Gly	Pro	Gly	Trp	Lys	Gln	Val	Pro	Asn
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Val															

[illegible]

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bacterial expression vector pGEX-4T-1 or similar
vector. gst:BST1:sp:K:trAPAO fusion, 3591 nt.
1-687 gst
+ polylinker, 688-2163, BEST1 mature; 2164-2199,
spacer, 2200-3588, K:trAPAO
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 act cga ctt ctt ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ttg 96
 Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu
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 tat gag cgc gat gaa ggt gat aaa tgg cga aac aaa aag ttt gaa ttg 144
 Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu
 35 40 45
 ggt ttg gag ttt ccc aat ctt cct tat tat att gat ggt gat gtt aaa 192
 Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys
 50 55 60
 tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac 240
 Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn
 65 70 75 80
 atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa 288
 Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu
 85 90 95
 gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt 336
 Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser
 100 105 110
 aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa 384
 Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu
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 130 135 140
 ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat 480

53

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Gly Glu Ser Ala Gly Ala Ser Ala Ile Gly Leu Leu Leu Thr Ser Pro	
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Leu Ser Lys Gly Leu Phe Arg Gly Ala Ile Leu Glu Ser Pro Gly Leu	
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Thr Arg Pro Leu Ala Thr Leu Ala Asp Ser Ala Ala Ser Gly Glu Arg	
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Leu Asp Ala Asp Leu Ser Arg Leu Arg Ser Thr Asp Pro Ala Thr Leu	
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Met Ala Arg Ala Asp Ala Ala Arg Pro Ala Ser Arg Asp Leu Arg Arg	
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Pro Arg Pro Thr Gly Pro Ile Val Asp Gly His Val Leu Pro Gln Thr	
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Asp Ser Ala Ala Ile Ala Ala Gly Gln Leu Ala Pro Val Arg Val Leu	
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Ile Gly Thr Asn Ala Asp Glu Gly Arg Ala Phe Leu Gly Arg Ala Pro	
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Met Glu Thr Pro Ala Asp Tyr Gln Ala Tyr Leu Glu Ala Gln Phe Gly	
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Asp Gln Ala Ala Ala Val Ala Ala Cys Tyr Pro Leu Asp Gly Arg Ala	
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Thr Pro Lys Glu Met Val Ala Arg Ile Phe Gly Asp Asn Gln Phe Asn	
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Ala Thr His Gly Ala Glu Ile Pro Tyr Val Phe Gly Val Phe Lys Leu	
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Lys Asn Gly Asp Pro Ala Gly Asp Ala Leu Thr Trp Pro Ala Tyr Ser		
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Val Ala Asp Val Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr		
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Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn		
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Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr		
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acc act aca gct cct tat ggt gac tcc ttg ctg agc gag gag gtt gca		2544
Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala		
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Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu		
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His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp		
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Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala		
885	890	895

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gcc Ala	cac His	gag Glu 915	atc Ile	agc Ser	atg Met	ctt Leu	ttt Phe 920	ctc Leu	acc Thr	gac Asp	tac Tyr	atc Ile 925	aag Lys	agt Ser	gcc Ala	2784
acc Thr	ggg Gly 930	ctc Leu	agt Ser	aat Asn	att Ile	ttc Phe 935	tcg Ser	gac Asp	aag Lys	aaa Lys	gac Asp 940	ggc Gly	ggg Gly	cag Gln	tat Tyr	2832
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acc Thr	ttg Leu 1010	aca Thr	ttt Phe	tca Ser	cca Pro	cct Pro 1015	ctt Leu	ccc Pro	gcc Ala	gag Glu	aag Lys 1020	caa Gln	gca Ala	ttg Leu	gcg Ala	3072
gaa Glu 1025	aat Asn	tct Ser	atc Ile	ctg Leu	ggc Gly 1030	tac Tyr	tat Tyr	agc Ser	aag Lys	ata Ile 1035	gtc Val	ttc Phe	gta Val	tgg Trp	gac Asp 1040	3120
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caa Gln	tgg Trp	tcc Ser 1075	att Ile	acc Thr	tgt Cys	ttc Phe	atg Met 1080	gtc Val	gga Gly	gac Asp	ccg Pro	gga Gly 1085	cgg Arg	aag Lys	tgg Trp	3264
tcc Ser	caa Gln 1090	cag Gln	tcc Ser	aag Lys	cag Gln	gta Val 1095	cga Arg	caa Gln	aag Lys	tct Ser	gtc Val 1100	tgg Trp	gac Asp	caa Gln	ctc Leu	3312
cgc Arg 1105	gca Ala	gcc Ala	tac Tyr	gag Glu	aac Asn 1110	gcc Ala	ggg Gly	gcc Ala	caa Gln	gtc Val 1115	cca Pro	gag Glu	ccg Pro	gcc Ala	aac Asn 1120	3360
gtg Val	ctc Leu	gaa Glu	atc Ile	gag Glu 1125	tgg Trp	tcg Ser	aag Lys	cag Gln	cag Gln	tat Tyr 1130	ttc Phe	caa Gln	gga Gly	gct Ala 1135	ccg Pro	3408
agc	gcc	gtc	tat	ggg	ctg	aac	gat	ctc	atc	aca	ctg	ggg	tcg	gcg	ctc	3456

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Val	Trp	Lys	Gly	Tyr	Met	Glu	Gly	Ala	Ile	Arg	Ser	Gly	Gln	Arg	Gly		
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Tyr	Glu	Arg	Asp	Glu	Gly	Asp	Lys	Trp	Arg	Asn	Lys	Lys	Phe	Glu	Leu		
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Val	Cys	Phe	Lys	Lys	Arg	Ile	Glu	Ala	Ile	Pro	Gln	Ile	Asp	Lys	Tyr		
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Leu	Lys	Ser	Ser	Lys	Tyr	Ile	Ala	Trp	Pro	Leu	Gln	Gly	Trp	Gln	Ala		
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Gln	Val	Gln	Gly	Leu	Ala	Gly	Asp	Val	Met	Ser	Phe	Arg	Gly	Ile	Pro		
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Cys	Phe	Gly	Ala	Ala	Tyr	Leu	Arg	Lys	Gly	Ser	Leu	Ala	Pro	Gly	Val		
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Ser	Glu	Asp	Cys	Leu	Tyr	Leu	Asn	Val	Trp	Ala	Pro	Ser	Gly	Ala	Lys		
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Pro Gly Gln Tyr Pro Val Met Val Trp Val Tyr Gly Gly Gly Phe Ala
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 Thr Gly Lys Ser Thr Met Thr Phe Gly Pro Glu Gly Arg Ala Ala Val
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 Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu
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Glu	Leu	Val	Pro	Gly	Ser	Val	His	Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	
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Glu	Gln	Ser	Ala	Ser	Gly	Cys	Thr	Val	Arg	Ser	Ala	Ser	Gly	Ala	Val	
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Phe	Arg	Ser	Lys	Lys	Val	Val	Val	Ser	Leu	Pro	Thr	Thr	Leu	Tyr	Pro	
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Lys	Pro	Trp	Trp	Arg	Glu	Gln	Gly	Phe	Ser	Gly	Val	Leu	Gln	Ser	Ser	
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Ser	Gln	Gln	Ser	Lys	Gln	Val	Arg	Gln	Lys	Ser	Val	Trp	Asp	Gln	Leu	
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Variable	Mean	Standard Deviation	Minimum	Maximum
Age	34.1	7.1	21	49
Gender	0.48	0.50	0	1
Marital Status	0.48	0.50	0	1
Education	12.5	1.5	9	16
Income	12.5	1.5	9	16
Occupation	12.5	1.5	9	16
Health	12.5	1.5	9	16
Stress	12.5	1.5	9	16
Depression	12.5	1.5	9	16
Life Satisfaction	12.5	1.5	9	16
Resilience	12.5	1.5	9	16
Optimism	12.5	1.5	9	16
Self-Esteem	12.5	1.5	9	16
Emotional Stability	12.5	1.5	9	16
Neuroticism	12.5	1.5	9	16
Extraversion	12.5	1.5	9	16
Conscientiousness	12.5	1.5	9	16
Agreeableness	12.5	1.5	9	16
Openness	12.5	1.5	9	16
Intelligence	12.5	1.5	9	16
Personality	12.5	1.5	9	16
Behavior	12.5	1.5	9	16
Attitudes	12.5	1.5	9	16
Values	12.5	1.5	9	16
Beliefs	12.5	1.5	9	16
Opinions	12.5	1.5	9	16
Preferences	12.5	1.5	9	16
Interests	12.5	1.5	9	16
Hobbies	12.5	1.5	9	16
Skills	12.5	1.5	9	16
Abilities	12.5	1.5	9	16
Talents	12.5	1.5	9	16
Strengths	12.5	1.5	9	16
Weaknesses	12.5	1.5	9	16
Flaws	12.5	1.5	9	16
Defects	12.5	1.5	9	16
Shortcomings	12.5	1.5	9	16
Limitations	12.5	1.5	9	16
Constraints	12.5	1.5	9	16
Barriers	12.5	1.5	9	16
Obstacles	12.5	1.5	9	16
Challenges	12.5	1.5	9	16
Problems	12.5	1.5	9	16
Difficulties	12.5	1.5	9	16
Struggles	12.5	1.5	9	16
Trials	12.5	1.5	9	16
Tests	12.5	1.5	9	16
Experiences	12.5	1.5	9	16
Events	12.5	1.5	9	16
Situations	12.5	1.5	9	16
Circumstances	12.5	1.5	9	16
Conditions	12.5	1.5	9	16
States	12.5	1.5	9	16
Phases	12.5	1.5	9	16
Stages	12.5	1.5	9	16
Steps	12.5	1.5	9	16
Processes	12.5	1.5	9	16
Methods	12.5	1.5	9	16
Techniques	12.5	1.5	9	16
Approaches	12.5	1.5	9	16
Strategies	12.5	1.5	9	16
Tactics	12.5	1.5	9	16
Plans	12.5	1.5	9	16
Programs	12.5	1.5	9	16
Systems	12.5	1.5	9	16
Frameworks	12.5	1.5	9	16
Models	12.5	1.5	9	16
Theories	12.5	1.5	9	16
Hypotheses	12.5	1.5	9	16
Concepts	12.5	1.5	9	16
Ideas	12.5	1.5	9	16
Thoughts	12.5	1.5	9	16
Feelings	12.5	1.5	9	16
Emotions	12.5	1.5	9	16
Moods	12.5	1.5	9	16
Attitudes	12.5	1.5	9	16
Beliefs	12.5	1.5	9	16
Opinions	12.5	1.5	9	16
Preferences	12.5	1.5	9	16
Interests	12.5	1.5	9	16
Hobbies	12.5	1.5	9	16
Skills	12.5	1.5	9	16
Abilities	12.5	1.5</		

245										250					255					
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Glu	Leu	Leu	Pro	Val	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Gln					
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gac	ctc	aag	gcg	agc	cct	cag	gcg	aag	cgg	ctc	gac	agt	gtg	agc	ttc	864				
Asp	Leu	Lys	Ala	Ser	Pro	Gln	Ala	Lys	Arg	Leu	Asp	Ser	Val	Ser	Phe					
		275					280					285								
gcg	cac	tac	tgt	gag	aag	gaa	cta	aac	ttg	cct	gct	gtt	ctc	ggc	gta	912				
Ala	His	Tyr	Cys	Glu	Lys	Glu	Leu	Asn	Leu	Pro	Ala	Val	Leu	Gly	Val					
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gca	aac	cag	atc	aca	cgc	gct	ctg	ctc	ggg	gtg	gaa	gcc	cac	gag	atc	960				
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Ser	Met	Leu	Phe	Leu	Thr	Asp	Tyr	Ile	Lys	Ser	Ala	Thr	Gly	Leu	Ser					
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aat	att	ttc	tcg	gac	aag	aaa	gac	ggc	ggg	cag	tat	atg	cga	tgc	aaa	1056				
Asn	Ile	Phe	Ser	Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys					
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aca	ggg	atg	cag	tcg	att	tgc	cat	gcc	atg	tca	aag	gaa	ctt	gtt	cca	1104				
Thr	Gly	Met	Gln	Ser	Ile	Cys	His	Ala	Met	Ser	Lys	Glu	Leu	Val	Pro					
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Gly	Ser	Val	His	Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	Glu	Gln	Ser	Ala					
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tcc	ggc	tgt	aca	gta	cga	tcg	gcc	tcg	ggc	gcc	gtg	ttc	cga	agc	aaa	1200				
Ser	Gly	Cys	Thr	Val	Arg	Ser	Ala	Ser	Gly	Ala	Val	Phe	Arg	Ser	Lys					
385					390				395						400					
aag	gtg	gtg	gtt	tcg	tta	ccg	aca	acc	ttg	tat	ccc	acc	ttg	aca	ttt	1248				
Lys	Val	Val	Val	Ser	Leu	Pro	Thr	Thr	Leu	Tyr	Pro	Thr	Leu	Thr	Phe					
				405					410					415						
tca	cca	cct	ctt	ccc	gcc	gag	aag	caa	gca	ttg	gcg	gaa	aat	tct	atc	1296				
Ser	Pro	Pro	Leu	Pro	Ala	Glu	Lys	Gln	Ala	Leu	Ala	Glu	Asn	Ser	Ile					
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ctg	ggc	tac	tat	agc	aag	ata	gtc	ttc	gta	tgg	gac	aag	ccg	tgg	tgg	1344				
Leu	Gly	Tyr	Tyr	Ser	Lys	Ile	Val	Phe	Val	Trp	Asp	Lys	Pro	Trp	Trp					
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cgc	gaa	caa	ggc	ttc	tcg	ggc	gtc	ctc	caa	tcg	agc	tgt	gac	ccc	atc	1392				
Arg	Glu	Gln	Gly	Phe	Ser	Gly	Val	Leu	Gln	Ser	Ser	Cys	Asp	Pro	Ile					
	450					455					460									
tca	ttt	gcc	aga	gat	acc	agc	atc	gac	gtc	gat	cga	caa	tgg	tcc	att	1440				
Ser	Phe	Ala	Arg	Asp	Thr	Ser	Ile	Asp	Val	Asp	Arg	Gln	Trp	Ser	Ile					
465					470				475						480					
acc	tgt	ttc	atg	gtc	gga	gac	ccg	gga	cgg	aag	tgg	tcc	caa	cag	tcc	1488				
Thr	Cys	Phe	Met	Val	Gly	Asp	Pro	Gly	Arg	Lys	Trp	Ser	Gln	Gln	Ser					
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Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr	
500 505 510	
gag aac gcc ggg gcc caa gtc cca gag ccg gcc aac gtg ctc gaa atc	1584
Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile	
515 520 525	
gag tgg tcg aag cag cag tat ttc caa gga gct ccg agc gcc gtc tat	1632
Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr	
530 535 540	
ggg ctg aac gat ctc atc aca ctg ggt tcg gcg ctc aga acg ccg ttc	1680
Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe	
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Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly	
565 570 575	
tat atg gaa ggg gcc ata cga tcg ggt caa cga ggt gct gca gaa gtt	1776
Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val	
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<210> 33
 <211> 600
 <212> PRT
 <213> Unknown

<400> 33

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35 40 45	
Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys	
50 55 60	
Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr	
65 70 75 80	
Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly	
85 90 95	
Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val	
100 105 110	
Pro Val Ser Ala Leu Ser Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala	
115 120 125	
Thr Ala Leu Val Pro Gly His Thr Thr Pro Asp Asn Val Ala Asp Val	
130 135 140	
Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val	
145 150 155 160	
Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val	
165 170 175	
Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile	
180 185 190	
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<211> 1929
 <212> DNA
 <213> *Exophiala spinifera*

<220>
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 <222> (739)...(811)

<221> intron
 <222> (1134)...(1186)

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gacgcttttg	gcgtgacaga	cccagcctac	gagaaacagg	ttgcccagc	attcgccaat	180
ctgcgagctt	gccttgctgc	agttggagcc	tcttcaaacg	acgtcaccaa	gctcaattac	240
tacatcgctg	actacgcccc	gagcaaaactc	accgcaattg	gagatgggct	gaagtctacc	300
tttgcccttg	acaggtctccc	tccttgcaag	ctgggtgccag	taccggcctt	ggcttcacct	360
gaatacctct	ttgaggttga	tgccacggcg	ctgggtgccag	gacactcgac	cccagacaac	420
gttgcggaag	tggttagtgg	gggcgctggc	ttgagcgggt	tgagagcggc	acgcaaagtc	480
caggccgcgc	gtctgtcctg	cctcgcttct	gaggcgatgg	atcgtgtagg	gggaaagact	540
ctgagcgtac	aatcgggtcc	cggcaggacg	actatcaacg	acctcggcgc	tgctgggatc	600
aatgacagca	accaaagcga	agtatccaga	ttgtttgaaa	gatttcattt	ggagggcgag	660
ctccagagga	cgaccggaaa	ttcaatccat	caagcacaag	acggtacaac	cactacagct	720
ccttatgggtg	actccccggg	aagcacaatc	ccactttgtg	atgagacctc	tgctgagtg	780
agaatacagt	cactgactcc	acttcgtcca	gctgagcgag	gaggttgcaa	gtgcaactgc	840
ggaactcctc	cccgtatgg	ctcagctgat	cgaagagtat	agccttgaag	acccaaggc	900
gagccctcag	gcgaagcggc	tcgacagtgt	gagcttcgcg	cactactgtg	agaaggacct	960
aaacttgcc	gctgttctca	gcgtggcaaa	ccagatcaca	cgcgctctgc	tcggtgtgga	1020
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cgggtgtctc	tcaggtaggg	gactcgtttc	ttagtgggtc	ttccagggtat	gcagtcgatt	1200
tgccatgcca	tgtcaaagga	acttgttcca	ggctcagtg	acctcaacac	ccccgtcgct	1260
ggaattgagc	agtcggcgct	cggctgtata	gtacgatcgg	cctcggggcg	cgtgttcoga	1320
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cctcttcccc	ccgagaagca	agcattggcg	gaaaaatcta	tcctcggcta	ctatagcaag	1440
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<210> 36
 <211> 600
 <212> PRT
 <213> *Exophiala spinifera*

<400> 36

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Thr	Ile	Ala	Gly	Gln	Ile	Gly	Gln	Asp	Ala	Leu	Gly	Val	Thr	Asp	Pro
Ala	Tyr	Glu	Lys	Gln	Val	Ala	Gln	Ala	Phe	Ala	Asn	Leu	Arg	Ala	Cys
Leu	Ala	Ala	Val	Gly	Ala	Ser	Ser	Asn	Asp	Val	Thr	Lys	Leu	Asn	Tyr
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Val Ala Ser Leu Val Pro Ala Ala
595 600

<210> 37
<211> 1929
<212> DNA
<213> *Exophiala spinifera*

<220>
<221> intron
<222> (739)...(811)

<221> intron
<222> (1134)...(1186)

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ctgcgagctt gccttgctgc agttggagcc tcttcaa acg acgtcaccaa gctcaattac 240
tacatcgtcg actacgcccc gagcaaaactc accgcaattg gagatgggct gaagtctacc 300
tttgcccttg acaggtctccc tccttgcaag ctggtgccag taccggcctt ggcttcacct 360
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gttgcggaag tggtagtggt gggcgctggc ttgagcggct tggagacggc acgcaaagtc 480
caggccgcgc gtctgtcctg cctcgttctt gaggcgatgg atcgtgtagg gggaaagact 540
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aatgacagca accaaagcga agtatccaga ttgtttgaaa gatttcattt ggagggcgag 660
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ccttatggtg actccccggg aagcacaatc ccactttgtg atgagacctc tgcgagtgt 780
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<210> 38
<211> 600
<212> PRT
<213> *Exophiala spinifera*

<400> 38
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Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe
545 550 555 560
Lys Cys Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly
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Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val
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Val Ala Ser Leu Val Pro Ala Ala
595 600

<210> 39
<211> 1930
<212> DNA
<213> *Exophiala spinifera*

<220>
<221> intron
<222> (739)...(811)

<221> intron
<222> (1134)...(1187)

<221> misc_feature
<222> (648)...(648)
<223> n = A,T,C or G

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ctgcgagctt gccttgctgc agttggagcc acttcaaacg acgtcaccaa gctcaattac 240
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ctccagagga cgactggaaa ttcaatccat caagcacaag acggtacaac cactacagct 720
ccttatgggtg actccttggt aagcacaatc ccactttgtg atgagacctc tgtcgagtgt 780
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gcogttcaag agtggttcatt tcgttggaac ggagacgtct ttagtttgga aagggtatat 1860
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agcagcatag 1930

<210> 40
 <211> 598
 <212> PRT
 <213> Exophiala spinifera

 <220>
 <221> VARIANT
 <222> (216)...(216)
 <223> Xaa = Any Amino Acid

<400> 40
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 1 5 10 15
 Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Gly Arg Tyr Val
 20 25 30
 Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro
 35 40 45
 Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys
 50 55 60
 Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr
 65 70 75 80
 Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly
 85 90 95
 Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val
 100 105 110
 Pro Val Ser Ala Leu Ser Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala
 115 120 125
 Thr Ala Leu Val Pro Gly His Thr Thr Pro Asp Asn Val Ala Asp Val
 130 135 140
 Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val Gln
 145 150 155 160
 Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val Gly
 165 170 175
 Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile Asn
 180 185 190
 Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Ser
 195 200 205
 Arg Leu Phe Glu Arg Phe His Xaa Glu Gly Glu Leu Gln Arg Thr Thr
 210 215 220
 Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro
 225 230 235 240
 Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu
 245 250 255
 Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln Asp
 260 265 270
 Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe Ala
 275 280 285
 His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val Asn
 290 295 300
 Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met
 305 310 315 320
 Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn Ile
 325 330 335
 Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr Gly
 340 345 350
 Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro Gly Ser
 355 360 365
 Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser Gly
 370 375 380
 Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys Lys Val
 385 390 395 400
 Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe Ser Pro


```

cttttccgc cgagaagcaa gcattggctg aaaaatccat cctgggctac tatagcaaga 1440
tagtcttcgt atgggacaag ccgtggtggc gcgaacaagg cttctcgggc gtctccaat 1500
cgagctgtga ccccatctca tttgccagag ataccagcat cgaagtcgat cggcaatggt 1560
ccattacctg tttcatggtc ggagaccggg gacggaagtg gtcccaacag tccaagcagg 1620
tacgacagaa gtctgtctgg aaccaactcc gcgcagccta cgagaacgcc ggggcccaag 1680
tcccagagcc ggccaacgtg ctcgagatcg agtggtcgaa gcagcagtat ttccaaggag 1740
cgccgagcgt cgtctatggg ctgaactgtc tcaacacact gggttcggcg ctcaaacgc 1800
cgttcaaggg tgttcatttc gttggaacgg agacgtcttt ggtttgaaa gggatatatgg 1860
aaggggccat acgatcgggt cagcgaggcg ctgcagaagt tgtggctagc ctggtgccag 1920
cagcatag 1928

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<210> 42
<211> 598
<212> PRT
<213> Rhinocycladiella atrovirens

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<400> 42
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Ala Gly Tyr Ser His Val Gly Val Gly Pro Asn Gly Gly Arg Tyr Ala
20 25 30
Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Ala Val Thr Asp Pro
35 40 45
Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys
50 55 60
Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Ile Thr Lys Leu Asn Tyr
65 70 75 80
Tyr Ile Val Asp Tyr Asn Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly
85 90 95
Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val
100 105 110
Pro Val Pro Ala Leu Ala Ser Pro Glu Tyr Pro Phe Glu Val Asp Ala
115 120 125
Thr Ala Leu Val Pro Gly His Ser Thr Pro Asp Asn Val Ala Asp Val
130 135 140
Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val
145 150 155 160
Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val
165 170 175
Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Ala Ile
180 185 190
Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val
195 200 205
Phe Lys Leu Phe Glu Arg Leu Glu Gly Glu Leu Gln Arg Thr Thr Gly
210 215 220
Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro Tyr
225 230 235 240
Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu Leu
245 250 255
Leu Pro Ala Trp Ser Gln Leu Ile Glu Glu His Ser Leu Glu Asp Pro
260 265 270
Lys Ala Ser Pro Gln Ala Lys Gln Leu Asp Ser Val Ser Phe Ala His
275 280 285
Tyr Cys Glu Lys Asp Leu Ser Leu Pro Ala Val Leu Gly Val Ala Asn
290 295 300
Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met
305 310 315 320
Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn Ile
325 330 335
Val Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr Gly
340 345 350
Met Gln Ser Leu Cys His Ala Met Ser Lys Glu Leu Val Pro Gly Ser

```

[illegible]

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<210> 43
<211> 1928
<212> DNA
<213> Rhinoclediella atrovirens
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```
<221> intron
<222> (1134)...(1186)
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gacgcttcgg	ccgtgacaga	ccctgcctac	gagaaacagg	ttgcccaagc	attcgccaac	180
ctgcgagctt	gtcttgctgc	agttggagcc	acttcaaacy	acattaccaa	gctcaattac	240
tacatcgctg	actacaaccc	gagcaaactc	accgcaattg	gagatgggct	gaaggctacc	300
tttgcccttg	acaggctccc	tccttgcacg	ctggtgccag	tgccggccct	ggcttcacct	360
gaatacctct	ttgaggttga	tgccacggcg	ctggttcag	gacactcaac	cccagacaat	420
gttcgggacg	tggctgtggg	gggcgctggc	ttgagcggtt	tggagacggc	acgcaaagtc	480
caggctgcgc	ggctgtcctg	cctcgttcct	gaggcgatgg	atcgtgtggg	gggaaagact	540
ctgagcgta	aatcgggtcc	cggcaggacg	actatcaatg	acctcgcgcg	tgcgtggatc	600
aatgacagca	accaaagcga	agtattcaaa	ttatttgaaa	gatttcattt	ggagggcgag	660
ctccagagga	cgaccggaaa	ttcaatccat	caagcacaa	acggtacaac	cactacagct	720
ccttatggtg	attccctggt	aagcacaatt	ccatcttgtg	atgagacctc	tgtcgtgtgt	780
agaatacagt	cgtgactccc	acatcgtcca	gctgagcgag	gaggttgcaa	gtgcactcgc	840
ggaactcctt	cccgactggg	ctcagctgat	cgaagagcat	agtcttgaag	accccaaggc	900
gagcctcaa	gcgaagcagc	tcgacagtgt	gagcttcgca	cactactgtg	agaaggatct	960
aaacttgcc	gctgttctcg	gcgtggcaaa	ccagatcaca	cgcgctctgc	tcggtgtgga	1020

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agcccacgag atcagcatgt tttttctcac cgactacatc aagagtgcc a cgggtctcag 1080
taatatgtgc tccgataaga aagacgggtg gcagtatatg cgatgcaaaa caggtgcgtg 1140
tggtgttctc tcagtgggag actcgtttct tagtgggtcat tccaggtatg cagtcgcttt 1200
gccatgccat gtcaaaggaa cttgttccag gctcagtga cctcaacacc cccgtcgccg 1260
aaattgagca gtcggcatcc ggctgtacag tacgatcggc ctcgggcggc gtgttccgaa 1320
gtaaaaaggt ggtggtttcg ttaccgacaa ccttgtatcc caccttgata ttttcaccac 1380
ctcttcccg cagagaagcaa gcattggctg aaaaatccat cctggggtac tatagcaaga 1440
tagtcttcgt atgggacaag ccgtgggtggc gcgaacaagg cttctcgggc gtctccaat 1500
cgagctgtga ccccatctca tttgccagag ataccagcat cgaagtcgat cggcaatggt 1560
ccattacctg tttcatggtc ggagaccgg gacggaagtg gtccaacag tccaagcagg 1620
tacgacagaa gtctgtctgg aaccaactcc ggcagccta cgagaacgcc ggggcccaag 1680
tcccagagcc ggccaactg ctcgagatcg agtggtcgaa gcagcagtat ttccaaggag 1740
cgccgagcgc cgtctatggg ctgaactgtc tcaacacact gggttcggcg ctcagaacgc 1800
cgttcaaggg tgttcatttc gttggaacgg agacgtcttt ggtttgaaa gggatatagg 1860
aaggggccat acgatcgggt cagcgaggcg ctgcagaagt tgtggctagc ctggtgccag 1920
cagcatag 1928

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<210> 44
<211> 591
<212> PRT
<213> Rhinocycladiella atrovirens

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<400> 44
Met Ala Leu Ala Pro Ser Tyr Ile Asn Pro Pro Asn Leu Ala Ser Pro
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Ala Gly Tyr Ser Tyr Val Gly Val Gly Pro Asn Gly Gly Arg Tyr Val
20 25 30
Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Ala Val Thr Asp Pro
35 40 45
Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys
50 55 60
Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Ile Thr Lys Leu Asn Tyr
65 70 75 80
Tyr Ile Val Asp Tyr Asn Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly
85 90 95
Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val
100 105 110
Pro Val Pro Ala Leu Ala Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala
115 120 125
Thr Ala Leu Val Pro Gly His Ser Thr Pro Asp Asn Val Ala Asp Val
130 135 140
Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val
145 150 155 160
Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val
165 170 175
Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Gly Arg Thr Thr Ile Asn
180 185 190
Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Lys
195 200 205
Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr Thr Gly
210 215 220
Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Thr Ala Pro Tyr
225 230 235 240
Gly Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu Leu Leu
245 250 255
Pro Ala Ser Gln Leu Ile Glu Glu His Ser Leu Glu Asp Pro Lys Ala
260 265 270
Ser Pro Gln Ala Lys Gln Leu Asp Ser Val Ser Phe Ala His Tyr Cys
275 280 285
Glu Lys Leu Asn Leu Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg
290 295 300
Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met Phe Phe Leu Thr

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[illegible]

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<220>
<221> intron
<222> (739)...(811)

<221> intron
<222> (1134)...(1185)
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74

[illegible]

<400> 47

76

Thr	Ala	Leu	Val	Pro	Gly	His	Thr	Thr	Pro	Asp	Asn	Val	Ala	Asp	Val
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Val	Val	Val	Gly	Ala	Gly	Leu	Ser	Gly	Leu	Glu	Thr	Ala	Arg	Lys	Val
145						150				155					160
Gln	Ala	Ala	Gly	Leu	Ser	Cys	Leu	Val	Leu	Glu	Ala	Met	Asp	Arg	Val
						165				170					175
Gly	Gly	Lys	Thr	Leu	Ser	Val	Gln	Ser	Gly	Pro	Gly	Arg	Thr	Thr	Ile
						180				185					190
Asn	Asp	Leu	Gly	Ala	Ala	Trp	Ile	Asn	Asp	Ser	Asn	Gln	Ser	Glu	Val
						195						205			
Ser	Arg	Leu	Phe	Glu	Arg	Phe	His	Leu	Glu	Gly	Glu	Leu	Gln	Arg	Thr
						215						220			
Thr	Gly	Asn	Ser	Ile	His	Gln	Ala	Gln	Asp	Gly	Thr	Thr	Thr	Thr	Ala
225						230						235			240
Pro	Tyr	Gly	Asp	Ser	Leu	Leu	Ser	Glu	Glu	Val	Ala	Ser	Ala	Leu	Ala
						245				250					255
Glu	Leu	Leu	Pro	Val	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Gln
						260				265					270
Asp	Leu	Lys	Ala	Ser	Pro	Gln	Ala	Lys	Arg	Leu	Asp	Ser	Val	Ser	Phe
						275							285		
Ala	His	Tyr	Cys	Glu	Lys	Glu	Leu	Asn	Leu	Pro	Ala	Val	Leu	Gly	Val
						295						300			
Ala	Asn	Gln	Ile	Thr	Arg	Ala	Leu	Leu	Gly	Val	Glu	Ala	His	Glu	Ile
305						310						315			320
Ser	Met	Leu	Phe	Leu	Thr	Asp	Tyr	Ile	Lys	Ser	Ala	Thr	Gly	Leu	Ser
						325				330					335
Asn	Ile	Phe	Ser	Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys
						340				345				350	
Thr	Gly	Met	Gln	Ser	Ile	Cys	His	Ala	Met	Ser	Lys	Glu	Leu	Val	Pro
						355						365			
Gly	Ser	Val	His	Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	Glu	Gln	Ser	Ala
						375						380			
Ser	Gly	Cys	Thr	Val	Arg	Ser	Ala	Ser	Gly	Ala	Val	Phe	Arg	Ser	Lys
385						390					395				400
Lys	Val	Val	Val	Ser	Leu	Pro	Thr	Thr	Leu	Tyr	Pro	Thr	Leu	Thr	Phe
						405				410					415
Ser	Pro	Pro	Leu	Pro	Ala	Glu	Lys	Gln	Ala	Leu	Ala	Glu	Asn	Ser	Ile
						420				425				430	
Leu	Gly	Tyr	Tyr	Ser	Lys	Ile	Val	Phe	Val	Trp	Asp	Lys	Pro	Trp	Trp
						435				440				445	
Arg	Glu	Gln	Gly	Phe	Ser	Gly	Val	Leu	Gln	Ser	Ser	Cys	Asp	Pro	Ile
						455						460			
Ser	Phe	Ala	Arg	Asp	Thr	Ser	Ile	Asp	Val	Asp	Arg	Gln	Trp	Ser	Ile
465						470					475				480
Thr	Cys	Phe	Met	Val	Gly	As									

<211> 1392
 <212> DNA
 <213> Unknown

<220>
 <221> CDS
 <222> (1)...(1392)

<223> Cys (-) APAO; removal of cysteine 461

<400> 48

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ttg gag acg gca cgc aaa gtc cag gcc gcc ggt ctg tcc tgc ctc gtt	96
Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val	
20 25 30	
ctt gag gcg atg gat cgt gta ggg gga aag act ctg agc gta caa tcg	144
Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser	
35 40 45	
ggg ccc ggc agg acg act atc aac gac ctc ggc gct gcg tgg atc aat	192
Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn	
50 55 60	
gac agc aac caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg	240
Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu	
65 70 75 80	
gag ggc gag ctc cag agg acg act gga aat tca atc cat caa gca caa	288
Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln	
85 90 95	
gac ggt aca acc act aca gct cct tat ggt gac tcc ttg ctg agc gag	336
Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu	
100 105 110	
gag gtt gca agt gca ctt gcg gaa ctc ctc ccc gta tgg tct cag ctg	384
Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu	
115 120 125	
atc gaa gag cat agc ctt caa gac ctc aag gcg agc cct cag gcg aag	432
Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys	
130 135 140	
cgg ctc gac agt gtg agc ttc gcg cac tac tgt gag aag gaa cta aac	480
Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn	
145 150 155 160	
ttg cct gct gtt ctc ggc gta gca aac cag atc aca cgc gct ctg ctc	528
Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu	
165 170 175	
ggg gtg gaa gcc cac gag atc agc atg ctt ttt ctc acc gac tac atc	576
Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile	
180 185 190	
aag agt gcc acc ggt ctc agt aat att ttc tcg gac aag aaa gac ggc	624
Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly	
195 200 205	

ggg Gly	cag Gln	tat Tyr	atg Met	cga Arg	tgc Cys	aaa Lys	aca Thr	ggt Gly	atg Met	cag Gln	tcg Ser	att Ile	tgc Cys	cat His	gcc Ala	672
210215220																
atg Met	tca Ser	aag Lys	gaa Glu	ctt Leu	gtt Val	cca Pro	ggc Gly	tca Ser	gtg Val	cac His	ctc Leu	aac Asn	acc Thr	ccc Pro	gtc Val	720
225230235240																
gct Ala	gaa Glu	att Ile	gag Glu	cag Gln	tcg Ser	gca Ala	tcc Ser	ggc Gly	tgt Cys	aca Thr	gta Val	cga Arg	tcg Ser	gcc Ala	tcg Ser	768
245250255																
ggc Gly	gcc Ala	gtg Val	ttc Phe	cga Arg	agc Ser	aaa Lys	aag Lys	gtg Val	gtg Val	gtt Val	tcg Ser	tta Leu	ccg Pro	aca Thr	acc Thr	816
260265270																
ttg Leu	tat Tyr	ccc Pro	acc Thr	ttg Leu	aca Thr	ttt Phe	tca Ser	cca Pro	cct Pro	ctt Leu	ccc Pro	gcc Ala	gag Glu	aag Lys	caa Gln	864
275280285																
gca Ala	ttg Leu	gcg Ala	gaa Glu	aat Asn	tct Ser	atc Ile	ctg Leu	ggc Gly	tac Tyr	tat Tyr	agc Ser	aag Lys	ata Ile	gtc Val	ttc Phe	912
290295300																
gta Val	tgg Trp	gac Asp	aag Lys	ccg Pro	tgg Trp	tgg Trp	cgc Arg	gaa Glu	caa Gln	ggc Gly	ttc Phe	tcg Ser	ggc Gly	gtc Val	ctc Leu	960
305310315320																
caa Gln	tcg Ser	agc Ser	tcc Ser	gac Asp	ccc Pro	atc Ile	tca Ser	ttt Phe	gcc Ala	aga Arg	gat Asp	acc Thr	agc Ser	atc Ile	gac Asp	1008
325330335																
gtc Val	gat Asp	cga Arg	caa Gln	tgg Trp	tcc Ser	att Ile	acc Thr	tgt Cys	ttc Phe	atg Met	gtc Val	gga Gly	gac Asp	ccg Pro	gga Gly	1056
340345350																
cgg Arg	aag Lys	tgg Trp	tcc Ser	caa Gln	cag Gln	tcc Ser	aag Lys	cag Gln	gta Val	cga Arg	caa Gln	aag Lys	tct Ser	gtc Val	tgg Trp	1104
355360365																
gac Asp	caa Gln	ctc Leu	cgc Arg	gca Ala	gcc Ala	tac Tyr	gag Glu	aac Asn	gcc Ala	ggg Gly	gcc Ala	caa Gln	gtc Val	cca Pro	gag Glu	1152
370375380																
ccg Pro	gcc Ala	aac Asn	gtg Val	ctc Leu	gaa Glu	atc Ile	gag Glu	tgg Trp	tcg Ser	aag Lys	cag Gln	cag Gln	tat Tyr	ttc Phe	caa Gln	1200
385390395400																
gga Gly	gct Ala	ccg Pro	agc Ser	gcc Ala	gtc Val	tat Tyr	ggg Gly	ctg Leu	aac Asn	gat Asp	ctc Leu	atc Ile	aca Thr	ctg Leu	ggc Gly	1248
405410415																
tcg Ser	gcg Ala	ctc Leu	aga Arg	acg Thr	ccg Pro	ttc Phe	aag Lys	agt Ser	gtt Val	cat His	ttc Phe	gtt Val	gga Gly	acg Thr	gag Glu	1296
420425430																
acg Thr	tct Ser	tta Leu	gtt Val	tgg Trp	aaa Lys	ggg Gly	tat Tyr	atg Met	gaa Glu	ggg Gly	gcc Ala	ata Ile	cga Arg	tcg Ser	ggc Gly	1344
435440445																
caa Gln	cga Gln	qgt Gln	gct Gln	qca Gln	gaa Gln	ggt Gln	gtg Gln	gct Gln	agc Gln	ctg Gln	gtg Gln	cca Gln	gca Gln	gca Gln	tag Gln	1392

Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu
 305 310 315 320
 Gln Ser Ser Ser Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp
 325 330 335
 Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly
 340 345 350
 Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp
 355 360 365
 Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu
 370 375 380
 Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln
 385 390 395 400
 Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly
 405 410 415
 Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu
 420 425 430
 Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly
 435 440 445
 Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala
 450 455 460

<210> 52

<211> 1392

<212> DNA

<213> Unknown

<220>

<221> CDS

<222> (1)...(1392)

<223> Cys (-) APAO; removal of cysteines 169, 359, and
 461

<400> 52

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Lys Asp Asn Val Ala Asp Val Val Val Val Gly Ala Gly Leu Ser Gly	
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ttg gag acg gca cgc aaa gtc cag gcc gcc ggt ctg agc tcc ctc gtt	96
Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Ser Leu Val	
20 25 30	
ctt gag gcg atg gat cgt gta ggg gga aag act ctg agc gta caa tcg	144
Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser	
35 40 45	
ggt ccc ggc agg acg act atc aac gac ctc ggc gct gcg tgg atc aat	192
Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn	
50 55 60	
gac agc aac caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg	240
Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu	
65 70 75 80	
gag ggc gag ctc cag agg acg act gga aat tca atc cat caa gca caa	288
Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln	
85 90 95	
gac ggt aca acc act aca gct cct tat ggt gac tcc ttg ctg agc gag	336
Asp Gly Thr Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu	
100 105 110	

